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# The Alysiinae (Hym. Braconidae) parasites of the Agromyzidae (Diptera)

VI. The parasites of Cerodontha RONDANI s. 1.<sup>1</sup>

With textfigures 186-209

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#### Introduction

Five parts of this paper have already appeared in this journal (GRIFFITHS, 1964, 1966a, 1966b, 1967 and 1968). The present sixth part deals with the parasites of *Cerodontha* RONDANI sensu lato, a monophyletic group whose larvae are associated exclusively with monocotyledonous host-plants. This genus has for some time been the subject of detailed study by Dr. J. T. NOWAKOWSKI of the Polish Academy of Sciences, and a preliminary summary of his conclusions has recently been published (NOWAKOWSKI, 1967). The nomenclature for host species used in this paper has been revised to accord largely with that proposed by NOWAKOWSKI. Until recently some of the subgenera now included in *Cerodontha* s.l. were considered as subgenera of "*Phytobia*", but NOWAKOWSKI (1962) clearly demonstrated that this concept was heterogeneous. The generic name *Phytobia* is now used in a restricted sense to include only those species whose larvae mine

<sup>1</sup> Part I in Beitr. Ent., 14, 823-914; 1964. – Part II in Beitr. Ent., 16, 551-605; 1966. – Part III in Beitr. Ent., 16, 775-951; 1966. – Part IV in Beitr. Ent., 17, 653-696; 1967. – Part V in Beitr. Ent., 18, 5-62; 1968.

in the cambium of trees (= Dendromyza HENDEL). This is now the only genus of Agromyzidae with known Alysiinae parasites which I have not yet treated in this series of papers. Whether suitable material will be available in the near future to enable me to remedy this omission is not yet clear.

I have been able to provide in this part revised keys to the outstanding groups of the large and difficult genus *Chorebus* HALIDAY, as previously promised.

Acknowledgements to the various persons who have helped me with material for this paper have already been given in previous parts. I also wish to express my special thanks to Dr. J. T. NOWAKOWSKI for his assistance in clarifying the identity and nomenclature of the host species.

The abbreviations used in this paper were explained in the introduction to Part II (GRIFFITHS, 1966a), except for the following addition: USNM — United States National Museum, Washington, D. C., U.S.A.

Notes on the bases of the measurements and ratios in the tables of biometric data have been given in Part I (GRIFFITHS, 1964, page 904).

### **Previous Records**

Table 24

As in previous parts of this paper I have prepared a table below explaining the discrepancies between my list of host records and the list given by FULMEK (1962): this table completes my reassessment of all records of Alysiinae parasites of Agromyzidae given in that work. The comments in the table exclude changes in the generic nomenclature which affect nearly all the names (both of hosts and parasites). A few old records which are in conflict with confirmed records must be rejected as not being based on any good authority, although I have not seen the material on which they were based.

Host	Parasite	Comments
Cerodonta sp.	Rhizarcha lestes NIXON	host was Scaptomyza graminum FALLÉN (Drosophilidae): see footnote 8 in Part III (GRIFFITHS, 1966b, page 893).
Phytobia incisa Meigen	Chorebus uliginosa HALIDAY	not accepted (parasite of <i>Hydrellia</i> , Ephy- dridae)
	Dacnusa sp.	record too imprecise to be of value
	Rhizarcha stramineipes THOMSON	not accepted (Dacnusa stramineipes
		(HALIDAY) has a long ovipositor and can
		hardly be a parasite of a leaf-miner).
Phytobia iraeos GOUREAU	Dacnusa raissa NIXON	accepted : the prior name Chorebus flavipes
		(GOUREAU) is used for the parasite in this
		paper.
	Rhizarcha maculipes THOMSON	host was C. (Dizygomyza) iridis HENDEL
Phytobia morosa Meigen	Alysia truncator NEES	not accepted (Alysia spp. are parasites of
	Ť	Calliphoridae)
Phytobia pygmaea Meigen	Dacnusa asramenes NIXON	accepted
	Rhizarcha stramineipes Thomson	not accepted (Dacnusa stramineipes
		(HALIDAY) has a long ovipositor and can
		hardly be a parasite of a leaf-miner).
Phytobia sönderupi HERING	Dacnusa morella NIXON	accepted (but spelling should be merella):
		the host name is considered synonymous
		with C. (Dizygomyza) caricicola HERING in
		this paper.
	Rhizarcha stramineipes HALIDAY	not accepted

Earlier Records of Alysiinae parasites of *Cerodontha* s.l. (after FULMEK, 1962) with comments thereon

# Dapsilarthra Förster

# Dapsilarthra sylvia (HALIDAY)

In addition to the records of this species from other host genera given in Parts II, III and V, I have seen the following specimens bred from *Cerodontha* hosts.

Cerodontha (Butomomyza) angulata LOEW -1  $\bigcirc$  from larva 30. ix. 64 on Carex silvatica, Clapham, Yorks., England, em. 16. iv. 65 (GCDG).

Cerodontha (Butomomyza) caricivora GROSCHKE — 1  $\bigcirc$  from larva 26. ix. 65 on Carex silvatica, Kiczera, Bieszczady, Poland, em. xii. 65, leg. NOWAKOWSKI (PAN). 1  $\bigcirc$  from larva 8. x. 65 on Carex silvatica, Hulskie, Otryt, Poland, em. 16. iii. 66, leg. NOWAKOWSKI (PAN).

Cerodontha (Poemyza) pygmaea MEIGEN  $-1 \circ$  from puparium 15. vii. 64 on Melica nutans, Żukowo, distr. Kartuzy, Poland, em. 15. viii. 64, leg. NOWAKOWSKI (PAN).

# Dapsilarthra balteata (THOMSON)

In addition to the records of this species from other host genera given in Parts II, III and V, I have seen the following material bred from *Cerodontha* subgenus *Poemyza*.

Cerodontha (Poemyza) pygmaea MEIGEN – 1 ex. from puparium 27. viii. 60 on Arrhenatherum elatius, Woodwalton Fen, Hunts., England, em. 9. ix. 60 (BM): 1  $\bigcirc$  from puparium 12. ix. 60 on Dactylis glomerata, same locality, em. 26. iii. 61 (GCDG). 1  $\bigcirc$  from puparium 14. viii. 35 on Festuca gigantea, Warsow, near Neukalen, Mecklenburg, Germany, em. 10. iii. 36, leg. BUHR (GCDG). 1  $\eth$  from puparium 21. vi. 64 on Dactylis glomerata, Warszawa-Pyry, Kabackie Forest, Poland, em. 2. ix. 64, leg. NOWAKOWSKI (PAN): 2  $\circlearrowright$  from puparia 17. vi. 64 on Deschampsia caespitosa, same locality, em. 16. vii and 30. viii. 64, leg. NOWAKOWSKI (PAN). 1  $\circlearrowright$  from puparium 11. vi. 64 on Dactylis glomerata, Warszawa-Młociny, em. 18. vii. 64, leg. NOWAKOWSKI (PAN). 2  $\eth$  from larvae and puparia 9. vii. 66 on Deschampsia caespitosa, Wierzba, near Popielno, Piska Forest, Poland, em. 28. vii. 66, leg. NOWAKOWSKI (PAN).

Cerodontha (Poemyza) incisa MEIGEN — 3 ex. from puparia 21. vi. 61 on Phalaris arundinacea, Woodwalton Fen, Hunts., England, em. 21. vii. 61 and 21. ix. 61 (2 ex.) (GCDG and BM). 1 ex. from puparium 31. viii. 54 on Phalaris arundinacea, Broxbourne, Essex, England, em. 30. x. 54 (BM). 1 3 from puparium 31. vii. 66 on Agropyron repens, Otwock Wielki, near Warszawa, Poland, em. 25. ix. 66, leg. Nowakowski (PAN). 1 3 from puparium 16. vi. 66 on Calamagrostis canescens, Sieraków reservation, Kampinoska Forest, Poland, em. 25. vii. 66, leg. Nowakowski (PAN).

The records for Woodwalton Fen have been previously published (GRIFFITHS, 1963). KÖNIGSMANN (1959) has already recorded *pygmaea* as a host.

### Dapsilarthra fuscula sp. nov.

This species is very similar to D. rufiventris (NEES), and may be compared with the description of that species in KÖNIGSMANN (1959) as follows.

Colour much darker. Palpi ochreous yellow. Labrum red-brown or almost black. Clypeus black. Antennae entirely dark. Legs ochreous yellow or yellowbrown, with the middle and hind coxae strongly infuscated, becoming virtually black towards their base. Gaster entirely dark.

29 and 31 antennal segments  $(2 \Im \Im)$ . Mesoscutum with some fairly dense pubescence on its anterior face, although its dorsal surface is almost bare and strongly shining (as in *rufiventris*): notaulices with only their lateral extensions

5 Beitr. Ent. 18, H. 1/2

distinct, not extending longitudinally on the dorsal surface of the mesoscutum. Ovipositor  $(\mathfrak{P})$  rather long, as in *rufiventris*, slightly projecting beyond the apical tergite in the retracted position.

Wing as in *rufiventris* (fig. 55), with vein 2r very short.

Body length: 2.2 mm. Wing length: 2.7-3.0 mm. Wing spread: 5.7 to 6.5 mm.

Holotype  $\mathcal{Q}$  from puparium 12. x. 66 of Cerodontha (Poemyza) tatrica NOWAKOWSKI on Festuca carpatica, Kraków ravine, Tatry, Poland, em. xii. 66, leg. NOWAKOWSKI (PAN). Paratype  $\mathcal{Q}$  from puparium 6. x. 65 of Cerodontha (Dizygomyza) caricicola HERING on Carex silvatica, Górna Solnika reservation, Bieszczady, Poland, em. 11. iii. 66, leg. NOWAKOWSKI (GCDG).

Although very similar to the well-known *D. rufiventris* (NEES), I think that these insects must be regarded as a distinct species on account of their very different coloration. They are also larger than any specimen of *rufiventris* which I have seen, although too much reliance should not be placed on this character as the size variation found in series of *Dapsilarthra* species is often very considerable.

In all published keys to *Dapsilarthra* (including that in Part II of this paper, page 555) this new species will run to the same couplet as *rufiventris* by virtue of its wing venation. I consider that it should be regarded as the sister-species of *rufiventris*, being clearly synapomorph with that species in respect of the short vein 2r and the long ovipositor.

# Dacnusini

Laotris NIXON

# Laotris striatula (HALIDAY)

Alysia (Dacnusa) striatula HALIDAY, 1839 Dacnusa striatula (HALIDAY), MARSHALL, 1891, 1895 and 1897, NIXON, 1937 Laotris striatula (HALIDAY), NIXON, 1943 and 1954, GRIFFITHS, 1964

Colour. Palpi ochreous or brownish. Labrum yellow-brown or brown. Clypeus black, like the face. Centre of mandibles red-brown. Antennae entirely dark. Legs varying from deep yellow to testaceous or red-brown, with the tarsi, the apex of the hind tibiae and occasionally the hind coxae somewhat infuscated. Gaster dark, at most with tergites 3 and 4 red-brown.

Morphology. Antennal segments:  $3, 26-28; \varphi, (24)-25-28$ . Palpi of moderate length (see the table of biometric data). Mandible (fig. 37) not expanded, with four teeth of which tooth 3 is long and pointed, but tooth 2 relatively small and inconspicuous (contrast *Chorebus*, in which tooth 2 is always the longest tooth). Face punctate, densely covered with fine pubescence which is directed mostly inwards over its centre but downwards along the eye-margins.

Thorax elongate, about 1.5 times as long as high. Pronotum with a large medial pit: its sides are largely bare, with only a little fine pubescence below the oblique suture; this suture is distinctly costate, but the surface above it is virtually smooth and strongly shining. Mesoscutum with its anterior face rough-

ened, with fine, rather long pubescence covering almost its entire surface (but tending to become sparse on the posterior half of the lateral lobes): notaulices well developed as rugose furrows at least anteriorly, sometimes almost reaching the posterior fovea: usually a smooth groove along the centre-line of the mesoscutum continues forward from the posterior fovea. Mesepisternum with a broad, conspicuously rugose-costate precoxal suture, which extends virtually to its hind margin: epicnemial suture less conspicuous, only weakly costate. Entire surface of metapleuron, propodeum and petiole strongly sculptured: the base of the hind coxa is also distinctly sculptured. Metapleural pubescence sparse and fine (compare fig. 16), with little differentiation in the direction of the hairs. Propodeum with only very fine, inconspicuous pubescence. Petiole usually slightly widened towards its apex, with only sparse inconspicuous pubescence. Surface of tergite 3 almost entirely longitudinally striate, with some fine pubescence at least at its sides. Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

Wing (fig. 186) with cell  $2R_1$  rather elongate; vein  $R_s$  sinuate; 1m-cu rejected from cell  $R_s$ ; cell 2Cu closed by  $Cu_{1b}$  at its lower distal corner.

# Breeding records

Host – Cerodontha (Dizygomyza) luctuosa MEIGEN (= effusi KARL)

25 ex. from puparia on Juncus effusus, Stuttgart (Wildpark and Feuerbach Tal), Germany, em. 10-20. xii. 53, leg. GROSCHKE (STGT and GCDG). 3 ex. from larvae and puparia 10 to 15. vii. 64 on Juncus effusus, Sieraków reservation, Kampinoska Forest, Poland, em. 22. vii-7. viii. 64, leg. NOWAKOWSKI (PAN). 2 33, 2 99 from puparia 6. xi. 66 on Juncus effusus, Palmiry, Kampinoska Forest, em. 6-10. ii. 67, leg. NOWAKOWSKI (PAN). 5 ex. from larvae and puparia 30. vii. 64 on Juncus effusus, Mirachowo, distr. Kartuzy, Poland, em. 17. viii-17. ix. 64, leg. NOWAKOWSKI (PAN). 1 ex. from larva 25. vii. 64 on Juncus effusus, Żukowo, distr. Kartuzy, em. 10. viii. 64, leg. NOWAKOWSKI (PAN). 1 ex. from larva 23. vii. 64 on Juncus effusus, Górki Wschodnie, near Gdansk, Poland, em. 17. viii. 64, leg. NOWAKOWSKI (PAN).

Hitherto this has been the only known species of *Laotris*.

# Laotris rupestris sp. nov.

Similar to *L. striatula* (HALIDAY), with which it may be compared as follows. Colour darker. Palpi and labrum dark brown. Legs brown with all coxae virtually black. Gaster entirely dark.

Morphology. 28 antennal segments ( $\mathcal{Q}$ ). Mandibles similar to those of *striatula*, but virtually 3-toothed, with the additional tooth (tooth 2 in *striatula*) represented only by a dilation at the base of the long central tooth.

Sides of pronotum almost completely bare. Metapleuron with most of its surface almost smooth and strongly shining, but there is some coarse rugose sculpture adjacent to the hind coxa. Propodeum and petiole strongly sculptured throughout, as in *striatula*. Tergite 3 with distinct longitudinal striation only on about its basal half, with fine pubescence sparsely distributed over most of its surface.

 $5^*$ 

Host – Cerodontha (Dizygomyza) sp.

Holotype ♀ from puparium 12. x. 66 on *Carex sempervirens*, Kraków Ravine, Tatry, Poland, em. 27. i. 67, leg. Nowakowski (PAN).

I am convinced that this specimen represents a second species of *Laotris*, as the smooth appearance of its metapleuron is unlike that of any specimen of *striatula* which I have seen. In other respects the two species are very similar.

# Exotela Förster

# Exotela flavicoxa (THOMSON)

Dacnusa (Dacnusa) flavicoxa Thomson, 1895
 Dacnusa melanocera Thomson sensu Nixon, 1937 (? Dacnusa (Dacnusa) melanocera Thomson, 1895)
 Antrusa melanocera (Thomson) sensu Nixon, 1943 and 1954

Exotela flavicoza (THOMSON), GRIFFITHS, 1964 and 1966a

I have already given a description of this species in Part II, where I recorded it from several hosts belonging to the Agromyza nigripes group and Cerodontha (Dizygomyza) incisa MEIGEN. The complete records known to me for Cerodontha hosts are as follows.

#### Host 5 — Cerodontha (Poemyza) incisa MEIGEN

1 9 from larva 21. vi. 61 on Phalaris arundinacea, Woodwalton Fen, Hunts., England, em. 10. vii. 61 (GCDG). 2 33 from larvae 5. ix. 60 on Phalaris arundinacea, Ash Vale, Surrey, England, em. 13 and 17. iii. 61 (GCDG). 2 99 from puparia 14. viii. 35 on Phalaris arundinacea, Warsow/Neukalen, Mecklenburg, Germany, em. 27. viii and l. x. 35, leg. BUHR (GCDG). 1 & from Calamagrostis epigejos, Rostock-Mönkweden, Mecklenburg, em. 11. viii. 36, leg. BUHR (GCDG). 3 99 from puparia 28. vi. 56 on Calamagrostis epigejos, Sieraków, Kampinoska Forest, Poland, em. 22-23. iv. 57, leg. Nowakowski (PAN): 1  $\bigcirc$  from puparium 26. vii. 56, same plant and locality, em. 29. iv. 57, leg. NOWAKOWSKI (PAN): 2 33 from larvae 16. vi. 66 on Calamagrostis arundinacea, same locality, em. 5. vii. 66, leg. No-WAKOWSKI (PAN): 1 3 from puparium 16. vi. 66 on Calamagrostis canescens, same locality, em. 17. ii. 67, leg. Nowakowski (PAN). 39 from puparia 31. vii. 66 on Agropyron repens, Otwock Wielki, near Warszawa, Poland, em. 12 and 18. viii. 66, leg. Nowakowski (PAN). 1 3 from larva 13. vii. 66 on Agropyron repens, Wierzba, near Popielno, Piska Forest, Poland, em. 28. vii. 66, leg. NOWAKOWSKI (PAN). 1 ex. from puparium 26. vi. 66 on Agropyron repens, Mierzeja Wiślana, Kąty Rybackie, Poland, em. 10. vii. 66, leg. NOWAKOWSKI (PAN). 1 3 from larva 24. vi. 66 on Poa trivialis, Mierzeja Wiślana, Krynica Morska, em. 15. ii. 67, leg. Nowakowski (PAN).

# Host 6 – Cerodontha (Poemyza) pygmaea Meigen

1  $\circ$  from puparium 16. vii. 61 on *Deschampsia caespitosa*, Chippenham Fen, Cambs., England, em. 17. vii. 61, leg. SPENCER (GCDG). 1  $\circ$  from puparium 14. viii. 35 on *Festuca gigantea*, Warsow, near Neukalen, Mecklenburg, Germany, em. 24. viii. 35, leg. BUHR (GCDG). 4  $\circ \circ$  from puparia 3. viii. 65 on *Deschampsia caespitosa*, Stadtwald, Mühlhausen, Thuringia, Germany, em. 14, 18. viii. 65 and 7-8. iii. 66, leg. BUHR no. 2558 (GCDG). 1  $\circ$  from puparium on *Dactylis glomerata*, Berlin-Dahlem, Germany, em. 30. ix. 65, leg. HERING no. 7475 (GCDG). 14 ex. from puparia 17 and 26. vi. 64 on *Dactylis glomerata*, Warszawa-Pyry, Kabackie Forest, Poland, em. 30. vi-19. viii. 64, leg. NOWAKOWSKI (PAN): 1 ex. from larva 21. vi. 64 on *Deschampsia caespitosa*, same locality, em. 14. vii. 64, leg. NOWAKOWSKI (PAN). 4 ex. from puparia 30. vi. 66 on *Dactylis glomerata*, Warszawa-Bielany, em. 5-22. vii. 66, leg. NOWAKOWSKI (PAN). 4 ex. from puparia 15. vii. 66 on *Calamagrostis arundi* 

nacea, Strzałowo reservation, Piska Forest, Poland, em. 25. vii (3 ex.) and 29. viii. 66, leg. NOWAKOWSKI (PAN): 1  $\bigcirc$  from puparium on *Dactylis glomerata*, same date and locality, em. 22. vii. 66, leg. NOWAKOWSKI (PAN). 1  $\heartsuit$  from larva 9. vii. 66 on *Deschampsia caespitosa*, Wierzba, near Popielno, Piska Forest, em. 22. vii. 66, leg. NOWAKOWSKI (PAN). 13 ex. from puparia 3. vii. 56 on *Dactylis glomerata*, Warszawa-Młociny, Poland, em. 6. vii— 10. viii. 56, leg. NOWAKOWSKI (PAN): 5 ex. from larvae and puparia 11. vi. 64, same plant and locality, em. 23. vi—20. viii. 64, leg. NOWAKOWSKI (PAN). 2  $\heartsuit$  from larvae 18. vi. 66 on *Deschampsia caespitosa*, Sieraków reservation, Kampinoska Forest, Poland, em. 5. vii and 16. ix. 66, leg. NOWAKOWSKI (PAN).

Host 7 — Cerodontha (Poemyza) phragmitidis NOWAKOWSKI (= atra auctt.) l  $\varphi$  from larva 13. vii. 66 on Phragmites communis, Wierzba, near Popielno, Piska Forest, Poland, em. 31. viii. 66, leg. NOWAKOWSKI (PAN).

#### Host 8 — Cerodontha (Poemyza) phalaridis Nowakowski

1 ζ from larva 18. vi. 66 on *Phalaris arundinacea*, Sieraków reservation, Kampinoska Forest, Poland, em. 10. vii. 66, leg. Νοwakowski (PAN). 1  $\varphi$  from larva 25. vi. 66 on *Phalaris arundinacea*, Mierzeja Wiślana, Krynica Morska, Poland, em. 30. viii. 66, leg. Nowakowski (PAN).

#### Host 9 — Cerodontha (Crastemyza) flavocingulata StrobL

 $1 \circ$  from larva 29. vi. 65 on *Poa trivialis*, Warszawa-Pyry, Kabackie Forest, Poland, em. vii. 65, leg. Nowakowski (PAN). 1 ex. from larva 3. vii. 56 on *Dactylis glomerata*, Warszawa-Młociny, em. 9. v. 57, leg. Nowakowski (PAN).

The study of the considerable material bred from C. (P.) pygmaea MEIGEN listed above has convinced me that there is no justification for regarding it as representing a distinct species, as was done in my key to *Exotela* in Part II (GRIFFITHS, 1966a). The average number of antennal segments in material bred from this host (29 in the male, and 27-28 in the female) is lower than that of material bred from C. (P.) incisa MEIGEN and the several Agromyza hosts recorded in Part II, but, since the host is smaller, this difference may be due merely to the correlation of the number of antennal segments with size.

It thus seems likely that Exotela melanocera (THOMSON) (= Antrusa persimilis NIXON) is a synonym of flavicoxa, although there remains a further point to be clarified. The lowest number of antennal segments in the material before me bred from C. (P.) pygmaea MEIGEN is 25 ( $\mathcal{Q}$ ). However NIXON (1954) states that the peak number of segments in his material of "Antrusa persimilis" was 24 ( $\mathcal{Q}$ ), and the lectotype of melanocera also has only 24 segments (see Appendix V in Part I). Until bred material is available which fully corresponds with these very small specimens, I am not proposing to establish a formal synonymy.

The range of antennal segments in the bred material which I have non accepted as *flavicoxa* (from all hosts) may be expressed as follows:  $\Im$ , (27)-29-34;  $\Im$ , (25)-26-32.

# Dacnusa Haliday

There are no *Dacnusa* species known as specific parasites of *Cerodontha* hosts, but a few specimens have been obtained of two species known also from other host genera.

# Dacnusa adducta (HALIDAY)

In addition to the records of this species from *Liriomyza* hosts given in Part V, I have received the following material bred from a *Cerodontha* species.

Host 2 – Cerodontha (Poemyza) pygmaea MEIGEN

 $1 \circ$  from puparium 17. vi. 64 on *Dactylis glomerata*, Warszawa-Pyry, Kabackie Forest, Poland, em. 6. ix. 64, leg. Nowakowski (PAN). 1  $\sigma$  from larva 11. vi. 64 on *Dactylis glomerata*, Warszawa-Młociny, em. 5. v. 65, leg. Nowakowski (PAN).

On the limited information available it is not clear whether this is an oligophagous species or a species with a disjunct host distribution.

# Dacnusa maculipes THOMSON

In addition to the records of this species bred from other genera of Agromyzidae given in Parts II, III and V, I have seen the following material bred from *Cerodontha* hosts.

 ${
m Host}~55-Cerodontha~(Poemyza)~calamagrostidis~{
m Nowakowski}$ 

 $1 \circ$  from larva 21. vi. 61 on *Calamagrostis epigejos*, Woodwalton Fen, Hunts., England, em. 23. ix. 61 (GCDG) (host previously recorded as "*Phytobia (Poemyza) muscina* MEIGEN" in GRIFFITHS, 1963).

#### Host 56 — Cerodontha (Poemyza) pygmaea MEIGEN

1 & from puparium 5. vii. 66 on *Lolium perenne*, Hauswalde, Kreis Kamenz, Saxony, Germany, em. 10. ix. 66, leg. BUHR no. 2960 (GCDG).

Host 57 — Cerodontha (Dizygomyza) iridis HENDEL

2 ex. from puparia 1. ix. 53 on Iris sp., Hampstead, London, leg. SPENCER (BM) (host previously recorded as "*Phytobia (Dizygomyza) iraeos* ROBINEAU-DESVOIDY" in GRIFFITHS, 1956).

This species occurs much more commonly on *Phytomyza*, *Trilobomyza* and *Liriomyza* hosts.

# The Chorebus talaris group

I include in this concept, here proposed for the first time, the three species treated below and one species of unknown life-history, C. rhanis (NIXON). These insects appear synapomorph in respect of their well-defined rugose metapleural swelling which is evenly covered with long, fairly dense pubescence (fig. 20), the presence of sculpturation at least on the base of the hind coxae, and their densely pubescent mesoscutum and petiole. Only one of the four species, the well known C. talaris (HALIDAY), seems to be widely distributed. The two new species described below are part of the high altitude fauna of the Tatry mountains, and rhanis is so far known only from Scotland.

# Chorebus talaris (HALIDAY)

Alysia (Dacnusa) talaris HALIDAY, 1839 (in part)

Dacnusa talaris (HALIDAY), MARSHALL, 1891 (in part), 1895 (in part) and 1897 (in part), NIXON, 1937

Etriptes talaris (HALIDAY), NIXON, 1943 and 1954 Chorebus talaris (HALIDAY), GRIFFITHS, 1964 Colour. Palpi deep yellow or yellow-brown. Labrum brown or black. Clypeus black. Centre of mandibles yellow-brown. Antennae entirely dark. Legs ochreous yellow or yellow-brown, with the tarsi (especially the hind tarsi), the hind coxae and the apex of the hind tibiae somewhat infuscated. Gaster entirely dark.

Morphology. Antennal segments: 3, 30-33; 9, 30-32. Palpi very long (see the table of biometric data). Mandible (fig. 201) not expanded, with tooth 2 relatively long and pointed but tooth 3 appearing only as a projection from its side. Face shining, weakly punctate, with fine pubescence directed inwards over its centre but downwards along the eye-margins.

Thorax 1.4—1.5 times as long as high. Sides of pronotum smooth, bare and shining centrally, but with some fine pubescence below the oblique suture. Mesoscutum slightly roughened anteriorly, with almost its entire surface densely pubescent: notaulices weak, with only their lateral extensions distinct or only shortly extending longitudinally on the dorsal surface of the mesoscutum. Precoxal suture narrow, only weakly rugose (occasionally virtually smooth). Metapleuron with a conspicuous rugose swelling which is evenly covered with long, fairly dense pubescence (fig. 20): the hind coxa is also rugose at its base. Propodeal pubescence fairly dense, but fine and inconspicuous, not obscuring the very strongly rugose surface beneath. Petiole slightly widened towards its apex, densely covered with fine inconspicuous pubescence. Tergite 3 with conspicuous rugose sculpture, similar to that of the petiole, at its base and with fine hairs distributed over most of its surface. Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

Wing (fig. 187) with cell  $2R_1$  rather elongate; vein  $R_s$  strongly sinuate; 1m-cu well rejected from cell  $R_s$ ;  $Cu_{1b}$  weak.

Breeding records

Host 1 – Cerodontha (Poemyza) pygmaea MEIGEN

1 9 from puparium 30. viii. 66 on Brachypodium silvaticum, Poulavallan, Clare, Ireland, em. 1. ix. 66 (GCDG). 1 9 from puparium 9. xi. 24 on Brachypodium sp., Cothill. Berks.. England, em. 22. v. 25, leg. HAMM (HD). 2 99 from puparia on Festuca gigantea, Mühlhausen, Thuringia, Germany, em. vii. 64, leg. BUHR, HERING no. 2140 (GCDG): 1 3 from puparium 19. ix. 66, same plant and locality, em. 27. x. 66, leg. BUHE no. 3197 (GCDG): 2 99 from puparia 5. viii. 65 on Dactylis aschersoniana, same locality, em. 14-15. viii. 65, leg. BUHR no. 2576 (GCDG): 2 99 from puparia 29. vii. 66, same plant and locality, em. 9. viii. 66, leg. BUHR no. 3073 (GCDG). 1 9 from puparium 9. vii. 66 on Agropyron repens, Rammenau, Saxony, Germany, em. 2. viii. 66, leg. BUHR no. 3003 (GCDG). 2 99 from Dactulis glomerata, Hedlandet, Södermanland, Sweden, em. 19. vii. 43, leg. LUNDQVIST (LUND). 2 ex. from puparia 24. viii. 65 on Festuca silvatica, Szeroki Wierch, Bieszczady, Poland, em. 8. x. 65 and 8. iv. 66, leg. NowAkowski (PAN). 1 3, 2 99 from puparia 13 and 28. viii. 65 on Dactylis glomerata, Cisna, Bieszczady, em. 20. viii. 65 (1 9) and 18. iii. 66, leg. Nowakowski (PAN). 1 ex. from puparium 25. vii. 64 on Festuca silvatica, Kartuzy, near Ciche lake, Poland, em. 8. v. 65, leg. NOWAKOWSKI (PAN). 2 ex. from puparia 30. vii. 64 on Molinia coerulea, Mirachowo, distr. Kartuzy, em. 3. v. 65, leg. NOWAKOWSKI (PAN). 1 9 from puparium 15. vii. 66 on Calamagrostis arundinacea, Strzałowo reservation, Piska Forest, Poland, em. 25. vii. 66, leg. NOWAKOWSKI (PAN).

# Host 2 – Cerodontha (Poemyza) incisa Meigen

d<sup>♀</sup> from puparia 15. vii. 66 on Calamagrostis arundinacea, Suchy Groń, Babia Góra, Poland, em. viii. 66 and 17. iii. 67, leg. Nowakowski (PAN): 10 ex. from puparia 15. viii. 66 on Festuca gigantea, same locality, em. 30. viii – 9. ix. 66, leg. Nowakowski (PAN). 4 ex. from puparia 14–15. viii. 65 on Calamagrostis arundinacea, Łopiennik, Bieszczady, Poland, em. 31. viii – 3. ix. 65, leg. Nowakowski (PAN): 3 ex. from puparia 13–14. viii. 65 on Agropyron repens, same locality, em. 3–6. ix. 65, leg. Nowakowski (PAN). 1 ex. from puparium 17. viii. 65 on Calamagrostis arundinacea, Falowa, Bieszczady, em. viii. 65, leg. Nowakowski (PAN). 1 ♂ from puparium 18. viii. 65 on Agropyron repens, Cisna, Bieszczady, em. 14. iii. 66, leg. Nowakowski (PAN). 1 ♀ from puparium 24. viii. 65 on Festuca silvatica, Szeroki Wierch, Bieszczady, em. 2. ix. 65, leg. Nowakowski (PAN). 2 ex. from puparia 21. ix. 64 on Calamagrostis canescens, Wilkowska Valley, Łysogóry, Poland, em. 10–12. v. 65, leg. Nowakowski (PAN).

### Host 3 — Cerodontha (Poemyza) tatrica Nowakowski

1 3 from larva 15. viii. 66 on *Calamagrostis arundinacea*, Suchy Groń, Babia Góra, Poland, em. 2. iii. 67, leg. Nowakowski (PAN). 1 3 from puparium 9. ix. 60 on *Calamagrostis* arundinacea, Spadowiec Valley, Tatry, Poland, em. 1. vi. 61, leg. Nowakowski (PAN).

This species differs from the other species of the *talaris* group by the presence of sculpturation at the base of tergite 3, the longer cell  $2R_i$  and the rather elongate mandibles.

### Chorebus subasper sp. nov.

Colour. Palpi and labrum yellow. Clypeus dark. Centre of mandibles redbrown. Antennae brownish at their base as far as about the second flagellar segment. Legs uniformly deep yellow, with the hind coxae slightly darker (more or less brown). Gaster entirely dark.

Morphology. 29 antennal segments ( $\mathcal{Q}$ ). Palpi very long (see the table of biometric data). Mandible not expanded, with tooth 2 relatively large (but not so long and pointed as in *talaris*) and tooth 3 appearing as a projection from its side. Face fairly strongly punctate, only weakly shining, with pubescence directed inwards over its centre but downwards along the eye-margins.

Thorax about 1.5 times as long as high. Sides of pronotum with their entire surface sculptured, largely bare and shining, but with some fine inconspicuous pubescence below the oblique suture. Mesoscutum entirely covered with dense pubescence, with rather coarse, punctate sculpture extending over almost its entire surface (particularly conspicuous on its anterior face and central lobe): notaulices weak, distinct anteriorly only. Rugose precoxal suture well developed. Metapleuron as in *C. talaris* (HALIDAY) (compare fig. 20), with a conspicuous rugose swelling which is evenly covered with long, fairly dense pubescence. The hind coxa is also largely covered with rugose sculpture. Propodeal pubescence fairly dense, but fine and inconspicuous, not obscuring the very strongly rugose surface beneath. Petiole slightly widened towards its apex, densely covered with fine inconspicuous pubescence. Tergite 3 almost bare, without any rugosity at its base. Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

Wing (compare fig. 188) with cell  $2R_1$  shorter than in *talaris*; vein  $R_s$  distinctly sinuate; 1m-cu rejected from cell  $R_s$ ;  $Cu_{1b}$  weak.

# Host — Cerodontha (Poemyza) alpina Nowakowski

Holotype  $\Im$  from puparium 24. ix. 66 on *Trisetum alpestre*, Kraków ravine, Tatry, Poland, em. 6. iii. 67, leg. Nowakowski (PAN).

Although lacking sculpturation of tergite 3, this species and C. asperrimus sp. nov. (described below) have the thorax more extensively sculptured than in *talaris*. They also clearly differ from that species in having a shorter cell  $2R_1$ .

# Chorebus asperrimus sp. nov.

Similar to C. subasper sp. nov., with which it may be compared as follows. Colour much darker. Palpi dark brown. Labrum and clypeus virtually black. Centre of mandibles red-black. Antennae entirely dark. Legs brown, with the coxae and hind femora virtually black. Gaster entirely dark.

Morphology. Antennal segments:  $\mathcal{J}, 35; \mathcal{Q}, 30$ . Face more strongly sculptured, coarsely and conspicuously rugose. The centre-line of the frons (from the front ocellus) is impressed, and in the holotype (the larger specimen) is indicated by a rugose furrow.

The sculpture on the hind coxa is very coarse and conspicuous, and the middle coxa, middle femur and hind femur are also largely covered with punctate sculpture.

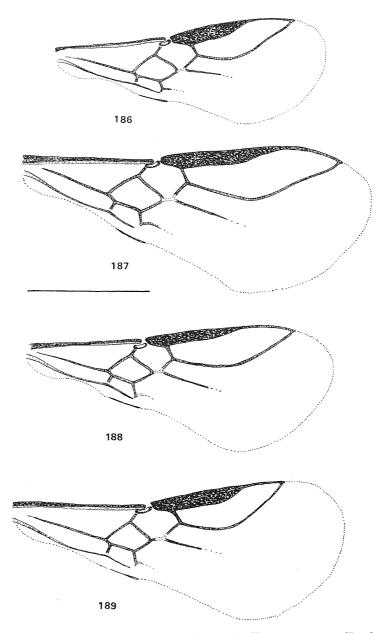
# Host — Cerodontha (Poemyza) tatrica Nowakowski

Holotype  $\Im$  from puparium 14. x. 66 on *Calamagrostis arundinacea*, Kraków ravine, Tatry, Poland, em. 14. iv. 67, leg. Nowakowski (PAN): paratype  $\Im$  from puparium 24. ix. 66, same plant and locality, em. 24. x. 66, leg. Nowakowski (GCDG).

The holotype of C. *rhanis* (NIXON), described from Scotland, is a similar darklegged insect, but I do not think that it can be conspecific with the above Tatry material because its face is only weakly punctate (as in *talaris*), and the sides of the pronotum virtually smooth.

# The Chorebus ovalis/lateralis complex

I have provisionally adopted the term "ovalis/lateralis complex" (see Part I, page 850) for those species of Chorebus which have a well-defined rosette of metapleural pubescence (as also in the C. senilis group, C. affinis group and related species), but with vein  $Cu_{1b}$  weak or absent. Species of this complex occur as parasites of most leaf-mining Agromyzidae, and many species have already been treated in Parts II, III and V of this paper. Twelve further species are now recorded as parasites of Cerodontha sensu lato, including both the known parasites of subgenus Cerodontha sensu stricto and a majority of the known parasites of subgenus Poemyza. However only two species of this complex are known to be associated with hosts of the subgenus Dizygomyza, in contrast with the affinis group and its relatives in which this host distribution is reversed (many species being known as parasites of subgenera Dizygomyza and Butomomyza, but only



Figs. 186–189. Wings of: 186, Laotris striatula (HALIDAY)  $\varphi$ ; 187, Chorebus talaris (HALIDAY)  $\varphi$ ; 188, Chorebus asperrimus sp. nov.  $\varphi$ ; 189, Chorebus vitripennis sp. nov.  $\varphi$ . (Scale 1 mm.)

two recorded from *Poemyza* hosts). This strongly suggests a former vicariance between the two groups, which may be of considerable evolutionary interest when it is possible to make a firmer assessment of the phylogenetic relationship between them.

It has not been possible to establish in any detail the relationships between the species of the *ovalis/lateralis* complex, because there has evidently been much convergent evolution of many of the characters important for diagnostic purposes, such as the mesoscutal and petiolar pubescence and the number of antennal segments. One species, described below as *C. difficilis* sp. nov., seems referable to the *alecto* group which contains many species of *Phytomyza*- and *Liriomyza*-parasites (see Parts III and V). Many of the other species also have similar petiolar pubescence to the *alecto* group, but combined (except in *asramenes* and *agraules*) with a much more sparsely pubescent mesoscutum. A firm interpretation of the degree of relationship between most of these species does not seem possible on present information.

It should be noted that rather more overlapping of host ranges occurs in the species of the *ovalis/lateralis* complex associated with *Cerodontha* subgenus *Poemyza* than has been found in the species associated with other genera of Agromyzidae treated in previous parts of this paper. In several bred series two or three closely related species were found to be present. Special care should therefore be taken when studying series bred from these hosts to ensure that they are homogeneous.

In the descriptions which follow the following characters may be assumed whenever nothing to the contrary is stated.

Mesepisternum with a well-defined rugose-costate precoxal suture. Metapleural pubescence dense, forming a rosette around a raised swelling. Wing venation rather uniform in most species (compare figs. 87 and 88), with the pterostigma longer than the metacarp, Im-cu clearly rejected from cell  $R_s$ , and vein  $Cu_{1b}$ weak or absent, so that cell 2Cu is more or less open at its lower distal corner. Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

### Chorebus flavipes (GOUREAU)

Dacnusa flavipes GOUREAU, 1851 Dacnusa raissa NIXON, 1937 and 1945 Chorebus flavipes (GOUREAU), GRIFFITHS, 1966b

Colour. Palpi and labrum yellow. Clypeus dark. Centre of mandibles yellowbrown. Antennae usually yellow or yellow-brown at their base as far as about the second or third flagellar segment (but with only the scape and pedicel obscurely yellow-brown in the Scottish specimen). Legs yellow except that tarsal segments 5 are contrastingly black. Gaster entirely dark or at most with tergites 3 and 4 brownish.

Morphology. Antennal segments:  $\mathcal{J}$ , (28)-30-32;  $\mathcal{Q}$ , 29-32. Palpi long (see the table of biometric data). Mandible not or hardly expanded, clearly 4-toothed but with tooth 3 relatively small. Face weakly punctate.

Thorax about 1.4 times as long as high. Sides of pronotum bare and smooth centrally, with a little fine pubescence mainly along and below the oblique suture. Mesoscutum with only its anterior face roughened and pubescent, but its dorsal surface smooth, shining and bare apart from a few hairs along the course of the notaulices: notaulices usually distinct anteriorly only (but occasionally extending as weak impressions almost to the posterior fovea). Precoxal suture rather broad. Metapleural swelling strongly rugose-punctate. Metapleural and propodeal pubescence very dense and whitish. Petiole widened towards its apex, with short fine inconspicuous pubescence distributed over most of its surface (except the central part of about its apical half), usually becoming denser towards the apical corners although no tufts are formed (the pubescence is often so inconspicuous that the petiole might at first sight be considered bare). Tergite 3 without basal hairs: the base of tergite 3 is rugose (as in *C. talaris* (HALIDAY)) in the two Stuttgart specimens, but this feature is not shown by the other material. Wing with vein  $R_s$  only weakly sinuate.

### Breeding records

Host - Cerodontha (Dizygomyza) iraeos ROBINEAU-DESVOIDY

1 δ from puparium 28. viii. 65 on Iris pseudacorus, Armadale, Skye, Scotland, em. 15. x. 65, leg. SPENCER (GCDG). 5  $\Im$  from puparia 16. vii. 66 on Iris pseudacorus, Woodwalton Fen, Hunts., England, em. 26. vii -1. viii. 66 (GCDG). 6 ex. from puparia 25. x and 8. xi. 24 on Iris pseudacorus, Oxford, England, em. 14-20. v. 25, leg. HAMM (HD and BM). 2  $\Im$  δ, Stuttgart-Wildpark, Germany, em. 10 and 25. ii. 54, leg. GROSCHKE (STGT). 1 ex. from puparium on Iris pseudacorus, Berlin-Grünewald, 1954, leg. HERING no. 6048 (BM). 4  $\Im$  δ from puparia on Iris pseudacorus, Kazuń, Kampinoska Forest, Poland, em. 16. v. 56, leg. Nowakowski (PAN). 10 ex. from puparia 27 and 31. vii. 55 on Iris pseudacorus, Granica, Kampinoska Forest, em. 31. vii-6. viii. 55, leg. Nowakowski (PAN). 1  $\Im$  from larva 19. ix. 54 on Iris pseudacorus, Sieraków reservation, Kampinoska Forest, em. 9. v. 55, leg. Nowakowski (PAN): 1 ex. from puparium 27. ix. 55, same plant and locality, em. 19. v. 56, leg. Nowakowski (PAN). 1 ex. from puparium 15. ix. 60 on Iris pseudacorus, Mątowski Forest reservation, Piekło near Sztum, Poland, em. 20. iv. 61, leg. Nowakowski (PAN).

GOUREAU's (1851) original material was bred from this same host in France.

The synonymy of NIXON'S *Dacnusa raissa* was proposed in Appendix VIII in Part III (GRIFFITHS, 1966b).

This species may be recognised by its bare mesoscutum, pale coloration and the inconspicuously pubescent petiole.

Chorebus crenulatus (THOMSON)

Dacnusa (Dacnusa) crenulata Thomson, 1895 Dacnusa elegantula Nixon, 1937 and 1945 Chorebus crenulatus (Thomson), Griffiths, 1964

Colour. Palpi and labrum deep yellow or yellow-brown. Clypeus dark. Centre of mandibles yellow-brown. Antennae more or less entirely dark, at most becoming reddish brown towards their base. Legs largely reddish yellow or yellowbrown, with the hind coxae usually somewhat infuscated. Gaster more or less entirely dark. Morphology. Head large and subcubical (1.5-1.6 times as wide as long). Antennal segments:  $3, 30-34; \varphi, 26-30$ . Palpi long (see the table of biometric data). Mandible not expanded, with tooth 2 long and pointed but tooth 3 appearing only as a projection from its side (compare *asramenes*, fig. 202). Face only weakly punctate.

Thorax elongate, about 1.5 times as long as high. Sides of pronotum with fine pubescence mainly along and below the oblique suture. Mesoscutum with its anterior face pubescent and slightly roughened, but its dorsal surface largely smooth and shining, with pubescence only along the course of the notaulices and sparsely on its central lobe (but occasionally this too is almost bare): notaulices well defined, reaching the posterior fovea: often the centre-line of the mesoscutum is impressed to form a longitudinal groove. Metapleural swelling strongly rugose-punctate. Metapleural and propodeal pubescence very dense and whitish. Petiole only slightly widened towards its apex, with dense pubescence covering its entire surface and forming distinct apical tufts (compare fig. 144). Tergite 3 without basal hairs.

Wing with vein  $R_s$  weakly sinuate.

Breeding records

Host – Cerodontha (Cerodontha) denticornis PANZER

1ð from larva 25. vi<br/>. 64 on Festuca pratensis, Warszawa-Ochota, Poland, em. 13. vi<br/>i. 64, leg. Nowakowski (PAN).

The synonymy of this species was established in Part I (GRIFFITHS, 1964).

This species and C. diremtus (NEES) are characterised by the combination of a large subcubical head and a very narrow, elongate thorax. Both these species are parasites of *Cerodontha* sensu stricto, whose long narrow puparia may be found in the leaf-sheaths of various Gramineae. Possibly other species with similar proportions will be discovered, when further material bred from this host-group is obtained. Another characteristic feature of *crenulatus* is the pubescence of the petiole, which is similar to that of *C. asramenes* (NIXON).

Chorebus diremtus (NEES), comb. nov.

Alysia diremta NEES, 1834

Alysia (Dacnusa) diremta NEES, HALIDAY, 1839 (in part)

Dacnusa diremta (NEES), MARSHALL, 1891 (in part), 1895 and 1897 (in part), NIXON, 1937, 1943 and 1946

Colour. Palpi yellow. Labrum and clypeus red-brown. Centre of mandibles yellow-brown. Basal antennal segments contrastingly yellow as far as about the second flagellar segment. Legs uniformly yellow or reddish yellow, with at most tarsal segments 5 slightly infuscated. Gaster beyond petiole largely reddish yellow, but becoming darker towards its apex.

Morphology. Head large and subcubical (less than 1.5 times as wide as long). Antennal segments:  $3, 30-33; \mathfrak{P}, (25)-26-28-(29)$ . Palpi long (see the table

of biometric data). Mandible not expanded, with tooth 2 long and pointed, much better developed than the other three teeth. Face weakly punctate.

Thorax elongate, about 1.5 times as long as high. Sides of pronotum largely smooth and bare, but with some fine inconspicuous pubescence along and below the oblique suture. Mesoscutum with only its "shoulders" roughened, largely smooth, bare and shining with pubescence almost confined to the shoulders apart from a few fine hairs along the course of the notaulices: notaulices weakly indicated anteriorly only. Precoxal suture long, extending towards the hind margin of the mesepisternum. Metapleural swelling strongly rugose-punctate. Metapleural and propodeal pubescence very dense and whitish. Petiole parallelsided or only slightly widened towards its apex, bare centrally but with a few hairs along its sides and thin accumulations of pubescence at its apical corners. Tergite 3 with few or no basal hairs.

Wing with vein  $R_s$  weakly sinuate.

# Breeding records

Host – Cerodontha (Cerodontha) fulvipes Meigen

l ${\tt q}$ from larva 6. vii. 64 on Poa trivialis, Warszawa-Bielany, Poland, em. 27. vii .64, leg. Nowakowski (PAN).

This species is readily recognisable by its very bright coloration, the proportions of the head and thorax, and the largely bare mesoscutum and petiole. My interpretation follows that of NIXON (NEES' types being all destroyed).

# Chorebus rubicundus sp. nov.

Colour. Palpi and labrum yellow. Clypeus black or reddish. Centre of mandibles orange-yellow. Antennae yellow or yellow-brown at their base as far as the second or third flagellar segment, this colour merging gradually into the dark colour of the more distal flagellar segments. Legs uniformly clear yellow, with only tarsal segments 5 weakly infuscated. Gaster beyond petiole entirely deep orange-yellow.

Morphology. 28 antennal segments  $(2 \Im \Im)$ . Palpi very long (see the table of biometric data). Mandibles strongly 4-toothed, only slightly expanded. Face weakly punctate.

Thorax about 1.4 times as long as high. Sides of pronotum almost bare centrally, but with some fine pubescence along and below the oblique suture. Mesoscutum somewhat roughened anteriorly, especially on its central lobe, with dense pubescence covering almost its entire surface: notaulices weak, with their longitudinal extensions virtually absent or indicated anteriorly only. Precoxal suture narrow, only weakly rugose. Metapleural swelling finely rugose-punctate. Metapleural and propodeal pubescence dense and whitish. Petiole slightly widened towards its apex, almost bare except for a few fine hairs at its apical corners (not forming distinct tufts). Tergite 3 almost bare.

Wing with vein  $R_s$  weakly sinuate.

#### Host — Cerodontha (Poemyza) pygmaea MEIGEN

Holotype  $\varphi$  from puparium on *Dactylis glomerata*, Mühlhausen, Thuringia, Germany, em. 25. ii. 62, leg. Buhr, HERING no. 1836 (GCDG). Paratype  $\varphi$  from puparium 3. vii. 56 on *Dactylis glomerata*, Warszawa-Młociny, Poland, em. 11. vii. 56, leg. NOWAKOWSKI (PAN).

This species may be recognised by its brightly coloured gaster, and the combination of an extensively public mesoscutum with a virtually bare petiole. The fact that only two specimens have been found in the considerable material of parasites bred from C. (P.) pygmaea MEIGEN available to me suggests that possibly this is not the principal host.

### Chorebus enephes (NIXON), comb. nov.

### Dacnusa enephes NIXON, 1945

Colour. Palpi and labrum yellow. Clypeus dark. Centre of mandibles yellowbrown. Antennae yellow-brown at their base, as far as the second or third flagellar segment. Legs largely yellow, with only tarsal segments 5 strongly infuscated (sometimes the apices of the hind femora and tibiae are also slightly infuscated). Gaster beyond petiole with most of its dorsal surface conspicuously reddish yellow, becoming darker only ventrally and towards its apex.

Morphology. Antennal segments: 3, (32)-34-37-(38); 9, 32-36. Palpi long (see the table of biometric data). Mandibles clearly 4-toothed, not or only slightly expanded (compare fig. 59). Face weakly punctate.

Thorax 1.2-1.3 times as long as high. Sides of pronotum almost bare centrally, but with dense fine pubescence along and below the oblique suture. Mesoscutum with only its anterior face roughened and pubescent, its dorsal surface being virtually bare, with only a few hairs along the course of the notaulices, and of strongly shining appearance with a faint indication of scaly-reticulate ground sculpture (but this is not so well developed as in *C. ganesa* (NIXON)): notaulices distinct anteriorly only. Metapleural swelling finely rugose-punctate. Metapleural and propodeal pubescence very dense and whitish. Petiole slightly widened towards its apex, bare and shining centrally, but with some fine pubescence near its base and along its sides, and conspicuous white apical tufts. Tergite 3 without basal hairs.

Wing with vein  $R_s$  only weakly sinuate.

Breeding records

### Host – Cerodontha (Poemyza) deschampsiae SPENCER

1 φ from larva 24. ix. 65 on *Deschampsia caespitosa*, Kiczerka near Tarnica, Bieszczady, Poland, em. 30. iii. 66, leg. NOWAKOWSKI (PAN). 1 δ from larva 22. vii. 66 on *Deschampsia caespitosa*, Stadtwald, Mühlhausen, Thuringia, Germany, em. 21. iv. 67, leg. BUHR no. 3055 (GCDG).

This species is readily recognisable by its brightly coloured gaster, bare mesoscutum and the contrasting apical tufts on the petiole.

### Chorebus ganesa (NIXON), comb. nov.

Dacnusa ganesa NIXON, 1945

Very similar to C. enephes (NIXON), with which it may be compared as follows. Colour. Gaster uniformly dark, or at most with tergites 3 and 4 reddish brown. Morphology. Antennal segments:  $\mathcal{J}, 34-36; \mathcal{Q}, 33-35$ . Mesoscutum entirely covered with fine scaly-reticulate ground sculpture: notaulices better developed, usually reaching the posterior fovea as well-defined linear grooves.

### Breeding records

Host – Cerodontha (Poemyza) deschampsiae Spencer

1 ♂ from larva 8. vii. 66 on *Calamagrostis canescens*, Röderbrunn, Oberlausitz, Saxony, Germany, em. 13-18. iv. 67, leg. BUHR no. 3012 b (GCDG).

I think that NIXON (1945) was probably right in separating this species and *enephes*, but it is desirable that additional material be obtained from the host to confirm that the differences given are constant.

# Chorebus ninella (NIXON)

# Dacnusa ninella NIXON, 1945 (in part)

Colour. Palpi and labrum yellow. Clypeus dark. Centre of mandibles yellowbrown. Antennae contrastingly yellow, yellow-brown or reddish at their base as far as about the second flagellar segment. Legs largely yellow, with only tarsal segments 5 distinctly infuscated (hind coxae either yellow or weakly infuscated, more or less brown): however in the Woodwalton female the legs are slightly darker, largely ochreous yellow with the hind coxae strongly infuscated. Gaster more or less entirely dark, at most with tergite 3 red-brown.

Morphology. Antennal segments:  $\mathcal{J}$ , (30) -31-34;  $\mathcal{Q}$ , 29-33. Palpi very long (see the table of biometric data). Mandible not expanded, clearly 4-toothed but with tooth 3 relatively small (compare fig. 59). Face weakly punctate or virtually smooth.

Thorax about 1.3 times as long as high. Sides of pronotum almost smooth and largely bare above the oblique suture, but with rather dense, fine pubescence along and below the suture. Mesoscutum with its anterior face and the anterior part of its central lobe roughened but its lateral lobes smooth and shining, with its anterior face and central lobe densely pubescent but its lateral lobes largely bare (usually with pubescence only about the "shoulders"): notaulices distinct at least anteriorly, in most specimens reaching the posterior fovea as well-defined linear grooves. Metapleural swelling strongly rugose- punctate. Metapleural and propodeal pubescence covering its entire surface and forming welldeveloped whitish apical tufts (compare C. asramenes (NIXON)). Tergite 3 with few or no basal hairs.

Wing with vein  $R_s$  weakly sinuate.

#### Breeding records

### Host — Cerodontha (Poemyza) calamagrostidis Nowakowski

4 33, 5 99 from larvae 8. vii. 66 on *Calamagrostis canescens*, Röderbrunn, Oberlausitz, Saxony, Germany, em. 13-26. iv. 67, leg. BUHR no. 3010b and 3011b (GCDG). 1 9 from larva 21. vi. 61 on *Calamagrostis epigejos*, Woodwalton Fen, Hunts., England, em. 23. iv. 62 (GCDG).

Important features for recognising this species are the range of antennal segments, the pale basal flagellar segments and the nature of the pubescence of the mesoscutum and petiole.

NIXON'S (1945) Swedish holotype is a female with 33 antennal segments which agrees fully with the German bred series listed above. The Woodwalton specimen is slightly more darkly coloured, but I have no doubt that it is conspecific. Like the host, this species has only a single generation a year.

NIXON (1945) also referred to this species, with some qualification, some specimens which I now refer to C. poemyzae sp. nov. See the remarks below under the description of that species.

In Part I (GRIFFITHS, 1966a) I referred to *ninella* a specimen bred from Agromyza hendeli GRIFFITHS. It seems unlikely that this specimen represented the true *ninella*, and it is desirable that further material should be obtained from this host in order that its identity may be clarified.

### Chorebus poemyzae sp. nov.

# Dacnusa ninella sensu NIXON, 1945 (in part)

Colour. Palpi and labrum yellow. Clypeus dark. Centre of mandibles yellowbrown. Antennae contrastingly yellow or yellow-brown at their base as far as about the second flagellar segment. Legs almost completely yellow, with only tarsal segments 5 distinctly infuscated. Gaster beyond petiole with tergites 3 and 4 usually red-brown, the following tergites dark.

Morphology. Antennal segments: 3, (34)-35-38; 9, (33)-34-37. Palpi long (see the table of biometric data). Mandible not expanded, clearly 4-toothed but with tooth 3 relatively small (compare fig. 59). Face weakly punctate.

Thorax 1.3-1.4 times as long as high. Sides of pronotum almost smooth, bare or only sparsely pubescent above the oblique suture, but with rather dense, fine pubescence along and below the suture. Mesoscutum with only its anterior face and sometimes also the anterior part of its central lobe roughened, with its anterior face and the anterior part of the central lobe covered with fine pubescence, but most of its dorsal surface smooth, bare and strongly shining: notaulices distinct as rugose furrows anteriorly, occasionally extending to the posterior fovea: a longitudinal groove often continues from the posterior fovea along the centre-line of the mesoscutum, but the development of this feature is variable. Metapleural swelling strongly rugose-punctate. Metapleural and propodeal pubescence very dense and whitish. Petiole widened towards its apex, with dense pubescence covering most of its surface and forming conspicuous white

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apical tufts: the centre-line tends to be more sparsely pubescent than the sides, and in some specimens becomes almost bare on its apical half. Tergite 3 without basal hairs.

Wing with vein  $R_s$  only weakly sinuate.

# Breeding records

# Host 1 – Cerodontha (Poemyza) incisa Meigen

62 paratypes, Denmark (25 ex., Donse, Sealand; 18 ex., Randers, Jutland; 6 ex., Damhusmose, Sealand; 6 ex., Utterslevmose, Sealand; 4 ex., Taastrup, Sealand; 3 ex., Køge, Sealand), leg. Schlick (KB). 3 99 paratypes from puparia 18 and 25. x. 24, probably on Phalaris arundinacea, Oxford, England, em. 6-25. v. 25, leg. HAMM (BM). 1  $\varphi$  paratype from puparium 8. vii. 66 on Calamagrostis canescens, Röderbrunn, Oberlausitz, Saxony, Germany, em. 19. iv. 67, leg. BUHR no. 2980b (GCDG). 19 paratype from puparium 28. vi. 56 on Calamagrostis arundinacea, Sieraków, Kampinoska Forest, Poland, em. 12. vii. 56, leg. Nowakowski (PAN): 1 3, 2 99 paratypes from puparia 25 and 28. vi. 56 on Calamagrostis epigejos, same locality, em. 5-13. vii. 56, leg. Nowakowski (PAN): 2 33 paratypes from puparia 31. x. 66 on Calamagrostis canescens, same locality, em. 6. ii. 67, leg. NowA-KOWSKI (PAN). 1  $\ominus$  holotype; 1 3, 1  $\ominus$  paratypes from larvae 13. vii. 66 on Agropyron repens, Wierzba, near Popielno, Piska Forest, Poland, em. 26-30. vii. 66, leg. Nowakowski (PAN). 1 3 paratype from puparium 14. viii. 65 on Agropyron repens, Lopiennik, Bieszczady, Poland, em. xii. 65, leg. Nowakowski (PAN). 3 paratypes from puparia 13 and 18. viii. 65 on Agropyron repens, Cisna, Bieszczady, em. 2. ix. 65 (2 ex.) and 18. iii. 66, leg. Nowakowski (PAN and GCDG). 1 3 paratype from larva 29. vi. 65 on Alopecurus pratensis, Warszawa-Pyry, Kabackie Forest, Poland, em. 14. iii. 66, leg. NOWAKOWSKI (PAN).

# Host 2 — Cerodontha (Poemyza) pygmaea Meigen

7 paratypes from larvae and puparia 17 and 26. vi. 64 on *Dactylis glomerata*, Warszawa-Pyry, Kabackie Forest, Poland, em. 20. vi-11. vii. 64 (6 ex.) and 20. v. 65 (1  $\Im$ ), leg. Nowakowski (PAN and GCDG). 1  $\Im$  paratype from puparium 13. viii. 65 on *Dactylis glomerata*, Cisna, Bieszczady, Poland, em. 18. iii. 66, leg. Nowakowski (PAN).

# Host 3 – Cerodontha (Poemyza) deschampsiae SPENCER

l $_{\rm J}$ paratype from puparium 8. vii. 66 on Calamagrostis canescens, Röderbrunn, Oberlausitz, Saxony, Germany, em. 13–18. iv. 67, leg. BUHR no. 3012b (GCDG).

This species is very similar to *C. ninella* (NIXON), and was not separated from that species by NIXON (1945). The main difference between these two species lies in the range of antennal segments, although there is a small degree of overlap which may sometimes lead to difficulty in identifying isolated specimens. The mesoscutal public entry of *poemyzae* is finer and usually less extensive than in *ninella*, only extending onto the anterior part of the central lobe. There is considerable variation in the emergence dates of *poemyzae*, and it is clear that the species normally has more than one generation a year, in contrast with the true *ninella* which has only a single generation.

The three paratypes from Oxford were referred to *ninella* by NIXON (1945), where the host is recorded as "Agromyza nigripes MEIGEN on reeds". I expressed some doubts about this record in Part I (GRIFFITHS, 1966a, page 575). The specimens concerned have since come to light, and it is clear that the source of the difficulty lies in inaccurate host data, not in NIXON's interpretation of the Braconids. Two of these specimens are mounted with *Poemyza* puparia and only one

with that of an Agromyza species. Evidently Hamm collected a mixed sample of Agromyza albipennis MEIGEN and Cerodontha (Poemyza) incisa MEIGEN, doubtless from Phalaris arundinacea, on which both species occur commonly in England. These produced two species of Dacnusini — Chorebus lugubris (NIXON) (see Part I) and C. poemyzae sp. nov. Clearly we must assume that the latter was bred from the Poemyza, in conformity with the other host data.

# Chorebus vitripennis sp. nov.

Colour. Palpi yellow-brown. Labrum and clypeus black. Centre of mandibles red-brown. Antennae entirely dark. Legs varying from yellow-brown to dark brown, with all coxae black. Gaster entirely dark.

Morphology. Antennal segments: 3, 28-31; 9, (26), 28-30 (29 in the holotype). Palpi long (see the table of biometric data). Mandible similar to that of *asramenes*, with tooth 2 relatively long and pointed but tooth 3 appearing only as a projection from its side. Face strongly punctate.

Thorax somewhat elongate, 1.3-1.5 times as long as high. Sides of pronotum largely sculptured, evenly covered with fine pubescence. Mesoscutum conspicuously roughened anteriorly, densely pubescent on about its anterior third, but thereafter becoming bare, smooth and shining except for some hairs along the course of the notaulices and near the posterior fovea: notaulices distinct to about the middle of the mesoscutum (sometimes continuing as weak impressions almost to the posterior fovea). Precoxal suture strongly developed, continuing almost to the hind margin of the mesepisternum. Metapleural swelling finely rugosepunctate. Metapleural and propodeal pubescence extremely dense and whitish. Petiole widened towards its apex, entirely covered with dense fine pubescence which forms conspicuous matted white apical tufts. Tergite 3 without basal hairs.

Wing (fig. 189) with vein  $R_s$  evenly curved or only weakly sinuate; the wing membrane has a strongly iridescent appearance (hence the specific name).

### Breeding records

#### Host – Cerodontha (Poemyza) superciliosa ZETTERSTEDT

Holotype 9 from puparium 19. ix. 62 on Ammophila arenaria on coastal dunes at Llangennydd, Gower, Wales, em. 17. v. 63 (GCDG). 1 3 paratype from puparium 3. viii. 66 on Ammophila arenaria, Mierzeja Wiślana, Przebrino, Poland, em. 14. viii. 66, leg. NOWAKOWSKI (PAN).

### Other records

Mr. A. W. STELFOX has obtained the following Irish and Scottish material, all from the coast, swept from marram grass (*Ammophila arenaria*).

8 paratypes, Murlough Ho., Co. Down, Ireland, 8. vi-3. vii. 57 (USNM and GCDG). 3 paratypes, Buckroney, Co. Wicklow, Ireland, 5. vi. 49 (2 ex.) and 21. vii. 50 (USNM): 4 paratypes, Killoughter, Co. Wicklow, 11. vi. 50 and 21. vi. 51 (3 ex.) (USNM). 8 paratypes, The Island, Co. Dublin, Ireland, 10. vi-22. vii. 43 (USNM): 1 paratype, Portrane, Co. Dublin, 23. viii. 35 (USNM). 1 J paratype, Bettyhill, Sutherland, Scotland, 2. vii. 54 (USNM). 1 J paratype, Löderup, Skåne, Sweden, 24. vii. 38, leg. D.M.S. and J. F. PERKINS (BM).

The Swedish paratype was provisionally ascribed to C. merion (NIXON) by NIXON (1945). It was also taken on coastal dunes with Ammophila.

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The holotype of this species is the specimen referred to in Part I (Beitr. Ent., 14, page 864) as having emerged from the rear end of the host puparium, contrary to the normal practice of the Dacnusini. Since the second bred specimen has also emerged in this manner, it is possible that this manner of emergence is normal in this species.

I am pleased to give this attractive species the name by which Mr. STELFOX has long known it in his collection. Its dark coloration and partly bare mesoscutum will readily distinguish it from other species associated with *Cerodontha* hosts. It is in many respects similar to *C. merion* (NIXON) and *C. crocale* (NIXON), with which I have compared it below in the key to the *ovalis/lateralis* complex.

### Chorebus asramenes (NIXON), comb. nov.

Dacnusa ovalis MARSHALL sensu NIXON, 1937 (in part) Dacnusa asramenes NIXON, 1945

Colour. Palpi and labrum yellow. Clypeus red-brown. Centre of mandibles yellow-brown or red-brown. Antennae yellow or yellow-brown at their base at least as far as the second flagellar segment. Legs entirely deep yellow, or with the hind coxae somewhat infuscated (more or less brown). Gaster beyond petiole entirely dark or with tergite 3 brownish.

Morphology. Antennal segments: 3, (31), 33-38; 9, 32-35 (bred specimens only). Palpi very long (see the table of biometric data). Mandible (fig. 202) not expanded, with tooth 2 long and pointed but tooth 3 appearing only as a projection from its side. Face weakly punctate.

Thorax elongate, about 1.4 times as long as high. Sides of pronotum evenly covered with fine pubescence. Mesoscutum conspicuously roughened anteriorly, especially on its central lobe, with dense pubescence covering most of its surface (except about the posterior half of the lateral lobes): notaulices weak, but usually extending to about the middle of the mesoscutum (sometimes difficult to distinguish from the surrounding sculpturation). Metapleural swelling strongly rugose-punctate. Metapleural and propodeal pubescence extremely dense and whitish. Petiole slightly widened towards its apex, with dense pubescence covering almost its entire surface and forming well-developed whitish apical tufts (compare fig. 144 and NIXON, 1945, fig. 165). Tergite 3 with few or no basal hairs. Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

Wing with vein  $R_s$  sinuate.

### Breeding records

Host — Cerodontha (Poemyza) pygmaea Meigen

1 φ from puparium 14. viii. 55 on *Brachypodium* sp., Betchworth, Surrey, England, em. 24. viii. 55 (GCDG). 1 δ from puparium 20. vi. 54 on Gramineae sp., Boxhill, Surrey, em. vii. 54, leg. SPENCER (GCDG). 1 δ from puparium 14. x. 62 on *Molinia coerulea*, Oxshott, Surrey, em. 18. iv. 63 (GCDG). 1 ex. from puparium 6. viii. 54 on *Deschampsia caespitosa*, Brookman's Park, Herts., England, em. 17. viii. 54 (BM). 2 ex. from puparia 14. viii. 54 on *Deschampsia caespitosa*, Eynsford, Kent, England, em. 16 and 21. viii. 54 (BM). 1 φ from

puparium 27. vi. 66 on Deschampsia caespitosa, Röderbrunn, Oberlausitz, Saxony, Germany, em. 14. vii. 66, leg. BUHR no. 2902 (GCDG). 1 3 from puparium 3. x. 64 on Festuca gigantea. Stadtwald, Mühlhausen, Thuringia, Germany, em. 16. iii. 65, leg. BUHR no. 2328 (GCDG):  $1 \leq 1 \neq$  from puparia 19. viii. 66 on *Brachypodium silvaticum*, same locality, em. 23-30. viii. 66 and 18. iv. 67, leg. BUHR no. 3114 (GCDG): 2 33 4 99 from puparia 22. vii. 66 on Deschampsia caespitosa, same locality, em. 30. vii-7. viii. 66, leg. BUHR no. 3057 (GCDG). 1 3 from puparium 8. viii. 36 on Molinia coerulea, Graal, Mecklenburg, Germany, em. 12. viii. 36, leg. BUHR (GCDG). 4 99 from puparia 15. vii. 66 on Melica nutans, Calamagrostis arundinacea and Brachypodium silvaticum (2 ex.), Strzałowo reservation, Piska Forest, Poland, em. 25-30. vii. 66, leg. NOWAKOWSKI (PAN). 12 ex. from larvae 9 and 13. vii. 66 on Deschampsia caespitosa, Wierzba, near Popielno, Piska Forest, em. 22. vii-1. viii. 66, leg. Nowakowski (PAN). 1 9 from puparium 15. vii. 64 on Deschampsia caespitosa, Sieraków reservation, Kampinoska Forest, Poland, em. 21. vii. 64, leg. Nowakowski (PAN). 2 ex. from puparia 17 and 26. vi. 64 on Deschampsia caespitosa, Warszawa-Pyry, Kabackie Forest, Poland, em. 30. vi. and 9. vii. 64, leg. NOWAKOWSKI (PAN). 1 9 from puparium 3. x. 65 on Deschampsia caespitosa, Obnoga, near Bukowe Berdo, Bieszczady, Poland, em. 21. iii. 66, leg. Nowakowski (PAN).

Two of the above records were previously published (GRIFFITHS, 1956).

This species and its sister-species *agraules* may be distinguished from other species of *Chorebus* with similar pubescence of the petiole by their elongate thorax, extensive mesoscutal pubescence, pale basal antennal segments and the form of the mandibles.

#### Chorebus agraules (NIXON), comb. nov.

Dacnusa agraules NIXON, 1945

Very similar to asramenes, with which it may be compared as follows.

Colour. Hind coxae brownish.

Morphology. 32-35 antennal segments ( $\mathcal{Q}$ ). Ovipositor ( $\mathcal{Q}$ ) much longer, projecting beyond the apical tergite in the retracted position by over half the length of the petiole.

Breeding records

Host – Cerodontha (Poemyza) muscina Meigen

 $1\ \circ$  from puparium on Festuca ovina, Jena, Thuringia, Germany, em. vii. 64, leg. Buhr, Hering no. 2084 (GCDG).

Apart from the much longer ovipositor I can see no significant difference between this species and *asramenes*. I consider that these are sister-species (synapomorph in respect of the elongate thorax and the form of the mandibles).

# Chorebus difficilis sp. nov.

Colour. Palpi and labrum yellow or ochreous yellow. Clypeus dark. Centre of mandibles yellow-brown. Antennae more or less entirely dark, at most becoming obscurely brown towards their base. Legs largely ochreous yellow or yellow-brown, with the hind coxae infuscated (more or less dark brown). Gaster more or less entirely dark. Morphology. Antennal segments:  $\mathcal{J}, 29-32; \mathcal{Q}, (27)-28-31$ . Palpi long (see the table of biometric data). Mandible hardly expanded, with tooth 3 relatively small (tooth 2 is usually not so long and pointed as in *asramenes* and *vitripennis*). Face weakly punctate.

Thorax about 1.3 times as long as high. Sides of pronotum bare and smooth centrally, with fine pubescence mainly along and below the oblique suture, becoming strongly shining (virtually bare or at most with very fine inconspicuous pubescence) above the suture. Mesoscutum extensively roughened (especially anteriorly), with dense pubescence covering almost its entire surface: notaulices usually reaching about the middle of the mesoscutum, but somewhat difficult to distinguish from the surrounding sculpturation. Metapleural swelling strongly rugose-punctate. Metapleural and propodeal pubescence very dense and whitish. Petiole slightly widened towards its apex, evenly covered with dense pubescence which tends to become denser towards the apical corners (forming distinct apical tufts, although these are sometimes weak). Tergite 3 with a little pubescence at its base (adjacent to the petiole).

Wing with vein  $R_s$  sinuate (compare *alecto*, fig. 88).

#### Host 1 – Cerodontha (Poemyza) pygmaea Meigen

Holotype 9; 1 & paratype from puparia 14. x. 62 on Dactylis glomerata, Oxshott, Surrey, England, em. 12 and 15. iv. 63 (GCDG). 1 9 paratype, Stuttgart (Feuerbach Tal), Germany, em. 29. iii. 54, leg. GROSCHKE (STGT). 1 & paratypes, Stuttgart-Kräherwald, em. 21. vii. 54, leg. GROSCHKE (STGT). 19 paratype from puparium 5. viii. 65 on Dactylis aschersoniana, Stadtwald, Mühlhausen, Thuringia, Germany, em. 20. iii. 66, leg. BUHR no. 2576 (GCDG): 1 9 paratype from puparium 29. vii. 66, same plant and locality, em. 14. iv. 67, leg. BUHR no. 3073b (GCDG): 1 3 paratype from puparium 29. vii. 66 on Brachypodium silvaticum, same locality, em. 11-14. viii. 66, leg. BUHR no. 3074 (GCDG). 9 paratypes from larvae 26. vi. 64 on Dactylis glomerata, Warszawa-Pyry, Kabackie Forest, Poland, em. 31. vii. 64, 17. viii. 64 and 5-12. v. 65, leg. Nowakowski (PAN).  $1 \Leftrightarrow$  paratype from larvae 6. vii. 64 on Dactylis glomerata, Warszawa-Bielany, em. 15. v. 65, leg. NOWAKOWSKI (PAN): 1 & paratype from puparium 30. vi. 66, same plant and locality, em. 22. vii. 66, leg. NOWAKOWSKI (PAN). 2 33 paratypes from larvae 3. vii. 56 on Dactylis glomerata, Warszawa-Młociny, em. 28. vii and 10. viii. 56, leg. Nowakowski (PAN): 1 paratype from larva 11. vi. 64, same plant and locality, em. 17. viii. 64, leg. Nowakowski (PAN). 1 3 paratype from larva 25. vii. 65 on Dactylis glomerata, Żukowo, distr. Kartuzy, Poland, em. 8. v. 65, leg. Nowakowski (PAN). 1 3 paratype from puparium 28. viii. 65 on Dactylis glomerata, Cisna, Bieszczady, Poland, em. 18. iii. 66, leg. NOWAKOWSKI (PAN).

#### Host 2 – Cerodontha (Poemyza) incisa MEIGEN

1 δ paratype from puparium 14. viii. 65 on *Briza media*, Łopiennik, Bieszczady, Poland, em. xii. 65, leg. Nowakowski (PAN). 1 δ paratype from puparium 8. x. 65 on *Phleum* pratense, Hulskie, Otryt, Poland, em. xii. 65, leg. Nowakowski (PAN).

### Host 3 — Cerodontha (Poemyza) calamagrostidis Nowakowski

1 δ paratype from larva 8. vii. 66 on *Calamagrostis canescens*, Röderbrunn, Oberlausitz, Saxony, Germany, em. 13-26. iv. 67, leg. BUHR no. 3010b (GCDG).

### Host 4 – Cerodontha (Poemyza) deschampsiae Spencer

2 33, 4 99 paratypes from larvae 8. vii. 66 on *Calamagrostis canescens*, Röderbrunn, Oberlausitz, Saxony, Germany, em. 1. viii. 66 (1 9) and 10-18. iv. 67, leg. BUHR nos. 3011 and 3012b (GCDG).

### Host 5 — Cerodontha (Dizygomyza) chaixiana GROSCHKE

4 paratypes from puparia on *Poa chaixii*, Stuttgart (Kräherwald and Wildpark), Germany, em. 8 and 20. iv. 54, leg. GROSCHKE (STGT).

This species is difficult to distinguish from certain species associated with *Phytomyza* hosts, particularly *C. armida* (NIXON) and *C. luzulae* GRIFFITHS (see couplets 91-93 in the key to the *ovalis/lateralis* complex below). It may be separated from other species of *Cerodontha*-parasites with apical tufts of pubescence on the petiole by its relatively dark coloration, the length of the thorax (fairly elongate, but not as much so as in *asramenes* and *agraules*) and the extensive mesoscutal pubescence. The pubescence of the petiole seems identical with that of *C. alecto* (MORLEY) and *C. armida* (NIXON) (compare fig. 144), the apical tufts not being so well developed as for instance in *asramenes* and *ninella*.

# Plesiomorph species of Chorebus related to the affinis and senilis groups

There are six species of Chorebus associated with Cerodontha hosts<sup>2</sup> which retain a well-developed vein  $Cu_{Ib}$ , a plesiomorph feature found also in the affinis and senilis groups, but lack the hind coxal tuft of pubescence characteristic of these groups. The species concerned are given below in key 2 in the section headed "Keys to Chorebus HALIDAY". Three of these species were included in NIXON'S (1944) concept of the merella-group, but the fourth species which he included in that group, C. alua (NIXON), is clearly referable to the affinis group as defined in this paper. There seems no firm evidence for considering these six species to represent a monophyletic group on the basis of the characteristic rosette of metapleural pubescence which occurs in several groups of Chorebus, the only clearly apomorph feature found in all these six species is the very elongate petiole. It is difficult to interpret resemblances in the shape of the petiole among Chorebus species, as similar conditions seem to have arisen frequently through convergence.

I consider these plesiomorph species as probably more closely related to the *affinis* and *senilis* groups than to the *ovalis/lateralis* complex, because there is a distinct tendency in most of them for the eyes to be more strongly convergent below in the female, as has been noted in many species of the *affinis* group. As far as I am aware there is no such tendency towards sexual dimorphism in any species of the *ovalis/lateralis* complex.<sup>3</sup>

<sup>&</sup>lt;sup>2</sup> Subject to confirmation in the case of *orissa*, whose life-history is not yet known.

<sup>&</sup>lt;sup>3</sup> In two of these species the antennal pubescence is also strongly sexually dimorphic, a feature which occurs in several species of the *affinis* group and one of the *senilis* group (i.e. *C. bathyzonus* (MARSHALL)). While clearly all the species in which this antennal dimorphism is well developed cannot be regarded as monophyletic, it is possible that the convergent development of this character in certain species may indicate some similarity in the genetic make-up of the groups concerned.

One of the species treated below, C. transversus (NIXON), is remarkable in retaining 3-toothed mandibles comparable with those of other genera of Dacnusini, such as *Exotela* and *Dacnusa* (see fig. 203). This condition should not be confused with the long and pointed 3-toothed mandibles found in certain species of the affinis group, which have clearly been derived secondarily from 4-toothed mandibles similar to those of other species of this group (compare figs. 204-207). If the 3-toothed mandibles of transversus are genuinely plesiomorph, the characters of this species are of considerable interest for the interpretation of the evolution of the species included in Chorebus, because in this case the rosette of metapleural pubescence has been evolved before 4-toothed mandibles. Since there are a number of Chorebus spp. (included in key 4 in the section below headed "Keys to Chorebus HALIDAY") in which the contrary combination occurs (relatively plesiomorph metapleural pubescence, but strongly 4-toothed mandibles), it must be concluded that some convergence has occurred in the evolution of one or both of the apomorph conditions. Some revision of the relationships between groups of Chorebus spp. suggested in Part I (GRIFFITHS, 1964, page 894) is undoubtedly necessary, but it seems premature to attempt to revise the analysis at the moment, as information on variation in the larval morphology is expected shortly as a result of research by Dr. M. ČAPEK.

In the descriptions which follow the following characters may be assumed, wherever nothing to the contrary is stated.

Clypeus dark, like the face. Face weakly punctate or almost smooth, covered with dense fine pubescence which is directed inwards or upwards over its centre, but downwards along the eye-margins. Back of head largely bare, with only some fine pubescence near the base of the mandibles. Sides of pronotum largely bare and shining except for some fine inconspicuous pubescence below the oblique suture. Pubescence at the base of the hind coxa not forming a distinct matted tuft. Metapleural pubescence very dense (compare figs. 22 and 23 in Part I), forming a rosette around the strongly rugose swelling: propodeum strongly rugose, covered with similar dense pubescence. Petiole elongate. Tergite 3 without basal pubescence. Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

Wing (figs. 190-192) with vein  $Cu_{1b}$  well developed, forming a distinct angle with the transverse section of  $Cu_1$ .

### Chorebus transversus (NIXON), comb. nov.

### Dacnusa transversa NIXON, 1954

Colour. Palpi and labrum yellow. Centre of mandibles yellow-brown. Antennae with their basal segments yellow-brown (about as far as the second flagellar segment), this colour merging gradually into the darker colour of the more distal flagellar segments. Legs uniformly deep yellow, with only tarsal segments 5 distinctly infuscated. Gaster entirely dark. Morphology. Head very strongly transverse. Antennal segments: 3, (33) – 34-36; 9, (28) – 30-36: flagellum with its more apical segments about  $2\frac{1}{2}$  times as long as wide: basal flagellar segments evenly covered with rather long pubescence in both sexes. Palpi very long (see the table of biometric data). Mandible (fig. 203) 3-toothed (without any trace of an additional tooth on the side of tooth 2). Eyes somewhat convergent below in the female (minimum distance between them less than half the head width), but hardly so in the male.

Thorax short, 1.1-1.3 times as long as high. The lower half of the sides of the pronotum is distinctly sculptured and covered with dense pubescence, although this is fine and not very conspicuous. Mesoscutum with its anterior face and central lobe slightly roughened, with dense pubescence covering its anterior face and central lobe and often extending onto the anterior half of the lateral lobes, although sometimes these are almost completely bare: notaulices usually well developed, extending as rugose furrows at least to the middle of the mesoscutum. Precoxal suture broad and conspicuously rugose-costate anteriorly, but becoming weak posteriorly (either fading out or reaching the hind margin of the mesepisternum only as a narrow smooth impression). Petiole very elongate, virtually parallel-sided in the male, but usually somewhat widened towards its apex in the female, almost bare.

Wing (fig. 190) with cell  $2R_1$  long and vein  $R_s$  strongly sinuate (as in many species of the *lateralis* group): 1m-cu only narrowly rejected from cell $R_s$ , sometimes almost interstitial.

# Breeding records

Host — Cerodontha (Butomomyza) angulata LOEW (= semiposticata HENDEL)

1 3 from larva 20. xi. 55 on Carex pendula, Scratch Wood, London, em. 25. v. 56, leg. SPEN-CER (GCDG). 19 from larva 27. ix. 61 on Carex sp., Bookham, Surrey, England, em. 22. v. 62 (GCDG). 1 9 from larva 12. viii. 62 on Carex sp., Hadley Wood, Herts., England, em. 4. ix. 62 (GCDG). 2 33, 1 9 from larvae 30. ix. 64 on Carex silvatica, Clapham, Yorks., England, em. 9-16. v. 65 (GCDG). 1 3 from larva on Carex sp., Stuttgart-Wildpark, Germany, em. 20. iii. 54, leg. GROSCHKE (STGT). 1 3, 3 99 from larvae on Carex sp., Stuttgart-Kräherwald, em. 3. ix. 54, leg. GROSCHKE (STGT). 18 ex. from larvae 26. vi. 54 on Carex hirta, Warszawa-Pyry, Kabackie Forest, Poland, em. 21. vii-22. viii. 64, leg. Nowa-KOWSKI (PAN and GCDG): 2 99 from larvae 5. xi. 65, same plant and locality, em. 25. iii. 66, leg. NOWAKOWSKI (PAN). 3 ex. from larvae 20. vi. 64 on Carex hirta, Warszawa-Powsin, Kabackie Forest, em. 18-21. vii. 64, leg. NOWAKOWSKI (PAN). 8 ex. from larvae 6. vii. 64 on Carex hirta, Warszawa-Bielany, em. 29. vii–20. viii. 64, leg. Nowakowski (PAN). 5 99 from larvae 1. vii. 61 on Carex hirta, Międzylesie, near Warszawa, em. 3. viii. 61, leg. Nowa-KOWSKI (PAN). 19 from larva 18. x. 64 on Carex hirta, Łazy, distr. Piaseczno, Poland, em. 27. v. 65, leg. Nowakowski (PAN). 13 ex. from larvae 18. vi. 66 on Carex hirta, Sieraków reservation, Kampinoska Forest, Poland, em. 14-19. vii. 66, leg. NOWAKOWSKI (PAN). 10 ex. from larvae 11. vii. 66 on Carex hirta, Iznota, Piska Forest, Poland, em. 3-11. viii. 66, leg. Nowakowski (PAN). 7 99 from larvae 15. vii. 66 on Carex hirta, Strzałowo reservation, Piska Forest, em. 5-19. viii. 66, leg. Nowakowski (PAN). 4 99 from larvae 5. vii. 66 on Carex sp., Wierzba, near Popielno, Piska Forest, em. 27-29. vii. 66, leg. NOWAKOWSKI (PAN). 1 ex. from larva 20. viii. 65 on Carex hirta, Smerek, Bieszczady, Poland, em. 15. ix. 65, leg. Nowakowski (PAN). 2 ex. from larvae 28. vii. 64 on Carex pseudocyperus, Mierzeja Wiślana, Krynica Morska, Poland, em. 25. viii and 8. ix. 64, leg. NOWAKOWSKI (PAN): 1 Q from larva 24. vi. 66, same plant and locality, em. 30. viii. 66, leg. Nowakowski (PAN):

4 qq from larvae 24. vi. 66 on *Carex vulpina*, same locality, em. 23. vii-1. viii. 66, leg. Nowakowski (PAN). 1 q from larva 30. vii. 64 on *Carex acutiformis*, Mirachowo, distr. Kartuzy, Poland, em. 9. ix. 64, leg. Nowakowski (PAN).

I have already commented above on the apparently plesiomorph 3-toothed mandibles of this species. This character will readily serve to separate it from C. humeralis sp. nov., which occurs on the same host. The wing venation of these species also differs, since in *transversus* cell  $2R_I$  is longer and vein  $R_s$  more strongly sinuate.

# Chorebus humeralis sp. nov.

Colour. Palpi golden yellow. Labrum yellow or yellow-brown. Centre of mandibles brown. Antennae largely black, but becoming brown or reddish at their base (as far as about the second flagellar segment). Legs largely golden yellow, with the tarsi somewhat infuscated towards their apex (especially segments 5). Gaster entirely dark.

Morphology. Head strongly transverse. Antennal segments: 3, (32)-33-36;  $\varphi, (30)-32-35$ : flagellum short, with its more apical segments only about twice as long as wide: basal flagellar segments densely covered with rather short pubescence in both sexes. Palpi of moderate length (see the table of biometric data). Mandible not expanded, 4-toothed with tooth 3 well developed as a projection from the side of tooth 2. Eyes only slightly convergent below in the female. Face strongly shining, only sparsely pubescent and almost smooth centrally.

Thorax short, 1.2-1.3 times as long as high. Mesoscutum with its anterior face roughened but its dorsal surface almost smooth (with only faint scaly-reticulate ground sculpture), with fine pubescence covering its anterior face and often extending sparsely onto the central lobe (although sometimes this is virtually bare, like the lateral lobes): notaulices complete, reaching the posterior fovea as deep rugose furrows, their lateral extensions (i.e. at the "shoulders") margined by a strongly raised crest: usually a distinct groove also runs along the centre-line of the mesoscutum. Precoxal suture strongly rugose-costate almost throughout, broad anteriorly, extending almost to the hind margin of the mesepisternum. Petiole usually slightly widened towards its apex, almost bare (with only a few fine isolated hairs on its dorsal surface).

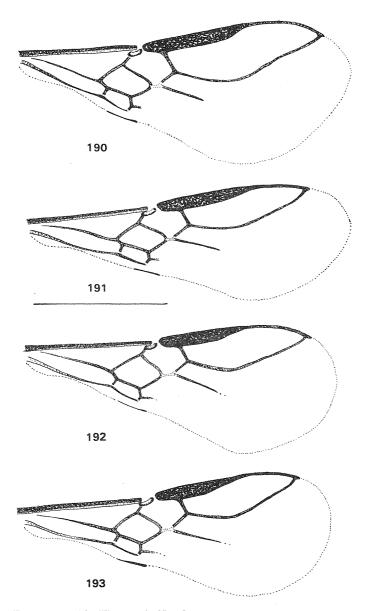
Wing (fig. 191) with cell  $2R_I$  rather narrow and elongate; vein  $R_s$  only slightly curved; Im-cu rejected from cell  $R_s$  (but sometimes only narrowly so).

#### Breeding records

Host — Cerodontha (Butomomyza) angulata LOEW (= semiposticata HENDEL) 28 33 45 99 paratypes, Rudehegn, 1 9 paratype, Køge, and 1 3 paratype, Damhusmose; Sealand, Denmark, leg. SCHLICK (KB and GCDG).

#### Other material

1  $\Im$  holotype, Newtownbabe, Co. Louth, Ireland, 30. vi. 62, leg. STELFOX (USNM). 1  $\Im$ , 1  $\Im$  paratypes, Killoughter, Co. Wicklow, Ireland, 15. viii. 51, leg. STELFOX (USNM): 1  $\Im$  paratype, same locality, 11. vi. 53, leg. STELFOX (USNM). 2  $\Im$  paratypes, The Murrough, Co.



Figs. 190–193. Wings of Chorebus spp.: 190, C. transversus (NIXON)  $\varphi$ ; 191, C. humeralis sp. nov.  $\varphi$ ; 192, C. merella (NIXON)  $\varphi$ ; 193, C. cytherea (NIXON)  $\varphi$ . (Scale 1 mm.)

Wicklow, 28. viii. 54, leg. STELFOX (USNM). 1 9 paratype, Glenasmole, Co. Dublin, Ireland, 2. vii. 42, leg. STELFOX (USNM).

This species is separable from *merella* and *nobilis* by its shorter thorax and tarsi. It has a superficial resemblance to C. glabriculus (THOMSON), included in the *ovalis/lateralis* complex, because of its coloration and the well developed notaulices and precoxal suture. However it has longer, entirely yellow palpi and a closed cell 2Cu: in glabriculus this cell is clearly open at its lower distal corner. A second species, C. transversus (NIXON), attacks the same host as humeralis, but is readily separable, as already mentioned above, by the form of its mandibles.

I am pleased to give this species the name by which Mr. A. W. STELFOX has long known it in his collection.

# Chorebus merella (NIXON), comb. no v.

Dacnusa merella NIXON, 1937 and 1944

Colour. Palpi and labrum clear yellow. Centre of mandibles yellow-brown. Antennae with their basal segments conspicuously yellowish (or rarely only brown) at least as far as the third flagellar segment, this colour merging gradually into the darker colour of the more distal flagellar segments. Legs almost entirely clear yellow, usually with only tarsal segments 5 somewhat infuscated. Gaster beyond petiole more or less entirely dark, or at most with tergites 3 and 4 yellow-brown or reddish.

Morphology. Antennal segments: 3, (36)-37-42-(43); 9, (33)-35-40: flagellum extremely long, with nearly all its segments at least  $2\frac{1}{2}-3$  times as long as wide: basal flagellar segments evenly covered with rather long, dense pubescence in both sexes. Palpi very long (see the table of biometric data). Mandible 4-toothed (fig. 25), with tooth 2 rather long and ponted and tooth 3 visible as a projection from its side. Eyes usually distinctly convergent below in the female, but hardly so in the male.

Thorax elongate, 1.3—1.4 times as long as high. Mesoscutum with its anterior face and about the anterior half of its central lobe roughened, with dense pubescence covering its anterior face and at least the anterior half of the central lobe but the lateral lobes almost bare: notaulices usually distinct as slightly sculptured furrows at least to the middle of the mesoscutum, sometimes reaching the posterior fovea, their rugose lateral extensions often margined by a slightly raised crest. Precoxal suture broad and conspicuously rugose-costate anteriorly, continuing as a narrow groove to the hind margin of the mesepisternum. Petiole virtually parallel-sided, with its dorsal surface almost bare (at most with a little pubescence near its base and a row of hairs along each side).

Wing (fig. 192) with vein  $R_s$  slightly sinuate and 1m-cu rejected from cell  $R_s$  (though sometimes only narrowly so).

### Breeding records

Host 1 - Cerodontha (Dizygomyza) caricicola HERING (= soenderupi HERING)

1 ex. from puparium 14. viii. 54 on *Carex* sp., Eynsford, Kent, England, em. 16. viii. 54 (BM).  $2 \sigma \sigma$ ,  $4 \circ \circ$  from puparia 14. ii. 54 on *Carex pendula*, Scratch Wood, London, em. 7. iv-v. 54, leg. SPENCER (GCDG): 3 ex. from puparia 25. vii. 54, same plant and locality,

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em. 4-11. viii. 54 (BM): 3 ex. from puparia 22. viii. 54 on Carex pendula and Carex sp., same locality, em. 23-27. viii. 54 (BM). 1 J. 2 99 from puparia mid-vii. 53 on Carex sp., Schongau, Oberbayern, Germany, em. 20-26. vii. 53, leg. GROSCHKE (STGT).  $1 \circ$  from puparium on Carex sp., Stuttgart-Kräherwald, Germany, em. 25. ii. 54, leg. GROSCHKE (STGT). 5 ex. from puparia 15. viii. 66 on Carex silvatica, Suchy Gron, Babia Góra, Poland, em. 20. viii-2. ix. 66, leg. Nowakowski (PAN). 6 ex. from puparia 17. viii. 66 on Carex silvatica, Mata Babia Góra, near Marków pond, em. 24. viii-14. ix. 66, leg. Nowakowski (PAN). 7 ex. from larvae and puparia 15. ix. 66 on Carex silvatica, Kazalnica, near Sarnia Skała, Tatry, Poland, em. 24. x. 66 (1 3) and 5. ii-11. iii. 67, leg. Nowakowski (PAN). 39 from puparia 15. x. 66 on Carex hirta and C. silvatica, Staników Żleb, near Hruby Regiel, Tatry, em. 6 and 11. ii. 67, leg. Nowakowski (PAN). 1 9 from puparium 21. viii. 57 on Carex silvatica, Spadowiec Valley, Tatry, em. 7. ix. 57, leg. NOWAKOWSKI (PAN). 1 & from puparium 9. vii. 66 on Carex sp., Wierzba, near Popielno, Piska Forest, Poland, em. 18. vii. 66, leg. NOWAKOWSKI (PAN). 1 9 from puparium 8. x. 65 on Carex silvatica, Otryt-Hulskie, Poland, em. 14. iii. 66, leg. Nowakowski (PAN). 1 ex. from puparium 13. viii. 65 on Carex silvatica, Cisna, Bieszczady, Poland, em. 20. viii. 65, leg. Nowakowski (PAN). 1 9 from puparium 17. viii. 65 on Carex pilosa, Falowa, Bieszczady, em. 31. viii. 65, leg. Nowa-KOWSKI (PAN).

### Host 2 — Cerodontha (Dizygomyza) luzulae GBOSCHKE

1 3, 2 99 from puparia on Luzula silvatica, Stuttgart-Kaltental, Germany, em. 1–23. vi. 55, leg. GROSCHKE (STGT). 2 33, 3 99 from larvae 6. x. 65 on Luzula silvatica, Górna Solinka reservation, Bieszczady, Poland, em. 18–23. iii. 66, leg. Nowakowski (PAN). 1 9 from larva 3. x. 65 on Luzula silvatica, Obnoga, near Bukowe Berdo, Bieszczady, em. 4. xi. 65, leg. Nowakowski (PAN). 1 9 from larva 22. ix. 66 on Luzula silvatica, near Smreczyński pond, Tatry, Poland, em. 17. iv. 67, leg. Nowakowski (PAN). 1 9 from larva 22. ix. 66 on Luzula silvatica, Kraków ravine, Tatry, em. 2. iii. 67, leg. Nowakowski (PAN).

### Host 3 — Cerodontha (Dizygomyza) chaixiana GROSCHKE

 $1 \circ$  from puparium on *Poa chaixii*, Kräherwald, Stuttgart, Germany, em. 25. ii. 54, leg. GROSCHKE (STGT). 1  $\sigma$  from puparium 19. viii. 65 on *Poa chaixii*, Łopiennik, Bieszczady, Poland, em. 26. viii. 65, leg. NOWAKOWSKI (PAN).

# Host 4 — Cerodontha (Butomomyza) staryi HERING

 $5 \sigma \sigma$ , 5 QQ from larvae and puparia 5. xii. 53 on *Carex* sp. cf. *remota*, Stuttgart (Feuerbach Tal), Germany, em. 8. i-29. iii. 54, leg. GROSCHKE (STGT) (all except one labelled by GROSCHKE as bred from "*Phytobia stilettata*", a name which he did not publish).

# Host 5 – Cerodontha (Butomomyza) scirpi KARL (= scutellaris auctt.)

1 3 from puparium 7. viii. 55 on *Scirpus silvaticus*, Godalming, Surrey, England, em. iv. 56, leg. SPENCER (GCDG). 1 9 from larva 27. vi. 66 on *Scirpus silvaticus*, Röderbrunn, Saxony, Germany, em. 21. vii. 66, leg. BUHR no. 2898 (GCDG). 3 33, 6 99, Stuttgart-Wildpark, Germany, em. 15. ii - 20. iv. 54, leg. GROSCHKE (STGT). 2 ex. from larvae 25. vii. 64 on *Scirpus silvaticus*, Żukowo, distr. Kartuzy, Poland, em. 17. viii. 64 and 15. v. 65, leg. Nowakowski (PAN).

### Host 6 — Cerodontha (Butomomyza) caricivora GROSCHKE

1 δ from puparium on *Carex silvatica*, Schöngeising an der Amper, Oberbayern, Germany, em. 2. vi. 53, leg. GROSCHKE (STGT). 1 ex. from larva 19. viii. 65 on *Carex silvatica*, Rożki, Bieszczady, Poland, em. 12. x. 65, leg. NOWAKOWSKI (PAN). 2 ex. from larvae 8. x. 65 on *Carex silvatica*, Hulskie, Otryt, Poland, em. 14. iii. 66, leg. NOWAKOWSKI (PAN).

A few of the above records for C. (D.) caricicola HERING have been previously published in GRIFFITHS (1956).

The differences between *merella* and other similar species are given in key 2 in the section below headed "Keys to *Chorebus* HALIDAY". It is important to note that in *merella* the central lobe of the mesoscutum is densely pubescent and roughened anteriorly.

### Chorebus nobilis sp. nov.

Similar to C. merella (NIXON), with which it may be compared as follows.

Colour. Centre of mandibles brown or red-brown. Antennae usually yellowbrown or brown at their base as far as about the first flagellar segment, but sometimes more or less entirely dark.

Morphology. Antennal segments: 3, 39-45;  $\bigcirc$ , 38-43: flagellum not so long proportionately, with its more apical segments only  $2-2\frac{1}{2}$  times as long as wide: basal flagellar segments in both sexes evenly covered with short dense pubescence. Mandible slightly broader. Eyes hardly convergent below in either sex.

Mesoscutum with only its anterior face roughened (the central lobe being virtually smooth like the lateral lobes), with dense pubescence covering its anterior face and usually extending sparsely onto the central lobe, but sometimes this too is bare like the lateral lobes (in which case there remain on the dorsal surface of the mesoscutum only a few hairs along the course of the notaulices and near the posterior fovea): notaulices always complete, reaching the posterior fovea as deep conspicuous furrows which are usually rugose throughout (at most becoming smooth posteriorly only), their rugose lateral extensions (i.e. at the "shoulders") margined by a strongly raised crest. Precoxal suture sculptured throughout its length, very broad and irregularly rugose-costate anteriorly, becoming narrower towards the hind margin of the mesepisternum: the central part of the mesepisternum (above the precoxal suture) is covered with fine scaly-reticulate ground sculpture, not completely smooth as in *merella* and *pelion*: the mesosternum (below the precoxal suture) also appears faintly sculptured. Propodeal pubescence slightly shorter and less dense.

Size usually larger (wing length up to 3 mm.).

### Breeding records

# Host — Cerodontha (Butomomyza) eucaricis Nowakowski

Holotype  $\Im$ ; 6  $\Im$ , 6  $\Im$  paratypes from puparia on *Carex* sp., Darmstadt-Kranichstein, Hessen, Germany, em. 15. xi. 52 (8 ex., including the holotype) and 14. ii-25. iii. 53 (5 ex.), leg. GROSCHKE (STGT and GCDG). 1  $\Im$ , 1  $\Im$  paratypes from larvae 3. vii. 56 on *Carex* sp., Warszawa-Młociny, Poland, em. 2 and 10. viii. 56, leg. NOWAKOWSKI (PAN). 1  $\Im$  paratype from puparium 31. vii. 55 on *Carex* sp., Granica, Kampinoska Forest, Poland, em. 9. viii. 55, leg. NOWAKOWSKI (PAN). 1 paratype (? sex) from larva 10. vii. 64 on *Carex vesicaria*, Sieraków, Kampinoska Forest, em. 3. viii. 64, leg. NOWAKOWSKI (PAN). 1 paratype (? sex) from puparium 7. vii. 64 on *Carex hirta*, Warszawa-Pyry, Kabackie Forest, Poland, em. 21. vii. 64, leg. NOWAKOWSKI (PAN).

#### Other material

1 & paratype, Cashegal, Co. Sligo, Ireland, 27. vi. 36, leg. STELFOX (USNM).

The host of GROSCHKE's series was originally identified as C. (B.) caricivora GROSCHKE, but is probably referable to the subsequently described C. (B.) eucaricis NOWAKOWSKI.

There is some overlap between this species and *merella* in size and the number of antennal segments. The most reliable diagnostic criteria are the presence in *nobilis* of complete notaulices and scaly-reticulate ground sculpture on the central part of the mesepisternum: also the central lobe of the mesoscutum is not roughened as in *merella*.

# Chorebus pelion (NIXON), comb. nov.

# Dacnusa pelion NIXON, 1944

Similar to C. merella (NIXON), with which it may be compared as follows.

Colour. Antennae usually only yellow-brown or red-brown at their base about as far as the first flagellar segment, rarely so brightly coloured as in *merella*: in some specimens the flagellum is entirely dark.

Morphology. Antennal segments:  $3, 31-35; \mathfrak{Q}, 30-35$ : flagellar pubescence sexually dimorphic, in the female the entire flagellum is evenly covered with rather long, dense pubescence (as in *merella*), but in the male the segments of about the basal half of the flagellum are strongly shining and almost bare, bearing only a few isolated short hairs (mainly near their apex). Mandible slightly less elongate than normally in *merella*.

Mesoscutum with only its anterior face partly roughened (the central lobe being completely smooth like the lateral lobes), with its pubescence confined to its anterior face except for a few fine hairs along the course of the notaulices and near the posterior fovea: notaulices with their longitudinal extensions completely smooth (although usually well-defined anteriorly and reaching the posterior fovea as distinct impressions), their lateral extensions (i.e. at the "shoulders") either smooth or rugose, margined by a strongly raised crest. Precoxal suture narrow, becoming only slightly broader anteriorly, but well-defined, regularly rugose-costate throughout its length (almost as far as the hind margin of the mesepisternum): both the mesepisternum and mesosternum (i.e. both above and below the precoxal suture) are completely smooth. Petiole slightly broader (see the table of biometric data).

### Breeding records

### Host 1 — Cerodontha (Butomomyza) eucaricis NowAKowski

5 33, 24 99 from puparia on *Carex* sp., Darmstadt-Kranichstein, Hessen, Germany, em. 25. i-10. v. 53, leg. GROSCHKE (STGT and GCDG) (host originally identified as *C. (B.) caricivora* GROSCHKE, but more probably referable to *eucaricis*). 4 33, 2 99 from larvae 24. vii. 56 on *Carex hirta*, Cybulice, Kampinoska Forest, Poland, em. 19. viii. 56 (1 9) and 21. iv-23. v. 57, leg. NOWAKOWSKI (PAN and GCDG). 5 ex. from larvae 12. vii. 64 on *Carex pseudocyperus*, Dziekanów Leśny, Kampinoska Forest, em. 3-15. viii. 64, leg. NOWA-KOWSKI (PAN). 1 9 from larva 10. vii. 64 on *Carex pseudocyperus*, Sieraków, Kampinoska Forest, em. 3. viii. 64, leg. NOWAKOWSKI (PAN). 1 3, 2 99 from larvae 6. vii. 64 on *Carex hirta*, Warszawa-Bielany, Poland, em. 31. vii-7. viii. 64, leg. NOWAKOWSKI (PAN). 1 3, 2 99 from puparia 7. vii. 64 on *Carex hirta*, Warszawa-Pyry, Kabackie Forest, em. 23. vii-12.

viii. 64, leg. Nowakowski (PAN): 1 3 1 9 from puparia 5. xi. 65, same plant and locality, em. 21. iii. 66, leg. Nowakowski (PAN). 7 99 from larvae 30. vii. 64 on Carex hirta, Mirachowo, distr. Kartuzy, Poland, em. 29-31. viii. 64 (3 ex.) and 19. v-vi. 65, leg. Nowa-KOWSKI (PAN). 2 99 from larvae 25. vii. 64 on Carex hirla, Zukowo, distr. Kartuzy, em. 20. v and 5. vi. 65, leg. Nowakowski (PAN). 4 99 from larvae 7. ix. 65 on Carex hirta, Nowy Most, near river Krutynia, Piska Forest, Poland, em. 4. xii. 65 (1 ex.) and 28-30. iii. 66, leg. NOWAKOWSKI (PAN). 1 3 from larva 6. xi. 65 on Carex hirta, Pranie near Nidzkie lake, Piska Forest, em. 21. iii. 66, leg. NOWAKOWSKI (PAN). 1 of from puparium 9. vii. 66 on Carex hirta, Wierzba, near Popielno, Piska Forest, em. 11. viii. 66, leg. Nowakowski (PAN): 1 & from larva 13. vii. 66 on Carex sp., same locality, em. 1. viii. 66, leg. NOWAKOWSKI (PAN). 2 99 from larvae 11. vii. 66 on Carex hirta, Iznota, Piska Forest, em. 11-14. viii. 66, leg. NOWAKOWSKI (PAN). 3 99 from larvae 15. vii. 66 on Carex hirta, Strzałowo reservation, Piska Forest, em. 14, 24. viii. 66 and 21. iv. 67, leg. NOWAKOWSKI (PAN). 1 9 from larva 23. vii. 64 on Carex hirta, Górki Wschodnie, near Gdansk, Poland, em. 29. viii. 64, leg. NOWAKOWSKI (PAN). 1 3 1 9 from puparia 28. vii. 64 on Carex pseudocyperus, Mierzeja Wiślana, Krynica Morska, Poland, em. 15 and 25. v. 65, leg. Nowakowski (PAN): 2 dd from larvae 4. viii. 66 on Carex vulpina, same locality, em. 14. iv. 67, leg. NOWAKOWSKI (PAN): 1 3 from puparium 28. vii. 64 on Carex pseudocyperus, same locality, em. 3. viii. 64, leg. NOWAKOWSKI (PAN).

### Host 2 – Cerodontha (Butomomyza) vigneae Nowakowski

1 δ from puparium 4. viii. 66 on *Carex vulpina*, Mierzeja Wiślana, Krynica Morska, Poland, em. 12. viii. 66, leg. Νοwakowski (PAN).

### Host 3 — Cerodontha (Dizygomyza) morosa MEIGEN

1 δ from larva 24. vi. 66 on *Carex vulpina*, Mierzeja Wiślana, Krynica Morska, Poland, em. 22. vii. 66, leg. Νοwakowski (PAN).

### Host 4 — Cerodontha (Dizygomyza) sp. probably caricicola HERING

1 φ from puparium 7. vii. 64 on Carex hirta, Warszawa-Pyry, Kabackie Forest, Poland, em. 23. vii. 64, leg. Nowakowski (PAN). 1 φ from puparium 12. vii. 64 on Carex pseudocyperus, Dziekanów Leśny, Kampinoska Forest, Poland, em. 27. vii. 64, leg. Nowakowski (PAN). 1 φ from puparium 27. vii. 55 on Carex pseudocyperus, Granica, Kampinoska Forest, em. 4. viii. 55, leg. Nowakowski (PAN). 1 ♂ from puparium 15. vii. 64 on Carex pseudocyperus, Sieraków reservation, Kampinoska Forest, em. 20. v. 65, leg. Nowakowski (PAN). 2 ♂♂, 2 ♀♀ from puparia 9 and 13. vii. 66 on Carex sp., Wierzba, near Popielno, Piska Forest, Poland, em. 14-22. vii. 66, leg. Nowakowski (PAN). 1 ♂, 1 ♀ from larvae 24. vi. 66 on Carex hudsonii, Mierzeja Wiślana, Krynica Morska, Poland, em. 18. vii. 64, leg. Nowakows-KI (PAN).

This species is readily distinguishable from *merella* by its bare mesoscutum and sexually dimorphic antennal public ence. NIXON'S (1944) original description suggests that it also differs from *merella* in the length of vein 2r ("abscissa 1 of the radius"). But from examination of the additional material now available I am not of the opinion that any clear distinction can be made in this respect.

NIXON (1944) has described as *Dacnusa orissa* a single male which closely resembles *pelion*, but is believed to represent a distinct species on account of differences in the sculpturation of its mesosternum and petiole. The differences are given in key 2 in the section below headed "Keys to *Chorebus* HALIDAY".

# The Chorebus affinis group

The species which I include in this concept are considered to represent a monophyletic group through their possession of the following three apomorph characters: (i) hind coxa with a conspicuous matted tuft or crest of pubescence on its posterior margin; (ii) precoxal suture represented by a smooth linear groove which extends almost to the hind margin of the mesepisternum; and (iii) mandible with tooth 2 long and pointed, but tooth 3 (the additional tooth) small or absent. The loss of the additional mandibular tooth in some species is clearly secondary, being correlated with the increasing length of tooth 2 (as can be seen from figs. 204-207).

Some of the species which I include in this group have been placed in separate genera (*Chorebus* HALIDAY in a restricted sense, *Gyrocampa* FÖRSTER and *Paragyrocampa* TOBIAS) by some previous authors. My reasons for rejecting these concepts were given in Part I (GRIFFITHS, 1964, page 849). It is not in practice possible to draw a clear-cut distinction between species in which vein  $R_s$  is evenly curved and those in which it is slightly sinuate, as was done in NIXON's (1943) generic key, and I have not therefore made much use of this character in the key to species given below in the section headed "Keys to *Chorebus* HALI-DAY".

Some of the species included in *Chorebus* in the former restricted sense appear to represent a smaller monophyletic group within the *affinis* group, characterised by their narrow pterostigma, 3-segmented labial palpi<sup>4</sup> and densely pubescent mesoscutum. I propose to refer to these as the *Chorebus uliginosus* group. The six species concerned are included in couplets 20-23 of the key below (in the section headed "Keys to *Chorebus* HALIDAY"). Most of these species are associated with the Ephydrid leaf-mining genus *Hydrellia*, and since the Dacnusini are not known as parasites of other genera of Ephydridae, I concluded (Part I, page 871) that this host association had arisen through transference from an Agromyzid host. The discovery of the life-history of *C. gracilipes* (THOMSON), which is clearly referable to the same group as the *Hydrellia*parasites, indicates that this transference was probably from an *Icteromyza* host to *Hydrellia*.

One of the new species described in this paper, *C. metallicus* sp. nov., is remarkable for the reduction of the rosette of metapleural pubescence, which is so fine as to be barely visible in some angles of light. However the species seems typical of the *affinis* group in most other respects, and I think it must be concluded that the reduction of its metapleural pubescence is secondary. (The plesiomorph condition of the metapleuron in the Dacnusini is as shown in fig. 16, with long sparse hairs, not dense fine pubescence of the type found in *metallicus*.)

There are a number of species with bare mesoscuta (included in couplets 2-10 of the key below), whose life-histories have not yet been discovered. I think it

<sup>&</sup>lt;sup>4</sup> C. miodes (NIXON) also has 3-segmented labial palpi, but this is probably due to convergence as this species is very different from those now under consideration in other respects.

<sup>7</sup> Beitr. Ent. 18, H. 1/2

probable that some or all of these are parasites of further species of *Cerodontha* (particularly subgenus *Icteromyza*). There is no information that the *affinis* group contains any parasites of non-Agromyzid hosts apart from the *Hydrellia*-parasites mentioned above included in the subordinate *uliginosus* group.

In Part I (GRIFFITHS, 1964, page 850) I suggested a connection betwen C. cytherea (NIXON) and the "merella-group". These remarks are perhaps somewhat misleading as they stand. C. cytherea (NIXON) shows certain features which are plesiomorph for the affinis group, namely the clearly 4-toothed mandibles and distinctly sinuate vein  $R_s$ , and in these respects resembles C. merella (NIXON) and certain similar species. Such resemblance in relatively plesiomorph characters cannot be considered to indicate monophyly. However the presence of such plesiomorph characters in some species of the affinis group associated with hosts of the subgenus Dizygomyza indicates that the association of the ancestral species of the group was probably with host species belonging to, or ancestral to, this host group.

It is possible that the *affinis* group is the sister-group of the *senilis* group s.l., treated in Part IV (GRIFFITHS, 1967). Both groups include species of elongate form with conspicuous hind coxal tufts of matted pubescence. But in the groundplan of the *senilis*-group the rugose precoxal suture is retained, and the mandibles are not long and pointed. Within the *senilis* group there is one subordinate group, the *bathyzonus* group, whose species might easily be confused with some species of the *affinis* group, since they possess (through convergence) a similar smooth precoxal suture. To avoid confusion careful attention should be paid to the form of the mandibles (see further the preliminary key in the section below headed "Keys to *Chorebus* HALIDAX"). As stated in Part IV, the presence of very dense opaque pubescence on the sides of the *senilis* group s.l. This character is absent from the groundplan of the *affinis* group (such as *iridis, fordi* and *stilifer*). Such species could easily be confused with species of the species of this group (such as *iridis, fordi* and *stilifer*).

Species included in *Chorebus* (in a restricted sense) and *Gyrocampa* by recent authors, but not included in my present concept of the *affinis* group, are *Chorebus striolus* STELFOX and *C. scabrifossus* STELFOX, now referred to the *rousseaui* group, *Gyrocampa thomsoni* ROMAN (synonymised with *Chaenusa conjungens* (NEES) in Appendix IX in Part III), and *G. pospelovi* KURDJUMOV. I have not been able to examine the holotype of the latter species, and the description (KURDJUMOV, 1912) is not sufficient to establish its generic position. If the insect was in fact bred from the Chloropid Oscinella frit (L.), as stated, probably it represents a *Coelinius* species, as no other genus of Dacnusini is known to contain parasites of Chloropidae.

THOMSON'S (1895) Dacnusa (Dacnusa) obliqua was doubtless a species of the affinis group, but it is impossible to give a firm interpretation of the name as the type has been destroyed: the four remaining specimens attributed to obliqua in the THOMSON collection are not from the type locality, and do not all belong to

the same species. In the circumstances I propose to discard the name as a nomen dubium.

In the descriptions which follow the following characters may be assumed, wherever nothing to the contrary is stated.

Clypeus dark, like the face. Face weakly punctate or almost smooth, covered with dense fine pubescence which is directed inwards or upwards over its centre, but downwards along the eye-margins. Precoxal suture represented by a well-defined smooth linear groove which extends virtually to the hind margin of the mesepisternum. Pubescence of hind coxa very dense, forming a conspicuous matted crest or basal tuft on its posterior margin. Metapleural pubescence very dense (compare figs. 22 and 23 in Part I), forming a rosette around the strongly rugose swelling: propodeum strongly rugose, covered with similar dense pubescence. Tergite 3 without basal pubescence. Wing (figs. 193-200) with vein  $Cu_{1b}$  well developed, forming a distinct angle with the transverse section of  $Cu_1$ .

Chorebus cytherea (NIXON), comb. nov.

Dacnusa cytherea NIXON, 1937 and 1944 Dacnusa tesmia NIXON, 1944, syn. nov. Dacnusa calliope NIXON, 1944, syn. nov.

Colour. Palpi and labrum yellow or ochreous yellow. Centre of mandibles yellow-brown. Antennae usually with their basal segments yellow-brown about as far as the third flagellar segment, but occasionally more or less entirely dark. Legs sometimes uniformly deep yellow with only tarsal segments 5 slightly infuscated, sometimes with more extensive brown coloration especially on the hind coxae and hind femora. Gaster beyond petiole more or less uniformly dark brown.

Morphology. Antennal segments: 3, (29)-31-36, (39); 9, 27-34-(35): flagellum long, with its more apical segments about  $2\frac{1}{2}$  times as long as wide: basal flagellar segments evenly covered with rather long pubescence in both sexes. Palpi long (see the table of biometric data). Mandible (fig. 204) narrowed towards its apex, 4-toothed, with tooth 2 relatively long and pointed and tooth 3 visible as a small projection from its side. Eyes slightly convergent below in the female, but not in the male. Back of head largely bare, with only some fine pubescence near the base of the mandibles.

Thorax elongate, 1.5-1.6 times as long as high. Sides of pronotum largely bare and shining, with only some fine pubescence along and below the oblique suture. Mesoscutum usually with its entire surface roughened (especially anteriorly) and densely covered with long whitish pubescence (but in some small specimens the lateral lobes tend to become smooth and bare posteriorly): notaulices with only their lateral extensions distinct (margined by a slightly raised crest). Pubescence of metapleuron and propodeum extremely dense. Petiole covered with similar, extremely dense whitish pubescence, which tends to be parted along its centre-line and radiates outwards around its apical half 7\* (fig. 208). Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

Wing (fig. 193) with vein  $R_s$  slightly sinuate and *Im-cu* rejected from cell  $R_s$ .

# Breeding records

# Host 1 — Cerodontha (Dizygomyza) spinata GROSCHKE

4 33, 5 99 from puparia on *Carex silvatica*, Schongau, Oberbayern, Germany, em. 1-20. viii. 53, leg. GROSCHKE (STGT and GCDG). 2 99 from larvae 13. v. 66 on *Carex* sp., Stadtwald, Mühlhausen, Thuringia, Germany, em. 2 and 12. viii. 66, leg. ВUHR no. 2727 (GCDG).

### Host 2 — Cerodontha (Dizygomyza) luctuosa MEIGEN (= effusi KARL)

1 δ, 4 QQ from puparia on Juncus effusus, Stuttgart (Feuerbach Tal), Germany, em. 10–16. xii. 53, leg. GROSCHKE (STGT). 1 δ from puparium on Juncus effusus, Stuttgart-Wildpark, em. 20. iv. 54, leg. GROSCHKE (STGT). 3 ex. from larvae and puparia 23. vii. 64 on Juncus effusus, Górki Wschodnie, near Gdansk, Poland, em. 20. vii-7. viii. 64, leg. NOWAKOWSKI (PAN and GCDG). 1 Q from larva 25. vii. 64 on Juncus effusus, Żukowo, distr. Kartuzy, Poland, em. 10. viii. 64, leg. NOWAKOWSKI (PAN). 1 Q from larva 30. vii. 64 *on Juncus* effusus, Mirachowo, distr. Kartuzy, em. 7. ix. 64, leg. NOWAKOWSKI (PAN). 1 δ from puparium 1. xi. 66 on Juncus effusus, Palmiry, Kampinoska Forest, Poland, em. 10. ii. 67, leg. NOWAKOWSKI (PAN). 1 Q from larva 7. xi. 57 on Juncus effusus, Sieraków reservation, Kampinoska Forest, em. 6. iii. 58, leg. NOWAKOWSKI (PAN): 1 δ 1 Q from puparia 15. vii. 64 on Juncus effusus, same locality, em. 20–23. vii. 64, leg. NOWAKOWSKI (PAN).

## Host 3 — Cerodontha (Dizygomyza) hirtae Nowakowski (= luctuosa auctt.)

2 dd from larvae 14. vii. 66 on Carex hirta, Iznota, Piska Forest, Poland, em. 8–11. viii. 66, leg. Nowakowski (PAN). 6 ex. from larvae 18. x. 64 on Carex hirta, Łazy, distr. Piaseczno, Poland, em. 14–22. xii. 64, leg. Nowakowski (PAN and GCDG). 1  $\circ$  from puparium 7. vii. 64 on Carex hirta, Warszawa-Pyry, em. vii. 64, leg. Nowakowski (PAN): 2  $\circ \circ$  from puparia 5. xi. 65, same plant and locality, em. 25. iii. 66, leg. Nowakowski (PAN). 1 dfrom larva 20. vi. 64 on Carex hirta, Warszawa-Powsin, em. 21. vii. 64, leg. Nowakowski (PAN). 2 ex. from larvae 6. vii. 64 on Carex hirta, Warszawa-Bielany, em. 27. vii and 10. viii. 64, leg. Nowakowski (PAN). 1  $\circ$  from larva 3. xi. 55 on Carex hirta, Cybulice, Kampinoska Forest, Poland, em. 23. v. 56, leg. Nowakowski (PAN). 1 ex. from larva 25. vii. 64 on Carex hirta, Żukowo, distr. Kartuzy, Poland, em. 17. v. 65, leg. Nowakowski (PAN).

Host 4 — Cerodontha (Dizygomyza) chaixiana GROSCHKE

3 33, 6 99 from puparia on *Poa chaixii*, Stuttgart (Kräherwald and Wildpark), Germany, em. 4. iii-20. iv. 54, leg. GROSCHKE (STGT and GCDG). 1 3 3 99 from puparia vi. 54 on *Poa chaixii*, Vaihingen, Stuttgart, em. 21-28. vii. 54, leg. GROSCHKE (STGT).

### Host 5 – Cerodontha (Poemyza) pygmaea MEIGEN

1 ex. from puparium 30. vii. 64 on *Deschampsia caespitosa*, Mirachowo, distr. Kartuzy, Poland, em. 15. viii. 64, leg. Nowakowski (PAN).

The single record of this species from C. (*Poemyza*) pygmaea MEIGEN is probably to be considered an example of xenophagy.

NIXON (1944) attempted to define three species of his "cytherea-subgroup", but commented on a number of specimens which did not satisfactorily fit his definitions. I have not been able to make any division of the bred material before me on the basis of the characters which he suggested, namely colour, the shape of the head and the number of antennal segments. The series bred from C. (Dizygomyza) chaixiana GROSCHKE, a very small host, are very small speci-

mens with a low number of antennal segments (3, 30-35; 9, 27-32), as in NIXON's proposed species *calliope*; but some of these insects have relatively large heads (not very strongly transverse), and thus erode the suggested difference between NIXON's calliope and his other two species. The larger specimens of this series seem fully comparable with specimens bred from C. (D.) luctuosa MEIGEN and C. (D.) hirtae NOWAKOWSKI, and it seems probable that the statistical difference in the number of antennal segments is due merely to correlation with the small size of the host. The small differences suggested between cytherea sensu stricto and tesmia seem to me to be within the range of variation of the material bred from C. (D.) luctuosa MEIGEN and C. (D.) hirtae NOWAKOWSKI, and I cannot therefore accept the separation of these species. The short series bred from C. (D.) spinata GROSCHKE pose a further problem of size correlation. This host is substantially larger than the other hosts and the parasites bred from it have a larger average size and higher range of antennal segments (3, 36, 39; $\mathcal{Q}$ , 32, 33 (2 ex.), 34 (2 ex.), 35). But there seems to be no clear-cut difference between these insects and the other material, and the statistical differences may again be due merely to correlation with the size of the host.

On balance I do not think that there is sufficient evidence for dividing the wide concept of *cytherea* here proposed into a number of species, although it should be recognised that the range of variation in size and number of antennal segments is abnormally wide. Final clarification of this problem may not be possible without breeding experiments.

The unique petiole of *cytherea* (fig. 208) will enable the species to be readily identified. In no other known species does the pubescence on the apical half of the petiole radiate outwards in this way.

# Chorebus stilifer sp. nov.

Colour. Palpi and labrum yellow. Centre of mandibles yellow-brown. Antennae with their basal segments yellowish (clearly so usually about as far as the second flagellar segment), this colour merging gradually into the darker colour of the more distal flagellar segments. Legs uniformly yellow. Gaster beyond petiole largely bright yellow or orange, in strong contrast with the shining black petiole.

Morphology. Antennal segments:  $\mathcal{J}$ , 32 (2 ex.);  $\mathcal{Q}$ , 31-32: flagellum long and slender, with all its segments over three times as long as wide: flagellar pubescence somewhat sexually dimorphic, in the female the entire flagellum is densely covered with rather long pubescence, but in the male the segments of about the basal third of the flagellum are somewhat shining and bear only rather short, sparse pubescence. Palpi very long (see the table of biometric data). Mandible weakly 4-toothed, with tooth 2 relatively long and pointed but tooth 3 weak, visible as a small projection from the side of tooth 2 (sometimes almost absent). Eyes strongly convergent below in the female (minimum distance between them about one-third of the head width), less markedly so in the male. Face smooth

and shining, only sparsely pubescent. Back of head with rather dense pubescence near the base of the mandibles, but otherwise virtually bare.

Thorax elongate, about 1.5 times as long as high. Sides of pronotum with their entire surface strongly sculptured (more or less punctate) and completely covered with very dense, adpressed pubescence. Mesoscutum with most of its surface roughened (especially its anterior face and central lobe), entirely covered with dense, rather long pubescence: notaulices with only their lateral extensions distinct (margined by a slightly raised crest). Subalar callus also covered with dense pubescence. Petiole extraordinarily long and narrow, virtually bare. Ovipositor ( $\varphi$ ) not projecting beyond the apical tergite in the retracted position.

Wing with vein  $R_s$  very slightly sinuate and 1m-cu rejected from cell  $R_s$  (compare cytherea, fig. 193).

# Host – Cerodontha (Butomomyza) staryi HERING

Holotype  $\Im$ ; 2 33, 6  $\Im$  paratypes from larvae and puparia 5. xii. 53 on *Carex* sp. cf. remota, Stuttgart (Feuerbach Tal), Germany, em. 9–15. ii. 54, leg. GROSCHKE (STGT and GCDG).

The above material was labelled by GROSCHKE as bred from "Phytobia stilettata". But in the event he did not publish this name, but described the host as "Phytobia (Pöemyza) starýi HERING var. normalis nov." (GROSCHKE, 1954).

This species may be easily distinguished from other species of the *affinis* group by the form of its petiole and the brightly coloured gaster. In these respects it has a strong superficial resemblance to C. *bathyzonus* (MARSHALL), included in the *senilis* group s.l. (see Part IV). But the form of the mandibles and the public public of the head are clearly different.

### Chorebus iridis sp. nov.

Colour. Palpi and labrum yellow or ochreous yellow. Centre of mandibles redbrown. Antennae largely black, but usually becoming brown or reddish towards their base (as far as about the second flagellar segment). Legs more or less uniformly ochreous yellow, testaceous or yellow-brown, with only tarsal segments 5 distinctly infuscated. Gaster entirely black.

Morphology. Antennal segments: 3, 31 (2 ex.);  $\varphi$ , 31 (3 ex.), 32: flagellum with its more apical segments  $2-2\frac{1}{2}$  times as long as wide: basal flagellar segments rather densely covered in both sexes with pubescence of moderate length (see the table of biometric data). Mandible (fig. 205) 4-toothed, with tooth 2 relatively long and pointed and tooth 3 developed as a distinct projection from its side. Eyes rather strongly convergent below in the female (minimum distance between them about two-fifths of the head width), but hardly so in the male. Back of head largely bare, but with some fine inconspicuous pubescence near the base of the mandibles.

Thorax elongate, about 1.4 times as long as high. Sides of pronotum with much of their area finely sculptured (more or less punctate) and covered with very dense adpressed whitish pubescence, but with a broad band of smooth bare shining surface along their dorsal margin. Mesoscutum with its anterior face and the anterior part of its central lobe somewhat roughened, with dense pubescence covering its anterior face and extending onto about the anterior third or half of its dorsal surface (both central and lateral lobes), but the posterior half of its dorsal surface is almost bare: notaulices with only their lateral extensions well developed (margined by a slightly raised crest), their longitudinal extensions being usually almost absent, at most represented by weak impressions. Petiole slightly widened towards its apex, with sparse hairs along its sides (but absent from its centre-line) and weak accumulations of fine pubescence at its apical corners. Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

Wing with vein  $R_s$  slightly sinuate and 1m-cu well rejected from cell  $R_s$  (compare cytherea, fig. 193).

### Host – Cerodontha (Dizygomyza) iridis HENDEL

Holotype 9; 2 33, 3 99 paratypes from puparia 21. iv. 66 on *Iris foetidissima*, near Sorrento, Campania, Italy, em. 29. iv-4. v. 66, leg. SPENCER (GCDG).

This species is a relatively plesiomorph member of the *affinis* group in respect of its 4-toothed mandibles and distinctly sinuate vein  $R_s$ . One of its most distinctive features is the presence of dense whitish pubescence on the sides of the pronotum, in which respect it resembles *C. fordi* (NIXON). The differences between these species are given in the key in the section below headed "Keys to *Chorebus* HALIDAY".

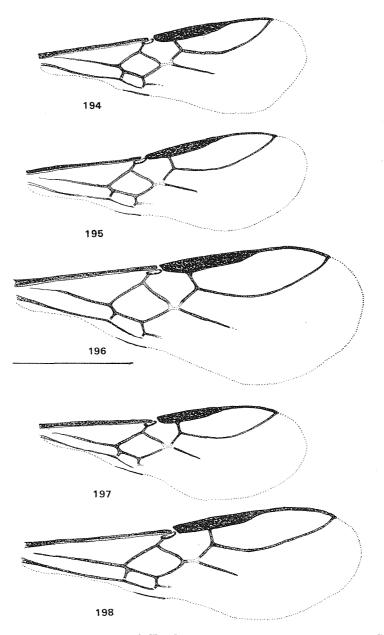
# Chorebus fordi (NIXON), comb. nov.

### Gyrocampa fordi NIXON, 1954

Colour. Palpi and labrum yellow or yellow-brown. Centre of mandibles brown. Antennae more or less entirely dark (at most with the scape, pedicel and annellus brownish). Legs largely deep yellow, with only the more apical tarsal segments and sometimes the hind coxae somewhat infuscated. Gaster beyond petiole dark brown.

Morphology. Antennal segments: 3, 30; 9, 30-32: flagellum with its more apical segments  $2\frac{1}{2}-3$  times as long as wide: all antennal segments densely pubescent in both sexes. Palpi long (see the table of biometric data). Mandible very small, with tooth 2 long and pointed but the additional tooth (tooth 3) represented only by a small projection from its side. Eyes only slightly convergent below. Back of head largely bare, but with some fairly dense pubescence near the base of the mandibles.

Thorax rather elongate, about 1.3 times as long as high. Sides of pronotum with most of their surface covered with conspicuous, densely matted, whitish pubescence (similar to that of the metapleuron), but with a band of bare shining surface along their dorsal margin (this bare area is sculptured, not smooth as in *iridis*). Mesoscutum with its anterior face roughened and densely pubescent, but its dorsal surface largely smooth, bare and shining, with only some sparse



Figs. 194–198. Wings of Chorebus spp.: 194, C. fordi  $(NIXON) \circ$ ; 195, C. siniffa  $(NIXON) \circ$ ; 196, C. vernalis sp. nov.  $\sigma$ ; 197, C. metallicus sp. nov.  $\circ$ ; 198, C. nigriscaposus  $(NIXON) \circ$ . (Scale 1 mm.)

pubescence on the anterior part of the central lobe and a few hairs along the course of the notaulices: notaulices with only their lateral extensions well developed (margined by a slightly raised crest), their longitudinal extensions visible only as weak impressions. Mesepisternum with a tuft of dense whitish pubescence (similar to that of the metapleuron) above the middle coxa. Pubescence of metapleuron and propodeum very dense and whitish. Petiole virtually parallel-sided, largely bare and shining, but with small patches of fairly dense pubescence near its base, a few fine hairs along its sides and weak accumulations of fine pubescence at the apical corners. Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

Wing (fig. 194) with vein  $R_s$  almost evenly curved, hardly sinuate; Im-cu well rejected from cell  $R_s$ .

### Breeding records

Host — Cerodontha (Poemyza) lateralis MACQUART

2 qq from puparia 17. vi. 64 on Agropyron repens, Warszawa-Pyry, Kabackie Forest, Poland, em. 24. vi. 64, leg. Nowakowski (PAN and GCDG).

The most characteristic features of this species are the dense whitish pubescence on the sides of the pronotum (as is found also in *C. iridis* sp. nov.), the very small mandibles and the about evenly curved vein  $R_s$ .

Chorebus siniffa (NIXON), comb. nov.

Dacnusa siniffa NIXON, 1937

Gyrocampa siniffa (NIXON), NIXON, 1949

Colour. Palpi and labrum yellow or ochreous yellow. Centre of mandibles redbrown. Antennae largely black, at most becoming brownish at their base as far as about the first flagellar segment. Legs largely golden yellow, but with the tarsi becoming infuscated towards their apex and sometimes also the hind coxae partly infuscated. Gaster entirely dark.

Morphology. Antennal segments: 3, 28-30;  $\Im$ , 26-29: flagellum with its more apical segments  $2\frac{1}{2}$ -3 times as long as wide: the basal flagellar segments have a strongly shining appearance in the male, but are evenly covered with pubescence which is almost as dense as in the female. Maxillary palpi long (see the table of biometric data). Mandible with tooth 2 very long and pointed, but the additional tooth (tooth 3) visible only as a small projection from its side. Eyes only slightly convergent below (hardly more so in the female than in the male). Back of head almost completely bare (with only some fine inconspicuous pubescence near the base of the mandibles). Facial pubescence very fine, hardly obscuring the shining, almost smooth surface beneath.

Thorax elongate, about 1.6 times as long as high. Sides of pronotum largely bare and shining, with only some fine inconspicuous pubescence along and below the oblique suture. Mesoscutum largely smooth, bare and shining, with some fine pubescence on its anterior face (especially at the "shoulders"), but otherwise only a few hairs along the course of the notaulices: notaulices with their lateral extensions (i.e. at the "shoulders") distinct, margined by a slightly raised crest, but hardly extending longitudinally on the dorsal surface of the mesoscutum (at most visible as smooth lines). Petiole not or only slightly widened towards its apex, almost completely bare. Ovipositor ( $\mathcal{Q}$ ) not projecting beyond the apical tergite in the retracted position.

Wing (fig. 195) with cell  $2R_1$  rather elongate; vein  $R_s$  almost evenly curved, hardly sinuate; 1m-cu rejected from cell  $R_s$ .

## Breeding records

Host – Cerodontha (Dizygomyza) morosa Meigen

l 3 2 99 from larvae 24. vii. 66 on Carex vulpina, Mierzeja Wiślana, Krynica Morska, Poland, em. 3-18. vii. 66, leg. Nowakowski (PAN and GCDG).

There are several species with bare mesoscuta similar to *siniffa* whose lifehistories have not yet been established. The differences are given in the key in the section below headed "Keys to *Chorebus* HALIDAY".

# Chorebus vernalis sp. nov.

Colour. Palpi and labrum yellow. Centre of mandibles yellow-brown. Antennae with their basal segments contrastingly yellowish as far as the second flagellar segment. Legs uniformly golden yellow, with only the tarsi becoming infuscated towards their apex. Gaster more or less entirely dark.

Morphology. 32 antennal segments (3): flagellum with all its segments entirely covered with dense pubescence, the more apical segments being  $2\frac{1}{2}-3$ times as long as wide. Palpi long (see the table of biometric data). Mandible with tooth 2 very long and pointed, but the additional tooth (tooth 3) visible only as a weak projection from its side. Back of head largely bare, but with some fairly dense whitish pubescence near the base of the mandibles.

Thorax elongate, about 1.5 times as long as high. Sides of pronotum with a broadly bare and shining area along their dorsal margin, but with dense pubescence covering much of their surface; however this is fine and somewhat upstanding, lacking the opaque whitish appearance of *fordi* and *iridis*. Mesoscutum with only its anterior face roughened, with dense pubescence covering its anterior face and extending onto the anterior half of the central lobe, but most of its dorsal surface is bare and shining: notaulices with their lateral extensions (i.e. at the "shoulders") rugose and margined by a raised crest, but only extending longitudinally as smooth lines on the dorsal surface of the mesoscutum. Petiole more or less parallel-sided, strongly rugose, with dense, rather long pubescence covering much of its surface, but becoming broadly bare along its centre-line on its apical half.

Wing (fig. 196) with vein  $R_s$  evenly curved, not at all sinuate; *1m-cu* only narrowly rejected from cell  $R_s$ .

(ç unknown).

### Host — Cerodontha (Dizygomyza) caricicola HERING

Holotype 3 from puparium 18. ii. 62 on *Carex* sp., Linton, Cambs., England, em. 13. iv. 62 (GCDG).

This insect clearly represents a distinct species characterised by the combination of dense pubescence on the sides of the pronotum, a partly bare mesoscutum (with pubescence extending only onto the anterior half of the central lobe), an extensively pubescent petiole and an evenly curved vein  $R_s$ .

# Chorebus metallicus sp. nov.

Colour. Palpi golden yellow or yellow-brown. Labrum brown. Centre of mandibles yellow-brown or testaceous. Antennae with the flagellum entirely dark, but the scape, pedicel and annellus testaceous or yellow-brown. Legs (including coxae) uniformly golden yellow, with only the tarsi becoming infuscated towards their apex. Gaster entirely dark.

Morphology. Antennal segments: 3, 23-24; 9, 21-22: flagellum with its more apical segments relatively long,  $2\frac{1}{2}-3$  times as long as wide: all antennal segments entirely covered with dense pubescence in both sexes. Maxillary palpi long (see the table of biometric data). Mandible 3-toothed, with tooth 2 extremely long and pointed (the additional tooth, tooth 3 in 4-toothed species, being absent). Eyes slightly more convergent below in the female than in the male. Back of head almost completely bare (without any dense pubescence near the base of the mandibles). Facial pubescence very fine, hardly obscuring the shining, almost smooth surface beneath.

Thorax elongate, 1.4-1.5 times as long as high. Sides of pronotum completely bare and shining. Mesoscutum largely smooth, bare and shining, with only a few fine hairs on the "shoulders" and along the course of the notaulices: notaulices with their lateral extensions (i.e. at the "shoulders") distinct, margined by a raised crest, but only extending longitudinally as smooth lines on the dorsal surface of the mesoscutum. Pubescence of hind coxa forming a whitish matted tuft on its posterior margin, but this is much less conspicuous than in other species of the *affinis* group. Metapleuron with a well-defined swelling on its lower half (as in other species of the *affinis* group), but the surrounding rosette of pubescence, though dense, is extremely fine and inconspicuous, only distinct in certain angles of light. Propodeum deeply rugose, strongly shining, with only fine inconspicuous pubescence towards its sides. Petiole virtually bare, slightly widened towards its apex. Gaster ( $\mathcal{Q}$ ) somewhat pointed apically, with the ovipositor fairly long, often slightly projecting beyond the apical tergite in the retracted position.

Wing (fig. 197) with vein  $R_s$  evenly curved, not at all sinuate; 1m-cu rejected from cell  $R_s$ ; vein  $Cu_{1b}$  represented only by a shadow, so that cell 2Cu is open at its lower distal corner.

### Host – Cerodontha (Icteromyza) geniculata FALLÉN

Holotype  $\varphi$ ; 7 33, 5  $\varphi\varphi$  paratypes from larvae and puparia 15-17. viii. 66 on *Eriophorum latifolium*, Mata Babia Góra, Marków pond, Poland, em. 31. i-11. ii. 67, leg. NOWAKOWSKI (PAN and GCDG).

This species is readily separable from other members of the *affinis* group by the remarkable reduction of its metapleural and propodeal pubescence, on which some comment has already been made above.

#### Chorebus nigriscaposus (NIXON), comb. nov.

## Gyrocampa nigriscaposa NIXON, 1949

Gyrocampa propodealis NIXON, 1949, syn. nov.

Colour. Palpi and labrum varying from deep yellow to brown. Centre of mandibles ochreous or yellow-brown. Antennae entirely dark except for the yellow-brown annellus. Legs often largely yellow, with only the tarsi and hind coxae somewhat infuscated; but in many of the Polish specimens the tibiae and hind femora are also brownish or testaceous, with only femora l and 2 remaining yellow. Gaster more or less entirely dark.

Morphology. Antennal segments:  $\mathcal{J}$ , (23)-24-27;  $\mathcal{Q}$ , 22-24: flagellum with its more apical segments  $2-2\frac{1}{2}$  times as long as wide: all antennal segments entirely covered with dense pubescence in both sexes. Palpi rather short (see the table of biometric data). Mandible (fig. 206) almost 3-toothed, with tooth 2 very long and pointed, but the additional tooth (tooth 3 in 4-toothed species) either absent or represented only by a weak notch on the side of tooth 2. Eyes slightly convergent below in the female, but hardly so in the male. Back of head largely bare, but with some rather dense pubescence near the base of the mandibles.

Thorax elongate, about 1.4-1.5 times as long as high. Sides of pronotum largely bare and shining, with distinct pubescence only at their ventral corner. Mesoscutum with its anterior face roughened, with fine pubescence covering its anterior face and central lobe, but the lateral lobes are largely bare: notaulices with their lateral extensions (i.e. at the "shoulders") distinct, margined by a raised crest, but hardly extending longitudinally on the dorsal surface of the mesoscutum. Mesepisternum with dense whitish pubescence similar to that of the metapleuron above the middle coxa. Pubescence of metapleuron and propodeum extremely dense, forming three distinct rosettes on either side — around the swelling on the lower half of the metapleuron (which is very prominent and sharply delimited), around the spiracle on the propodeum and around a very prominent tubercle behind the spiracle (as also in *lanigerus*). Petiole widened towards its apex, with its dorsal surface almost bare. Gaster ( $\mathcal{Q}$ ) somewhat pointed apically, with the ovipositor fairly long, usually slightly projecting beyond the apical tergite in the retracted position.

Wing (fig. 198) with cell  $2R_1$  rather elongate; vein  $R_s$  more or less evenly curved, hardly sinuate; 1m-cu rejected from cell  $R_s$ .

Breeding records

Host 1 – Cerodontha (Icteromyza) geniculata FALLÉN

20 33 21 99 from larvae and puparia 15-17. viii. 66 on *Eriophorum latifolium*, Mata Babia Góra, Marków pond, Poland, em. 22. viii-12. ix. 66 and 2-8. ii. 67, leg. Nowakowski (PAN and GCDG).

Host 2 — Cerodontha (Icteromyza) calosoma HENDEL 1 3, Randers and 1 9, Aarhus, Jutland, Denmark, leg. Schlick (KB).

The two specimens bred from C. (I.) calosoma HENDEL emerged from the posterior end of the host puparium, as did the specimens of C. lanigerus (STELFOX) bred from the same host: however the series bred from C. (I.) geniculata FALLÉN emerged from the anterior end of the puparia, as is normally the case in the Dacnusini and other Alysiinae.

The type material of *nigriscaposus* and *propodealis* was taken at the same locality. I have not been able to examine the holotype of *propodealis*, which is now in Washington: but the small differences in wing venation and propodeal pubescence given in NIXON'S (1949) original descriptions do not seem to me sufficient to demonstrate that two species are involved, and I am therefore proposing to regard both names as referring to the same species. One typographical error in the description of *propodealis* should be noted: Mr. STELFOX informs me that the female holotype has 23 antennal segments, not 28 as printed.

The prominent tubercle behind the spiracle on the propodeum in *nigriscaposus* is a feature found also in *lanigerus* and the *uliginosus* group. But *nigriscaposus* is the only species showing this feature in which the lateral lobes of the meso-scutum are bare.

### Chorebus lanigerus (STELFOX), comb. nov

## Gyrocampa lanigera STELFOX, 1957

Colour. Palpi yellow or ochreous yellow. Centre of mandibles yellow-brown. Antennae more or less entirely dark. Legs deep yellow or testaceous, with all tarsi and usually the hind coxae somewhat infuscated. Gaster more or less entirely dark (at most with tergite 3 reddish).

Morphology. Antennal segments:  $\mathcal{J}$ , (25), 28-30;  $\mathcal{Q}$ , 25 (holotype): flagellum with its more apical segments  $2-2\frac{1}{2}$  times as long as wide: all antennal segments entirely covered with very dense pubescence in both sexes. Maxillary palpi of moderate length (see the table of biometric data): labial palpi 4-segmented. Mandible (compare fig. 206) almost 3-toothed, with tooth 2 very long and pointed, but the additional tooth (tooth 3 in 4-toothed species) represented only by a weak notch on the side of tooth 2. Vertex, temples and back of head largely covered with short, rather dense, evenly distributed pubescence (similar to that of many species of the *senilis* group s.l. except that it shows no tendency to form tufts near the base of the mandibles).

Thorax elongate, about 1.4 times as long as high. Sides of pronotum with dense whitish pubescence covering much of their surface (especially along and below the oblique suture), although there is an almost bare area towards their posterodorsal corner. Mesoscutum roughened anteriorly, entirely covered with dense pubescence: notaulices with only their lateral extensions distinct, not extending longitudinally on the dorsal surface of the mesoscutum. Mesepisternum with dense whitish pubescence, similar to that of the metapleuron, above the middle coxa. Pubescence of metapleuron and propodeum extremely dense: the posterior angles of the propodeum (behind the spiracles) bear very prominent tubercles, around which the pubescence tends to radiate, forming a rosette: there are thus three rosettes of radiating pubescence which can be seen on either side of the metapleura/propodeum (as also in *nigriscaposus*) — the rosette on the lower half of the metapleuron (where the swelling is unusually prominent and sharply delimited), the rosette around the spiracle on the propodeum, and the rosette around the tubercle behind this. Petiole widened towards its apex, almost entirely covered with extremely dense pubescence which tends to form whitish tufts at the apical corners. Ovipositor ( $\mathcal{Q}$ ) short according to STELFOX's (1957) description (I have not seen a female).

Wing (fig. 199) with cell  $2R_I$  narrow and elongate; pterostigma narrow and elongate, merging gradually into the metacarp; vein  $R_s$  more or less evenly curved, hardly sinuate; 1m-cu rejected from cell  $R_s$ .

# Breeding records

Host – Cerodontha (Icteromyza) calosoma HENDEL 3 33, Randers, Jutland, Denmark, leg. SCHLICK (KB).

The life-history of the host, identified from a fly in the SCHLICK collection associated with an identical puparium, is still unknown (see the remarks on the SCHLICK collection in the introduction to Part III, GRIFFITHS, 1966b). One unusual biological feature of this species should be noted. All the bred specimens have emerged from the posterior end of the host puparium by making a complete vertical split through the eighth abdominal segment.

This species is well characterised by the presence of dense pubescence on the back of its head, the pronotum, the entire surface of the mesoscutum and the petiole.

# Chorebus gracilipes (THOMSON), comb. nov.

## Dacnusa (Dacnusa) gracilipes THOMSON, 1895 (in part)

Colour. Palpi yellow-brown or brown. Labrum dark brown. Centre of mandibles yellow-brown or red-brown. Antennae entirely dark except for the yellowbrown annellus. Legs largely unicolorous deep yellow or yellow-brown, with the tarsi somewhat infuscated towards their apex. Gaster entirely dark.

Morphology. Antennal segments:  $\mathcal{J}$ , 25 (lectotype);  $\mathcal{Q}$ , 22-24: flagellum entirely covered with dense pubescence in both sexes, with its more apical segments about  $2\frac{1}{2}$  times as long as wide. Maxillary palpi of moderate length (see the table of biometric data): labial palpi short, appearing only 3-segmented (the apical two segments being virtually fused). Mandible (fig. 207) 3-toothed, with tooth 2 extremely long and pointed (the additional tooth, tooth 3 in 4-toothed species, being absent). Eyes only slightly convergent below. Back of head largely bare, but with some dense pubescence near the base of the mandibles.

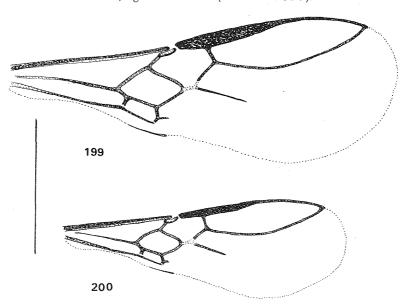
Thorax elongate, about 1.4 times as long as high. Sides of pronotum largely bare and shining (in strong contrast with the mesoscutum), but with some dense,

fine pubescence below the oblique suture. Mesoscutum with most of its surface slightly roughened, entirely covered with short, very dense pubescence: notaulices with their lateral extensions (i.e. at the "shoulders") distinct, margined by a slightly raised crest, but hardly extending longitudinally on the dorsal surface of the mesoscutum. Mesepisternum with dense whitish pubescence, similar to that of the metapleuron, above the middle coxa. Pubescence of metapleuron and propodeum extremely dense: the tubercle behind the spiracles on the propodeum, although not so prominent as in *nigriscaposus* and *lanigerus*, bears a conspicuous fringe of matted pubescence. Petiole strongly widened towards its apex, longitudinally striate, with its dorsal surface shining and almost bare. In the female the gaster becomes strongly pointed towards its apex, so that the ovipositor, although long and stout (fig. 209), only slightly projects beyond the apical tergite in the retracted position.

Wing (fig. 200) with vein  $R_s$  evenly curved, not at all sinuate; 1m-cu rejected from cell  $R_s$ .

# Breeding records

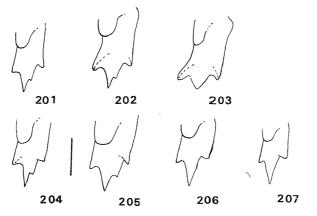
Host — Cerodontha (Icteromyza) geniculata FALLÉN 2 99 from puparia 17. viii. 66 on Eriophorum latifolium, Mata Babia Góra, Marków pond, Poland, em. 30. viii and 9. ix. 66, leg. NOWAKOWSKI (PAN and GCDG).



Figs. 199–200. Wings of Chorebus spp.: 199, C. lanigerus (STELFOX)  $\sigma$ ; 200, C. gracilipes (THOMSON) Q. (Scale 1 mm.)

This species is very similar to the well-known C. uliginosus (HALIDAY), a parasite of *Hydrellia* (Ephydridae), but has a longer, stouter ovipositor and less extensive pubescence on the back of the head.

The two syntypes of THOMSON'S *Dacnusa* (*Dacnusa*) gracilipes (from Örtofta, near Lund, Sweden) represent different species. Only the male, which is hereby designated lectotype, belongs to the species described above. The second syntype (a female) belongs to a species not otherwise known to me.



Figs. 201–207. Mandibles of Chorebus spp.: 201, C. talaris (HALIDAY); 202, C. asramenes (NIXON); 203, C. transversus (NIXON); 204, C. cytherea (NIXON); 205, C. iridis sp. nov.; 206, C. nigriscaposus (NIXON); 207, C. gracilipes (THOMSON). (Scale 0.1 mm.)



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Fig. 208. Petiole of Chorebus cytherea (NIXON). (Scale 0.1 mm. Scale line see fig. 209.)

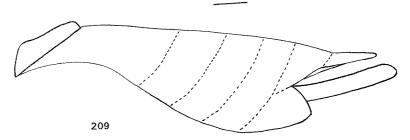


Fig. 209. Gaster in lateral view of *Chorebus gracilipes* (THOMSON) ♀. (Scale 0.1 mm.)

# Keys to Chorebus HALIDAY

A fundamental revision of the existing keys (in NIXON, 1943-54) to many groups of *Chorebus* spp. has been necessary, both as a result of my revision of the group concepts and to accomodate the large number of new species described in this and previous parts of this paper. Hitherto I have only given a fully revised key for the *senilis* group s.l. and the *cybele* group (in Part IV, GRIFFITHS, 1967). I must take full responsibility for what follows, although a few of the couplets still follow those in NIXON's keys.

### 1. Preliminary Key

- 2 Species of elongate form (thorax at least 1.3 times as long as high), with a conspicuous crest or basal tuft of matted pubescence on the posterior margin of the hind coxa. Vein Cu<sub>1b</sub> well developed in all except a few species, forming a distinct angle with the transverse section of Cu<sub>1</sub> (see fig. 154). (All species in which the precoxal suture is represented by a smooth linear groove, extending virtually for the entire length of the mesepisternum, are included under this alternative.)
   Hind coxa usually without a distinct tuft of matted pubescence. (Rugose precoxal suture present in all species except C. ampliator (NEES))

Note. A few species in the *ovalis/lateralis* complex included under this second alternative have weak tufts of pubescence on the hind coxa. Most of these have a short thorax, but three species of elongate form included in couplets 43-44 of the key to the *ovalis/lateralis* complex below may possibly be confused with species of the *senilis* group. They may be recognised as belonging to the *ovalis/lateralis* complex by careful attention to the form of cell 2Cu, which is open at its lower distal corner through the virtual loss of vein  $Cu_{1b}$ .

- 3 Tergite 3 (the tergite immediately following the petiole) longitudinally striate and separated from the following tergite by a well-defined groove. Precoxal suture very broad, strongly rugose-costate. Mandibles virtually 3-toothed, dominated by the long pointed central tooth. Labial palpi 3-segmented . . . . C. rousseaui group<sup>5</sup>

Note. This concept includes the *bathyzonus* group, whose species have a smooth precoxal suture similar to that of the *affinis* group. However in the *bathyzonus* group the mandibles are strongly 4-toothed, not long and pointed as in the *affinis* group.

<sup>5</sup> The available names for species of this group are *Dacnusa rousseaui* SCHULZ (1907), *Dacnusa obscuripes* RUSCH-KA (1913), *Chorebus striolus* STELFOX (1957) and *Chorebus seabrifossus* STELFOX (1957). BURGHELE (1960) has suggested that the first three of these names refer to a single species. The life-history of these curious insects is still unknown. They are probably closely related to the affinis and senilis groups (with which they appear synapomorph in respect of the tuft of matted publescence on the hind coxa), but further investigation is needed.

8 Beitr. Ent. 18, H. 1/2

3

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Precoxal suture represented by a well-defined smooth linear groove, which extends virtually for the entire length of the mesepisternum. Mandibles with tooth 2 long and pointed, the additional tooth (tooth 3 in 4-toothed species) being either weak or absent (see figs. 204-207). Pubescence of pronotum variable. (Species in which the sides of the pronotum are densely pubescent may be separated from the bathy-zonus group by the form of their mandibles.) . . . . . . . . . C. affinis group (see key 3 below)

5	Wing with vein $Cu_{1b}$ well developed, forming a distinct angle with the transverse
	section of $Cu_I$ (compare fig. 154). Petiole elongate, at least 1.9 times as long as wide,
	almost bare plesiomorph species related to the affinis and senilis groups
	(see key 2 below)

### 2. Plesiomorph species related to the affinis and senilis groups

1	Mandibles (fig. 203) 3-toothed (similar to those of most species of <i>Exotela</i> and <i>Dac-nusa</i> ). Wing (fig. 190) with vein $R_s$ strongly sinuate. Mesoscutum extensively pubescent, at most with the lateral lobes bare C. transversus (NIXON) Host: Cerodontha (Butomomyza) angulata LOEW
	Mandibles clearly 4-toothed (see fig. 25). Wing as figs. 191 and 192, with vein $R_s$ only weakly sinuate $\ldots \ldots 2$
2	Thorax fairly short, $1.2-1.3$ times as long as high: hind tarsi short, about 0.8 times the length of the hind tibiae. Notaulices complete, reaching the posterior fovea as deep rugose furrows: precoxal suture very broad anteriorly. $30-36$ antennal segments
	Host: Cerodontha (Butomomyza) angulata LOEW
	Species of more elongate form (thorax $1.3-1.5$ times as long as high), with longer hind tarsi $(0.9-1.0$ times as long as the hind tibiae) $\ldots \ldots \ldots \ldots \ldots 3$
3	Notaulices complete, reaching the posterior fovea as deep conspicuous furrows which are usually rugose throughout. Central part of mesepisternum (above the broad pre- coxal suture) covered with fine scaly-reticulate ground sculpture. Antennae with 38-45 segments: all flagellar segments entirely covered with short dense pubescence in both sexes
	Notaulices usually not so well developed. Central part of mesepisternum completely
	smooth $\ldots$
4	Central part of mesoscutum densely pubescent, roughened anteriorly. All flagellar segments covered with dense pubescence in both sexes <i>C. merella</i> (NIXON) Hosts: Cerodontha (Butomomyza) caricivora GROSCHKE, C. (B.) scirpi KARL, C. (B.) staryi HERING, C. (Dizygomyza) caricicola HERING, C. (D.) luzulae GROSCHKE and C. (D.) chaixiana GROSCHKE
	Central lobe of mesoscutum smooth and bare. Basal flagellar segments strongly shin- ing and almost bare in the male, bearing only a few short hairs (mainly near their apex)
5	Mesosternum ventral to the precoxal suture smooth and strongly shining. Petiole with rather coarse, rugose sculpture
	Hosts: Cerodontha (Butomomyza) eucaricis NOWAKOWSKI, C. (B.) vigneae NOWAKOWSKI, C. (Dizygomyza) morosa MEIGEN and C. (D.) sp. probably caricicola HERING

## 3. The Chorebus affinis group

I have not seen material of the species described by BURGHELE (1959a and 1959b), DOCAVO ALBERTI<sup>6</sup> (1962 and 1965) and TOBIAS (1962), which are included in this key on the basis of their descriptions. The host records for *Hydrellia* (Ephydridae) are mainly based on BURGHELE's work. She records the host as *Hydrellia griseola* FALLÉN in every case, but I think that the identity of the host species requires further investigation, as her mass collecting method can hardly have allowed a separation of different species of *Hydrellia*. It would not be surprising if some of the parasites concerned proved to be host vicariants.

1	Mesoscutum with its dorsal surface completely bare except for a few hairs along the course of the notaulices. Sides of pronotum strongly shining and almost bare.	
	Back of head largely bare	<b>2</b>
	Not as above. Mesoscutal pubescence more extensive, extending at least onto the anterior part of the central lobe	11
2	Metapleuron with a bare, shining appearance, the rosette of pubescence around the swelling on its lower half being extremely fine and inconspicuous. Vein $Cu_{1b}$ represented only by a shadow, so that cell $2Cu$ is open at its lower distal corner (fig. 197). 21-24 antennal segments	nov,
	Host: Cerodontha (Icteromyza) geniculata FALLÉN	
	Metapleural pubescence forming a conspicuous rosette. Cell $2Cu$ closed by vein $Cu_{Ib}$ at its lower distal corner	3
3	Petiole with dense pubescence along its sides (but bare along its centre-line). Basal flagellar segments almost bare in the male. $28-34$ antennal segments. Ovipositor ( $\mathfrak{P}$ ) stout, projecting beyond the apical tergite in the retracted position (see Nixon, 1040 for 202).	
	1949, fig. 297)	nov. 4
4	Pronotum entirely bright testaceous. $26-27$ antennal segments. Ovipositor ( $\mathfrak{P}$ ) stout, projecting beyond the apical tergite in the retracted position	*
	Pronotum black, like the rest of the thorax	<b>nov.</b> 5
5	Labial palpi 3-segmented. Basal flagellar segments pubescent in both sexes: in the female the flagellum is conspicuously thickened between roughly the third and	0
	tenth segments	
6	Labial palpi 4-segmented. Flagellum not unusually thickened Eyes $(\mathfrak{P})$ very strongly convergent below, only very narrowly separated above the clypeus (see TOBIAS, 1962, fig. 38). Petiole elongate, about $2^1/_2$ times as long as wide.	6
	29 antennal segments ( $\mathfrak{P}$ ). ( $\mathfrak{F}$ unknown) C. ophthalmicus (TOBIAS), comb.	nov.
7	Eyes not so strongly convergent below, well separated above the clypeus $31-35$ antennal segments ( $\mathfrak{P}$ ). Ovipositor ( $\mathfrak{P}$ ) very stout, strongly projecting	7
	beyond the apical tergite in the retracted position, its sheaths being obviously longer than hind tarsal segment 1 (see NIXON, 1949, fig. 296). Tergite 3 with an area of striation centrally at its base (adjacent to the petiole). (3 unknown)	
	$\ldots \ldots C.$ foveolus (Haliday), comb.	nov.
	Fewer antennal segments. Ovipositor $(\mathfrak{Q})$ more slender, only slightly projecting beyond the apical tergite in the retracted position except in <i>risilis</i>	8

<sup>6</sup> After completion of this paper a further work by Dr. DOCAVO has appeared (DOCAVO ALBERTI, 1967). This includes a very detailed description of *Chorebus costai* DOCAVO, which will supplement the brief original description (DOCAVO ALBERTI, 1962).

8 9	Surface of vertex, mesoscutum, mesepisternum and gastral tergites covered with fine punctate ground-sculpture. Petiole fairly long, about 1.8 times as long as wide. 25 antennal segments (both sexes): basal flagellar segments almost bare in the male. Ovipositor ( $\mathcal{Q}$ ) short, not projecting beyond the apical tergite in the re- tracted position
	Host: Cerodontha (Dizygomyza) morosa MEIGEN
-	Petiole shorter, more widened towards its apex. $20-26$ antennal segments 10
10	Ovipositor $(\mathfrak{P})$ only slightly projecting beyond the apical tergite in the retracted position (see NIXON, 1949, fig. 295), its sheaths hardly as long as the basal segment of the hind tarsus. Basal flagellar segments almost bare in the male
	Ovipositor ( $\mathfrak{P}$ ) more strongly projecting, its sheaths fully as long as the basal segment of the hind tarsus. ( $\mathfrak{G}$ unknown)
11	Petiole elongate, well over $l^1\!/_2$ times as long as wide $\ . \ . \ . \ . \ . \ . \ . \ . \ 12$
*****	Petiole very short, less than $1^{1}_{2}$ times as long as apically wide $\ldots \ldots \ldots 17$
12	Petiole extraordinarily long and narrow, over 3 times as long as wide, virtually bare. Gaster beyond petiole bright yellow or orange. Sides of pronotum entirely punctate and covered with dense public entirely public entirely public entirely antennal segments: public entire of basal flagellar segments sparse in the male
~~~~	Not as above. Petiole not so extremely narrow
13	Petiole as fig. 208, entirely covered with extremely dense publication which radiates outwards around its apical half. Sides of pronotum largely bare, in strong contrast with the densely public public mesoscutum. Basal flagellar segments densely public public entire in both sexes. $27-39$ antennal segments C. cytherea (NIXON) Hosts: Cerodontha (Dizygomyza) spinata GROSCHKE, C. (D.) luctuosa MEIGEN, C. (D.) hirtae NOWAROWSKI,
	C. (D.) chaixiana GROSCHKE and C. (Poemyza) pygmaea MEIGEN (xenophagy)
~	Pubescence of petiole less extensive, not radiating outwards in this way. $30-32$ antennal segments
14	Basal flagellar segments virtually bare in the male. Mesoscutum entirely covered with dense pubescence. Sides of pronotum virtually bare
-	All flagellar segments densely public publi
15	Pubescence on sides of pronotum dense, but somewhat upstanding, lacking the opaque whitish appearance of <i>fordi</i> and <i>iridis</i> . Petiole with dense, rather long pubescence covering much of its surface, bare only along its centre-line. C.vernalis sp.nov.
	Host: Cerodontha (Dizygomyza) caricicola HERING
	Sides of pronotum with densely matted, opaque whitish pubescence (similar to that of the metapleuron) covering most of their surface. Petiole more sparsely pubescent. 16

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16 Sides of pronotum with the bare area along their dorsal margin virtually smooth. Mesoscutum with dense pubescence extending onto the anterior half or third of its dorsal surface (including both central and lateral lobes). Mandibles clearly 4-toothed (fig. 205). Wing with vein  $R_s$  slightly sinuate (compare fig. 193). C.iridis sp. nov.

Host: Cerodontha (Dizygomyza) iridis Hendel

- Sides of pronotum with the bare area along their dorsal margin sculptured. Mesoscutal public extending onto the anterior part of the central lobe, but the lateral lobes are virtually bare. Mandibles very small, with tooth 2 long and pointed but the additional tooth (tooth 3) represented only by a small projection from its side. Wing (fig. 194) with vein  $R_s$  almost evenly curved. . . . . . . . . C. fordi (NIXON) Host: Cerodontha (Poemyza) lateralis MACQUART

17	Lateral lobes of mesoscutum bare. Sides of pronotum largely bare and shining, with distinct public constrained on the intervention of the segments: $\mathfrak{F}$ , (23)-24-27; $\mathfrak{P}$ , 22-24. Labial palpi 4-segmented. Wing as fig. 198
	Mesoscutum entirely covered with pubescence
18	Petiole densely publicent
	Petiole almost bare. Labial palpi 3-segmented uliginosus group (in part) 21
19	Labial palpi 4-segmented. Wing as fig. 199, with cell $2Cu$ closed below. Antennal segments: $3, 28-30; 9, 25. \ldots \ldots$
	Host: Cerodontha (Icteromyza) calosoma HENDEL
-	Labial palpi 3-segmented. Cell $2Cu$ open below (see BURGHELE, 1959b, fig. 16) or bounded only by a semitranslucent vein (1a). $20-24$ antennal segments 
207	Frons with an oval projection below the front ocellus. Petiole (see STELFOX, 1957, fig. 1) without a delimited apical area
	Frons without such a projection. Petiole apically with a medial area of longitudi- nal striation, delimited by a depression around its edges (see BURGHELE, 1959b, fig. 6)
	Host: <i>Hydrellia</i> sp. (Ephydridae)
21	Mesoscutum and mesepisternum densely covered with large punctures. Ovipositor $(\mathfrak{P})$ not extending beyond the apical tergite in the retracted position $\ldots \ldots \ldots$ . $\ldots \ldots $
-	Mesoscutum and mesepisternum not as above, at most the former finely roughened. 22
22	Ovipositor ( $\mathcal{Q}$ ) (fig. 209) long and stout: female gaster strongly pointed towards its apex. Back of head largely bare, but with some dense pubescence near the base of the mandibles: temples and vertex with only a few scattered hairs
	Ovipositor $(2)$ shorter: female gaster not so strongly pointed apically. Back of head largely covered with short dense pubescence $\ldots$ $\ldots$ $\ldots$ $23$
	mean margery converse with anore dense purescence

<sup>7</sup> Gyrocampa stagnalis HEYMONS (1908) is probably a prior name for one of the two species in this couplet, as stated by BURGHELE (1960).

23<sup>8</sup> Legs with the tibiae, tarsi and hind coxae strongly infuscated. Mesoscutum with a distinct posterior fovea along its centre-line . . . . . . . . C. uliginosus (HALIDAY) (= Gyrocampa thienemanni RUSCHKA)

Hosts: Hydrellia spp.

- Legs largely clear testaceous with only segment 5 of the hind tarsi distinctly infuscated: coxae only slightly darkened. Posterior fovea absent . . . C. costai DOCAVO

## 4. Various plesiomorph species of Chorebus

This key includes the species included in the key in Part I (GRIFFITHS, 1964, page 878), which may now be regarded as superseded. All species have been seen by me except C. geminus (TOBIAS) and C. singularis (TOBIAS). However Mr. TOBIAS has confirmed in correspondence that he thinks these correctly included in the key.

	Metapleuron (fig. 19) only feebly sculptured on its lower half, with two or three long hairs. $20-21$ antennal segments. Maxillary palpi brown, very short. Coxae, hind femora and hind tibiae darkened
5	Host: Phytomyza rufipes MEIGEN 28-34 antennal segments

Hosts: Cerodontha (Poemyza) pygmaea MEIGEN, C. (P.) incisa MEIGEN and C. (P.) tatrica NOWAKOWSKI

<sup>8</sup> Due to my imminent departure from England to Canada I have not had time to examine the lectotype of *Alysia (Dacnusa) uliginosa* HALDDAY (1839), and the interpretation here offered therefore requires checking. Since HALDDAY stated that the head of his species was finely pubescent ("subtiliter pubescens"), I am assuming that he had before him a species known to me as a *Hydrellia*-parasite. Probably this is the same as the species bred from *Hydrellia* which BURGHELE (1959a) has accepted as *uliginosus*.

	Tergite 3 without sculpture
7	Mandibles either with tooth 4 reduced, smaller than tooth 3 (fig. 165), or with both teeth 3 and 4 reduced (fig. 166). Metapleural pubescence as fig. 21. Propodeal pubescence dense, usually rather short. Petiole more or less parallel-sided. Elongate species with the thorax $1.4-1.7$ times as long as high. Hind tarsus about as long as the hind tibia
8	Hind coxa conspicuously rugose, at least at its base: metapleuron as in <i>talaris</i> (com- pare fig. 20), with a well-defined rugose swelling clothed with long, fairly dense pub- escence. Petiole entirely covered with fine pubescence. 29–35 antennal segments 
9	Hind coxa without any conspicuous sculpturation
 	Legs largely dark. Palpi and labrum infuscated
	Legs with at least the tibiae and femora bright golden- or orange-yellow (but with the coxae contrastingly dark in <i>nydia</i> and <i>coxator</i> ). Palpi yellow
— 13	Coxae dark. Metapleural pubescence sparser (fig. 17)
	Tarsal segments $1-4$ yellow, but segment 5 contrastingly black. C. coxator (THOMSON) Hosts: Agromyza phragmitidis HENDEL and A. hendeli GRIFFITHS
14 —	31-33 antennal segments; palpi long and yellowish C. singularis (TOBIAS), comb. nov. Antennae short, with $21-24$ segments; maxillary palpi short (see table 23), always infuscated
15	Antennal segments: $9, 23-24$ ( $\sigma$ unknown). Mesoscutal pubescence covering the entire central lobe and sometimes extending onto the lateral lobes. Metapleuron slightly rugose on its lower half but without a distinct raised swelling (its pubescence similar to that shown in fig. 21 but slightly denser) C. asphodeli GRIFFITHS Host: Liriomyza asphodeli SPENCER
	Antennal segments: $3, 21-22$ (9 unknown). Mesoscutal pubescence less extensive, the central lobe being pubescent anteriorly only or almost bare. Metapleuron with a distinct rugose swelling on its lower half, its pubescence as in <i>asphodeli</i>
	Host: Liriomyza buhri DE MEIJERE

# 5. The Chorebus ovalis/lateralis complex

(Key 6 below is a supplementary key to females of this group in which the ovipositor distinctly projects beyond the apical tergite in the retracted position.)

All species included in this key have been seen by me with the exception of C. oltenicus (BURGHELE) and C. interjectus (TOBIAS). However the authors of these species have confirmed in correspondance that they think them correctly included in the key.

1	Precoxal suture represented by a short smooth impression of absent. Small species (less than 2 mm.), with unusually short antennae, the flagellar segments being very short (mostly not more than $1\frac{1}{2}$ times as long as wide), and few in number (3, 19; $\wp, 15-17$ ). Legs entirely dark. Wing (fig. 179) with an unusually broad pterostigma.
	Host: Liriomyza lulea MEIGEN
$\frac{-}{2}$	Rugose precoxal suture present
	bare except for a few hairs along the course (or former course) of the notaulices, i.e. both its central and lateral lobes are virtually bare
	Pubescence of mesoscutum more extensive, extending on its dorsal surface at least over the anterior part of the central lobe
3	Small species, $1.6-2$ mm., with a subcubical head (less than 1.5 times as wide as long) and elongate thorax. Mandible not expanded, with tooth 2 long and pointed (NIXON, 1943. fig. 37). Petiole bare centrally but with a thin accumulation of pubescence at its apical corners (NIXON, 1946, fig. 215). Basal antennal segments contrastingly yellow: legs yellow
	Host: Cerodontha (Cerodontha) fulripes MEIGEN
	Note. Some specimens of <i>crenulatus</i> (see couplet 72) may be taken to this couplet. This species has a subcubical head $(1.5-1.6 \text{ times}$ as wide as long), but clearly differs from <i>diremtus</i> by its darker coloration and in having its petiole entirely pubescent with distinct apical tufts.
4	Head more transverse (over 1.5 times as wide as long) $\dots \dots \dots \dots \dots \dots \dots \dots \dots \dots 4$ Petiole parallel-sided, $2\frac{1}{2}$ times as long as wide, with evenly distributed pubescence covering its entire surface. 36 an nnal segments ( $\mathcal{Q}$ ). Legs largely yellow, but with the base of the hind coxae infuscated. Ovipositor ( $\mathcal{Q}$ ) projecting beyond the apical tergite in the retracted position $\dots \dots \dots$ (BURGHELE), comb. nov.
	Petiole not more than twice as long as wide. Ovipositor $(2)$ not projecting beyond the apical tergite in the retracted position $\cdot \cdot \cdot$
5	Petiole bare or sparsely pubescent, without distinct whitish tufts of pubescence at its apical corners, although in some species ( <i>lugubris, hilaris, solstitialis,</i> and <i>flavipes</i> ) the pubescence usually becomes slightly denser here
	Petiole with distinct whitish tufts of pubescence at its apical corners 11
6	$18-24$ antennal segments $\ldots$ $\ldots$ $\ldots$ $\ldots$ $\ldots$ $.$
	At least 26 antennal segments
7	Basal antennal segments contrastingly yellow at least as far as the third flagellar segment (often about as far as the sixth flagellar segment). Legs usually entirely yellow (rarely with the coxae brown) C. albipes (HALIDAY) Hosts: Phylagromyza populi KALTENBACH, P. tremulae HERING and P. tridentata LOEW
	Antennae entirely dark. Legs dark brown with all coxae black $C.$ lar (MORLEY) (= Dacnusa innana NIXON)

Hosts: Agromyza johannae DE MEIJERE and A. genistae HENDEL

- 9 Maxillary palpi very short, with their apical segment contrastingly darkened. No-taulices deep, extending more or less to the posterior fovea. Legs bright yellowish red with at most the two apical segments of the tarsi blackened. Antennal segments: 3, 33; 2, 29-32. Petiole almost bare. . . . . . . . . . C. glabriculus (THOMSON) (= Dacnusa cortipalpis NIXON)
- 10 Legs yellow. Antennal segments: ♂, (28)-30-32; ♀, 29-32: basal two or three flagellar segments usually yellow or yellow-brown. Petiole with pubescence distributed over most of its surface (except for the central part of about its apical half), although this is very fine and inconspicuous.
   C. flavipes (GOUREAU) (= Dacnusa raissa NIXON)

Host: Cerodontha (Dizygomyza) iraeos Robineau-Desvoidy

- Legs varying from deep yellow to light brown or testaceous, either unicolorous or with the hind coxae somewhat darker (more or less brown). Antennal segments:  $\mathcal{J}$ , (27)-28-31;  $\mathcal{Q}$ , 26-28-(29): flagellum usually dark throughout, but sometimes becoming obscurely yellow towards its base. Petiole similar to that of *lugubris* and *hilaris*, but slightly more pubescent, with some scattered hairs near its base, as well as along its sides and towards its apex . . . . . . . . . . C. fallaciosae GRIFFITHS Hosts: *Phytomyza fallaciosa* BRISCHKE and *P. rydeni* HEEING
- 11 Extremely small species (about 1.4 mm.) with 21-23 antennal segments. Petiole entirely covered with dense publication with only their lateral extensions distinct. Legs deep yellow. . . C. melanophytobiae GRIFFITHS (see also couplets 88 and 94)

Host: Melanophytobia chamaebalani HERING

- 12 Mandible almost 3-toothed, with tooth 3 developed as no more than a feeble angulation on the side of tooth 2. Antennae with 30-33 (3) and 29-31 (9) segments, yellow-brown at their base as far as the first or second flagellar segment. Petiole with fairly dense, but fine and inconspicuous pubescence distributed over most of its surface but tending to be absent from the centre-line apically. Legs deep yellow (occasionally with the hind coxae brown). . . . . . . . . . . . C. xanthaspidae GRIFFITHS. Host: Metopomyza xanthaspida HENDEL

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(see also couplet 34)

	Mandible with tooth 3 distinct, although relatively small in <i>dirona</i> (fig. 183). Pe- tiole bare centrally (see NIXON, 1945, fig. 198)
_ 14	At least 32 antennal segments: basal two or three flagellar segments contrastingly yellow or yellow-brown. Legs (including hind coxae) yellow
	Host: Cerodontha (Poemyza) deschampsirae SPENCER Gaster uniformly dark, or at most with tergites 3 and 4 reddish brown. Mesoscutum entirely covered with fine scaly-reticulate ground sculpture C. ganesa (NIXON) Host: Cerodontha (Poemyza) deschampsiae SPENCER
15	Head quadrate, hardly wider than long (NIXON, 1943, fig. 56). Large species, 3.8 to 4.2 mm., with very numerous antennal segments $(\mathcal{J}, 49-52; \mathcal{Q}, 42-45)$ . Petiole with distinct apical tufts of pubescence, but otherwise only sparse scattered hairs
	Head transverse, much wider than long, except in <i>crenulatus</i> (see couplet 72) 16 Mandibles large, much widened towards their apex, with at least tooth 1 much ex- panded (compare fig. 57). Petiole with evenly distributed pubescence (rather sparse except in <i>atis</i> ) or bare centrally ( <i>iphias</i> ) <i>atis</i> group 17 Mandibles smaller, not or only slightly widened towards their apex 21
	Note. Some species of the <i>lateralis</i> group (e.g. <i>eros</i> , fig. 60) in which the mandibles are slightly expanded are included under this second alternative.
17	Legs largely dark       18         Legs largely yellow or orange-testaceous, at most with the base of the hind coxae and the apical tarsal segments infuscated       20
18	41 antennal segments (3). Large species, about 2.8 mm., with the swelling of the temples very exaggerated (head width about 1 mm.) and the mandibles about 0.3 mm. wide. Maxillary palpi very short, the two apical segments clearly shorter than the scape $+$ pedicel (see table 8 of biometric data) C. atis (NIXON)
— 19	At most 36 antennal segments $\dots$ 19 34-36 antennal segments $(\mathcal{J})$ . Mesoscutum pubescent except for the posterior half of the lateral lobes. Petiole almost parallel-sided, about twice as long as wide (fig. 71) $\dots \dots \dots$
	29 antennal segments ( $\mathfrak{F}$ ). Mesoscutum almost entirely pubescent. Petiole some- what widened towards its apex, only about 1.7 times as long as wide (TOBIAS, 1962, fig. 58)
	Host: Agromyza baetica GRIFFITHS Note. C. solstitialis (STELFOX), which differs from the species included in this couplet in having an almost bare mesoscutum, has been included in couplet 8.
20	Petiole almost parallel-sided, twice as long as wide, bare centrally but with some

	Petiole widened towards its apex, about $1\frac{1}{2}$ times as long as wide, with evenly dis- tributed pubescence. Antennal segments: $3, 29; 9, 25-26$
	· · · · · · · · · · · · · · · · · · ·
21	Petiole elongate, at least twice as long as wide, almost or completely parallel-sided, virtually bare (at most with a little fine pubescence at its base and a few isolated hairs along its sides and near its apex)
	Petiole more pubescent or, if virtually bare, not elongate and parallel-sided 24
	Note. C. perkinsi (NIXON) and C. knautiae GRIFFITHS could possibly be misplaced at this couplet (see fig. 67).

- 22 42 antennal segments (φ). Petiole extremely narrow, about 3 times as long as wide.
   Legs largely yellow: basal flagellar segments yellow.
   *C. resa* (NIXON) Host: Agromyza sp. (ambigua group)

- Dorsal surface of mesoscutum with only sparse public public

- 24 Elongate species (thorax 1.3-1.6 times as long as high), with an extraordinarily long upcurved ovipositor ( $\mathfrak{Q}$ ), projecting beyond the apical tergite by about half the gastral length in the retracted position, its sheaths about equal in length to the hind tibia. Mandibles (fig. 134) with tooth 4 reduced, obviously smaller than the large tooth 3. Petiole strongly widened towards its apex, only 1.1-1.4 times as long as wide, with most of its surface covered with short, adpressed, fairly dense pubescence. Antennal segments:  $\mathfrak{Z}, 30-34; \mathfrak{Q}, 27-31 \ldots \ldots \ldots C.$  crassipes (STELFOX) Host: Phytomyza diversicornis HENDEL
- Ovipositor (\$\overline\$) shorter except in *bensoni* (couplet 36) which is a short-bodied species (thorax about 1.1 times as long as high). Mandibles usually with tooth 4 well developed (except in *tenellae*)

25	Antennal segments: $3, 21-22; 9, 18-19$ . Petiole partly bare or with evenly distributed public publ
26	Petiole raised centrally, rather coarsely rugose, not strongly shining, with short hairs rather densely distributed over almost its entire surface C. sativi (NIXON) Host: Phytomyza atricornis MEIGEN
	Petiole almost flat, strongly shining, its shallow sculpture having a distinct longi- tudinal element, bare centrally but with a few hairs near its sides and apical corners (fig. 143)
27	Hind coxa with a distinct matted tuft of whitish pubescence at its base. Thorax short, $1.1-1.2$ times as long as high. $24-32$ antennal segments. Legs yellow or ochreous. Petiole either parallel-sided, about twice as long as wide, and densely pubescent ( <i>nana</i> ); or, if widened towards its apex, densely pubescent on its basal half only ( <i>thecla</i> )
	Pubescence of hind coxa usually not forming a distinct matted tuft. Species in which a weak tuft is present are of much more elongate form ( <i>ergias</i> , <i>oritias</i> and <i>fallax</i> ), or with a different form of petiole (e.g. <i>tamiris</i> and <i>calthae</i> )
28	Petiole (fig. 145) widened towards its apex, densely public public on its basal half but more sparsely public public towards its apex. Antennal segments: $\sigma$ , 28–31; $\varphi$ , (24) – 25–27: flagellum entirely dark or only obscurely paler towards its base 
	Hosts: Phytomyza lithospermi NOWAKOWSKI and Liriomyza sp. on Hieracium pilosella
	Petiole parallel-sided, about twice as long as wide, with dense whitish pubescence distributed over its entire surface and tending to form apical tufts. Antennal segments: $\mathfrak{F}$ , $(28) - 29 - 32$ ; $\mathfrak{P}$ , $27 - 32$ : basal segments usually yellow-brown (clearly so as far as about the first flagellar segment)
	bescence on the hind coxa, has been included in couplet 23 on account of its elongate, virtually bare petiole.
29	38-45 antennal segments. Petiole strongly widened towards its apex, only 1.2 to 1.6 times as long as apically wide, largely bare (fig. 63) <i>lateralis</i> group (part). 30 Fewer antennal segments, or the petiole not of this type
30	Gaster beyond petiole reddish yellow. Legs bright yellow-red throughout
	Gaster dark brown. Legs largely yellow, but with the hind tibiae infuscated on about their apical third
31	Petiole wholly or partly bare (at most with fine pubescence at its base, along its sides and towards its apex), usually strongly widened towards its apex (not more than 1.5 times as long as wide in most species, at most 1.8 times as long as wide in <i>rubi- cundus</i> ): never with distinct whitish apical tufts. Not more than 35 antennal seg- ments

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	Petiole with more extensive pubescence, usually covering its entire surface: if the pubescence tends to be absent from the centre-line or apical half, then the petiole is relatively elongate (more than 1.5 times as long as apically wide) or bears distinct
32	whitish apical tufts
	surface (fig. 62). $27-34$ antennal segments. Notaulices well developed. Legs yellow, at most with the tarsi partly infuscated $\ldots \ldots \ldots \ldots \ldots \ldots 33$ Mandibles as fig. 135, with tooth 2 long and pointed, but tooth 3 weak. (Petiole only
	slightly widened towards its apex)
	Mandibles with tooth 3 well developed. Petiole not completely bare, at least with a
33	little fine pubescence near its base and along its sides
	Host: Agromyza distorta GRIFFITHS
	Mandible with tooth 3 visible as a small projection from the side of tooth 2 (fig. 182). Oblique bands of propodeal pubescence inconspicuous, not white and matted 
	Host: Liriomyza virgo Zetterstedt
34	Maxillary palpi very short, with their apical segment contrastingly blackened. No- taulices deep, extending more or less to the posterior fovea. Legs bright yellowish red with at most the two apical segments of the tarsi blackened. Antennal segments: 3, 33; 9, 29-32. (Dorsal surface of mesoscutum largely bare, at most with fine in- conspicuous pubescence on the central lobe) C. glabriculus (THOMSON) (= Dacnusa cortipalpis NIXON) (see also couplet 9)
	Maxillary palpi more or less uniformly coloured, the apical segment not contrast-
35	ingly blackened $\dots \dots \dots$
	Host: Cerodontha (Poemyza) pygmaea MEIGEN
	Petiole usually not more than 1.5 times as long as wide, strongly widened towards its apex
36	Ovipositor $(\mathfrak{P})$ extremely long, projecting beyond the apical territe in the retracted position by almost half the gastral length, its sheaths almost as long as the hind tibia. Antennal segments: $\mathfrak{F}, 33-35$ ; $\mathfrak{P}, 30-33$ . Petiole broadly bare along its centre-line. Legs yellow
37	Ovipositor $(Q)$ not or only shortly projecting beyond the apical tergite in the retracted position. Fewer antennal segments (considering each sex separately) except in <i>tamiris</i> , which has a more densely public except periode
	Host: <i>Paraphytomyza xylostei</i> ROBINEAU-DESVOIDY Note. This species differs from <i>C. albipes</i> (HALIDAY) (couplet 7) in having longer palpi, a shorter petiole, and a more densely pubescent mesoscutum. Occasional specimens of the latter species which have a little pubescence on the central lobe of the mesoscutum might be taken to this couplet.

	Larger species with the basal antennal segments not contrastingly pale 38 Mandibles with tooth 4 small, close to their base (fig. 137). Antennal segments: $\delta$ , 28-31; $\varphi$ , (25)-26-28. Legs golden yellow or yellow-brown <i>C. tenellae</i> GRIFFITHS Host: <i>Phytomyza tenella</i> MEIGEN
	Mandibles with tooth 4 normally developed
40	Fewer antennal segments (at most 27 in the male and 26 in the female) 40 Gastral tergites 3 and 4 contrastingly yellow-brown. Legs 1 and 2 ochreous yellow or yellow-brown; hind legs somewhat darker, more or less red-brown: coxae not con- trastingly infuscated. Petiole finely pubescent on about its basal half, but almost bare on its apical half. Ovipositor (?) not projecting beyond the apical tergite in the retracted position
	Gaster beyond petiole uniformly dark. At least the hind coxae distinctly infuscated, dark brown or black
	Petiole usually less strongly widened towards its apex, its pubescence sparse but usually distributed over most of its surface. Female gaster more or less rounded apically: ovipositor short, not projecting beyond the apical tergite in the retracted position
	Elongate species (thorax 1.4–1.5 times as long as high) with long, largely yellow legs: hind tarsi as long as or longer than the hind tibiae, with segment 3 clearly lon- ger than segment 5. Petiole about twice as long as wide, almost parallel-sided, enti- rely covered with dense pubescence which becomes denser towards the apical cor- ners (forming weak tufts). $30-35$ antennal segments $\ldots \ldots \ldots \ldots \ldots \ldots 43$ Not as above. If the thorax is elongate, the hind tarsi are clearly shorter than the hind tibiae (e.g. asramenes and agraules), or the form of the petiole different $\ldots 45$ Mandible with tooth 2 very long and curved outwards (fig. 136). Ovipositor ( $\mathfrak{P}$ ) di- stinctly projecting beyond the apical tergite in the retracted position. <i>C. fallax</i> (NIXON) Host: <i>Phytomyza cardui</i> HERING
 44	Mandibles with tooth 2 not so markedly long (fig. 131)
	Ovipositor $(\mathcal{Q})$ longer, probably projecting beyond the apical tergite in the retracted position

- Mandibles not as above: if tooth 2 is somewhat long and pointed, tooth 3 is better developed (e.g. buhri, fig. 130) or the antennal segments more numerous (e.g. asramenes and agraules)
   47

- 47 Centre of mesepisternum (above the precoxal suture) evenly covered with long hairs, similar to those below the precoxal suture. 34 antennal segments (\$). Pubescence of petiole largely fine and inconspicuous, but forming distinct apical tufts. Legs dull reddish with the hind coxae dark brown . . . C. abrota (NIXON), comb. nov.
  Centre of mesepisternum usually completely bare, at most with a few fine hairs form-

are aphantus and avesta: these have inter alia paler legs and a more strongly rugose metapleural swelling than in bres.
Not as above. If the petiole is similar to that of species of the *lateralis*-group (elon-

	gate and parallel-sided), the number of antennal segments does not exceed $30^{\circ}$ $58$
49	Pubescence of petiole sparse (fig. 67). Gaster beyond petiole predominantly pale (yellow-brown or orange-yellow) with at most suffusions of darker colouring along its
	sides. $34-39$ antennal segments $\ldots$ $\ldots$ $\ldots$ $\ldots$ $\ldots$ $\ldots$ $50$
	Pubescence of petiole denser (figs. 65, 66, 68 and 69)
50	Legs yellow except for the tarsi and occasionally the extreme apex of the hind tibiae. Mesoscutal pubescence sparse (along the course of the notaulices and in about $4-5$ rows over the central lobe)
	Hind legs with brown coxae, the apical half of the femora dark brown and the entire tibiae and tarsi almost black. Mesoscutum with denser pubescense, extending also onto the anterior half of its lateral lobes C. knautiae GRIFFITHS Host: Agromyza weerzi GROSCHKE
51	Legs largely yellow, at most with the tarsi, the apex of the hind tibiae and the base of the hind coxae infuscated

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-52	Mesoscutum bare over much of its surface, with pubescence over its anterior face, along the course of the notaulices and sparsely on the anterior part of its central lobe. $31-36$ antennal segments. Hind tibia entirely yellow (not infuscated api- cally)
	Central lobe of the mesoscutum densely pubescent
-53	Mesoscutum almost entirely pubescent. Metapleural swelling weakly rugose-punc- tate (but more strongly so than in <i>lateralis</i> and <i>deione</i> ). Antennal segments: $3, 34-38$ ; $9, (32)-33-36$ C. trilobomyzae GRIFFITHS Hosts: Trilobomyza labiatarum HENDEL, T. flavifrons MEIGEN and T. verbasci BOUCHÉ
_	Mesoscutal public extending onto the anterior part of the lateral lobes. Meta- pleural swelling coarsely rugose, hardly shining (as in <i>eros</i> ). $34-39$ antennal seg- ments
	Lateral lobes of mesoscutum almost bare. Metapleural swelling of strongly shining appearance, only feebly rugose. Usually at least 36 antennal segments
54	Basal flagellar segments yellow tipped with black. Hind coxae entirely pale yellow 
	Flagellum entirely dark. Hind coxae dark yellow, usually infuscated at their base
55	45-48 antennal segments. Gaster beyond petiole red or orange. Hind femora and tibiae extensively infuscated
	At most 43 antennal segments. Gaster usually entirely dark (at most with tergite 3
56	reddish)
 57	Metapleural swelling shining, only shallowly rugose $\dots \dots \dots$
<b></b>	Antennal segments: $\eth$ , $39-42$ ; $\wp$ , $(35)-37-43$ . Notaulices usually well defined to the middle of the mesoscutum
	Hosts: Agromyza nigrescens HENDEL and A. polygoni HERING
58 	Petiole with pubescence evenly distributed over its entire surface, or becoming sparser on its apical half; not becoming obviously denser at its apical corners 59 Petiole with dense pubescence covering usually its entire surface and becoming den- ser towards its apical corners, often forming conspicuous whitish tufts: if its centre-
59	line is bare (as sometimes in <i>ninella, daimenes</i> and <i>testaceipes</i> ), these apical tufts are well developed $\ldots \ldots \ldots$

	Basal antennal segments contrastingly yellow or yellow-brown as far as about the second flagellar segment. Legs entirely yellow. Palpi long (see table 23). Antennal segments: $3$ , $29-30$ ; $9$ , $(25)-27-29$ . (Pubescence of petiole dense, usually tending to form weak apical tufts, but occasionally these are absent). C. avesta (NIXON) (see couplet 89)	
	Not as above	
60 <sup>9</sup>	<sup>9</sup> Thorax very short, only about 1.1 times as long as high. Pubescence of petiole sparse and inconspicuous	
61	Legs largely deep yellow-brown or testaceous with the coxae and apical tarsal seg- ments (and sometimes also the hind femora) infuscated. Antennal segments: $\Im$ , $25-27$ ; $\Im$ , $(23)-25-26$	
	Host: Phytomyza angelicae KALTENBACH	
	Legs deep yellow or ochreous with only tarsal segments 5 distinctly infuscated (at most with the hind coxae brown). Antennal segments: $3, 28-32-(33); 9, 27-31$ - (32)	
	HOSE, I Rytomyza caenoprata mexing	
62	Wing length about equal to the body length. Head relatively large (head width 1.5times the width of the mesoscutum). Mandibles with tooth 2 rather long and pointed(fig. 130). Antennal segments: $\mathcal{J}, 26-27; \mathcal{Q}, 22-23$ . Palpi fairly short (see table 15).Legs largely ochreous or brown. (Pubescence of petiole usually showing a slighttendency to become denser at the apical corners). $\mathcal{C}$ . buhri GRIFFITHS(see also couplet 76)Host: Phytomyza griffithsi SPENCER	
	Wing length usually clearly longer than the body length (except sometimes in scabiosae). Mandibles with tooth 2 not unusually long and pointed $\ldots \ldots \ldots$	
63	Hind tarsi about as long as the hind tibiae. Antennal segments: $\eth, 22-24; \heartsuit, 21$ -23. Petiole rather densely publicent near its base but more sparsely so towards its apex (sometimes bare centrally on its apical half). Palpi short (see table 15 of biometric data). Small species, $1.4-1.6$ mm C. scabiosae GRIFFITHS Host: Phytomyza scabiosae HERING	
	Hind tarsi clearly shorter than the hind tibiae, except sometimes in <i>punctus</i> (a larger species with more numerous antennal segments)	
64 <sup>1</sup>	<sup>10</sup> Maxillary palpi short (the length of the two apical segments together being only about one third of the height of the head — see table 15). Antennae entirely dark. Legs largely ochreous brown with the coxae somewhat darker. 24 antennal seg- ments $(\mathcal{J})$	
	Maxillary palpi obviously longer (aphantus, sylvestris, xylostellus), or antennal seg- ments more numerous (aphantus, prosper, punctus)	
<sup>9</sup> The measurement of thorax length used throughout this paper is the longitudinal distance from the anterior edge of the mesoscutum to the posterior edge of the propodeum.		

<sup>10</sup> The measurement of head height used throughout this paper is the perpendicular distance from the lower margin of the clypeus to the apex of the vertex (measured in lateral view).

9 Beitr. Ent. 18, H. 1/2

- Maxillary palpi long (the length of the two apical segments together being almost half the height of the head — see table 15). Thorax 1.2—1.3 times as long as high. Number of antennal segments strongly sexually dimorphic
- 67 Antennal segments: δ, (22)-23-25-(26); φ, 19-21: basal flagellar segments contrastingly yellow or orange-yellow (at least as far as the third flagellar segment, often as far as the fifth).
  Host: Phytomyza periclyment DE MEIJERE
- Antennal segments more numerous in the female (and often so in the male): basal flagellar segments dark or at most deep yellow or yellow-brown no further than the third flagellar segment (never so strongly contrasting in colour as in *xylostellus*) . . 68
- 68 Antennal segments: J, (23)-24-27; Q, 22-25 . . . . . C. sylvestris GRIFFITHS Hosts: Phytomyza alpigenae HENDEL, P. lonicerella HENDEL and P. xylostei KALTENBACH

- white apical tufts. Ovipositor ( $\mathfrak{Q}$ ) not projecting beyond the apical tergite in the retracted position. Wing somewhat iridescent, with vein  $R_s$  evenly curved (fig. 189) or only weakly sinuate. Antennal segments:  $\mathfrak{Z}, 28-31; \mathfrak{Q}, (26)-28-30$ .... Host: Cerodontha (Poemyza) superciliosa ZETTERSTEDT

130

- 71 Ovipositor (φ) clearly projecting beyond the apical tergite in the retracted position (NIXON, 1945, fig. 197), slightly upcurved. 28-31 antennal segments (φ). Wings somewhat iridescent (compare vitripennis). . . . . . C. crocale (NIXON), comb. nov.
- Ovipositor (φ) not projecting beyond the apical tergite in the retracted position.
   33 antennal segments (both sexes). Wings not unusually iridescent . . C. merion (NIXON) Host: Phytomyza taraxacceccis HERING
- 72 Small species (about 2 mm.) with large subcubical head (1.5-1.6 times as long as wide) and elongate thorax. Dorsal surface of mesoscutum with pubescence only along the course of the notaulices and sparsely on its central lobe (occasionally this too is almost bare): notaulices well defined, reaching the posterior fovea. Mandibles small, with tooth 2 long and pointed. Antennal segments: 3, 30-34; 9, 26 30. Legs largely reddish yellow or yellow-brown. . . . C. crenulatus (THOMSON) (= Dacnusa elegantula NIXON)

(see also couplet 3)

#### Host: Cerodontha (Cerodontha) denticornis PANZER

73 73 Elongate species (thorax about 1.4 times as long as high) with at least 31 antennal segments. Legs largely deep yellow, at most with the hind coxae brown, Mesoscutum with dense pubescence covering its entire surface except the posterior half of the lateral lobes. Hind tarsus slightly shorter than the hind tibia. Mandibles (fig. 74- Elongate species (thorax about 1.4 times as long as high) with not more than 29 antennal segments. Legs dark, their dominant colour varying from yellow-brown to almost black: hind coxae always virtually black. Hind tarsus not or only slightly 75- Not as above. Thorax shorter in most species. Hind tarsus clearly shorter than the 7674 Ovipositor ( $\mathfrak{Q}$ ) not projecting beyond the apical tergite in the retracted position  $\ldots$ Host: Cerodontha (Poemyza) pygmaea MEIGEN - Ovipositor ( $\varphi$ ) strongly projecting beyond the apical tergite in the retracted position

- Ovipositor ( $\mathfrak{Q}$ ) not or hardly projecting beyond the apical tergite in the retracted position. 19-23 antennal segments. Hind tarsus not longer than hind tibia, usually slightly shorter. Legs uniformly blackish brown. *C. anasella* (STELFOX), **comb. nov.** Host: *Phytomyza plantaginis* ROBINEAU-DESVOIDX<sup>11</sup>
- 76 Wing length about equal to body length. Head relatively large (head width 1.5 times the width of the mesoscutum): thorax rather elongate, about 1.3 times as long as high. Mandibles with tooth 2 rather long and pointed (fig. 130). Legs largely ochreous or brown. Antennal segments: 3, 26-27; 9, 22-23. C. buhri GRIFFITHS (see also couplet 62)

Host: Phytomyza griffithsi Spencer

<sup>11</sup> New host record: details will be given in a later paper.

9\*

- Wing length usually clearly longer than the body length (except sometimes in poemyzae and melanophytobiae). Mandibles with tooth 2 not unusually long and pointed (figs. 132 and 133)
- 77 Ovipositor (\$\overline\$) and sheaths shortly projecting beyond the apical tergite in the retracted position: when extruded the ovipositor itself can be seen to be strongly downcurved (NIXON, 1945, fig. 193). Hind tarsus not or only slightly shorter than the hind tibia. Antennal segments: \$\delta\$, 28-32; \$\overline\$, 27-31. Legs dull yellow or yellow-brown with the hind coxae somewhat infuscated, at least at their base. Central lobe of mesoscutum roughened; notaulices well developed . . . . . . . C. kama (NIXON) Host: Phytomyza ranunculi SCHRANK

- 79 Palpi somewhat short (NIXON, 1945, fig. 185). In the female the gaster appears somewhat pointed towards its apex in dorsal view; apical sternite pointed (NIXON, 1945, fig. 177). Antennae with 32-35 (3) and 30-34 (9) segments, the basal segments being more or less dark or at most pale ventrally. *C. ovalis* (MARSHALL), comb. nov.

(GRIFFITHS, 1966a) requires clarification.
81 Eyes strongly convergent below, the minimum distance between them being only about two-fifths of the width of the head (NIXON, 1945, fig. 168). Antennae with 21-26 segments, unusually short (with most of the flagellar segments less than twice as long as wide). Palpi short (see table 23). Legs largely ochreous yellow or

<sup>12</sup> This character is subject to confirmation in the case of *apollyon*, whose female is unknown.

82 Wing (fig. 89) with the pterostigma and cell  $2R_I$  very elongate. Thorax short, only about 1.1 times as long as high: hind tarsus short (only about 0.8 times the length of the hind tibia). Petiole strongly widened towards its apex, only 1.3-1.5 times as long as apically wide. Mandibles (fig. 133) with tooth 2 somewhat blunt, not projecting much beyond tooth 3. Antennal segments: 3, 28-30; 9, 24-27. Legs dull yellow or light brown with the coxae and tarsi somewhat darker (brown or dark Host: Phytomyza calthivora HENDEL 83 83 Sides of pronotum with only fine inconspicuous pubescence, which hardly obscures 84 Sides of pronotum with denser, more conspicuous pubescence at least along and below the oblique suture. Petiole strongly widened towards its apex, 1.3-1.6 times 9484 Very dark species with all coxae blackened and at least the hind femora strongly infuscated (dark brown or black): palpi brown or virtually black . . . . . . . . . 85 - Paler species, at most with yellow-brown legs and the hind coxae infuscated: palpi 87 85 26 antennal segments ( $\varphi$ ). Petiole strongly widened towards its apex, about 1.4 times as long as wide, with conspicuous white apical tufts . . . C. pimpinellae GRIFFITHS Host: Phytomyza melana HENDEL - Antennal segments usually more numerous. Petiole more elongate, 1.8-2.0times as long as wide, its pubescence becoming denser towards the apical corners 86 Pubescence of petiole long, hardly adpressed. Antennal segments:  $\vec{\sigma}$ , (29) -30-33; (see also couplet 59) Host: Phytomyza campanulae HENDEL - Pubescence of petiole shorter, more adpressed (as in alecto and armida). 32 anten-87 Small species (1.4-1.7 mm.) with yellow legs (at most the hind coxae brown basally) and the petiole strongly widened towards its apex (less than  $1\frac{1}{2}$  times as long as 88 - Petiole more elongate, usually over 1½ times as long as apically wide. Antennal 89 88 Mesoscutum with its entire surface punctate and densely pubescent. Antennae en-Host: Phytomyza crassiseta ZETTERSTEDT - Mesoscutum with its dorsal surface smooth and largely bare, at most with a little pubescence anteriorly. Antennae usually with the scape, pedicel and about the first flagellar segment yellow-brown . . . . . . . . . . . . C. melanophytobiae GRIFFITHS (see also couplets 11 and 94) Host: Melanophytobia chamaebalani HERING 89 Basal antennal segments contrastingly yellow or yellow-brown as far as about the second flagellar segment. Legs entirely yellow. Antennal segments: 3, 29-30; 9, (see also couplet 59) Hosts: Liriomyza morio BRISCHKE and L. eupatoriana SPENCER - Antennae entirely dark, or at most obscurely brownish at their base . . . . . 90

- 90 Antennal segments: 3, (19), 23; 9, 21-22. Thorax about 1.3 times as long as high. Mesoscutal pubescence not extending onto the lateral lobes. Very small species (1.2-1.4 mm.)
  Host: Liriomyza congesta BECKER
- Antennal segments more numerous. Size usually larger . . . . . . . . . . . . . . . . . 91  $91^{13}$ Thorax very short, only 1.1-1.2 times as long as high. Mesoscutum usually almost
- Thorax 1.2-1.3 times as long as high. Mesoscutal publication in the result of the lateral lobes. Antennal segments: 3, 26; 9, 25-26. Maxillary palpi rather short (see table 15) . . . . . . . . . . . C. oreoselini GRIFFITHS Host: Phytomyza pauliloewi HENDEL
- 92 Antennal segments: 3, (27)-28-31;  $\mathcal{G}$ , (26)-27-31 . . . . . . C. armida (NIXON) Hosts: several Phytomyza spp. on Umbelliferae and Compositae
- Antennal segments: 3, (25)-26-28; 9, (23)-24-26-(27) . . . C. alecto (MORLEY) (= Dacnusa turissa NIXON) Hosts: several Phytomyza spp. on Compositae and P. crassiseta ZETTERSTEDT

Host: Melanophytobia chamaebalani HERING

- 95 18-24 antennal segments. Legs yellow-brown or brown with the hind coxae infuscated. Wing length less than 2 mm.
   96 24-31 antennal segments: if 24 (daimenes), the legs (including coxae) are vellow.
   97
- 24-31 antennal segments: if 24 (daimenes), the legs (including coxae) are yellow. 97
   96 Antennal segments: 3, (21)-22-24; 9, 21-22. Pubescence on sides of pronotum conspicuously dense and matted . . . . . . . . . . . . . . . C. misellus (MARSHALL) Hosts: Liriomyza congesta BECKER, L. centaureae HERING, L. cyparissiae GROSCHKE and Phylomyza atricornis MEIGEN (xenophagy)
- Antennal segments: 3, 20-22; 9, 18-20. Pubescence on sides of pronotum not so dense as in misellus, similar to that of daimenes.
   C. artemisiellus GRIFFITHS Hosts: Liriomyza artemisicola DE MEIJERE and L. dracunculi HERING
- 97 Legs (including coxae) uniformly deep yellow. 24-29 antennal segments . . . . . 98
- Legs varying from ochreous yellow to brown, with at least the hind coxae infuscated. 99
- 98 Entire surface of mesoscutum pubescent . . . . . . . . . C. incertus (GOUREAU) Host: Liriomyza pascuum MEIGEN

<sup>13</sup> While populations of the species in couplets 91-93 are clearly separable, the differences found may not be sufficient to enable all isolated specimens to be identified with confidence.

- Hosts: Liriomyza ameona MEIGEN, L. bryoniae KALTENBACH, L. millefolii HERING, L. pascuum MEIGEN. L. pumila MEIGEN, L. sonchi HENDEL and L. strigata MEIGEN Note. The distinction of the above two species is subject to some doubt.
- 99 Entire surface of mesoscutum pubescent. 30-31 antennal segments . . . . . Host: Liriomyza pusilla MEIGEN
- Lateral lobes of mesoscutum largely bare. 26-29 antennal segments  $\ldots$ Hosts: Liriomyza puella MEIGEN, L. pusilla MEIGEN and L. scorzonerae RYDÉN

In most species of the *Chorebus ovalis*/lateralis complex the ovipositor ( $\mathcal{Q}$ ) does not project beyond the apical tergite in the retracted position. But there are a few species in which a more elongate ovipositor has been evolved. These are not all grouped together in the above key, which is intended to apply to both sexes, because many are very dissimilar in other respects. It is evident that an elongate ovipositor has been evolved independently on a number of occasions (i.e. that resemblance in this respect often represents convergence, not synapomorphy). But in order to allow quicker identification of females which show this feature, the following supplementary key has been produced. Couplets 7-10 are partly based on couplets 4-7 of NIXON's (1945) key to his ovalis-group.

## 6. Females of the Chorebus ovalis/lateralis complex in which the ovipositor distinctly projects beyond the apical tergite in the retracted position

1	Ovipositor extremely long, projecting beyond the apical tergite in the retracted po- sition by about half the gastral length
2	Thorax elongate, $1.3-1.6$ times as long as high. Mandibles (fig. 134) with tooth 4 reduced, obviously smaller than the large tooth 3. Petiole with most of its surface covered with short, adpressed, fairly dense public ence. 27-31 antennal segments.
	Thorax short, about 1.1 times as long as high. Mandibles with tooth 4 well developed, not smaller than tooth 3. Petiole broadly bare along its centre-line, with only fine inconspicuous pubescence along its sides. 30-33 antennal segments
3	Dorsal surface of mesoscutum virtually bare except for a few hairs along the course of the notaulices. Petiole parallel-sided, $2\frac{1}{2}$ times as long as wide, with evenly distri- buted pubescence covering its entire surface. 36 antennal segments. Legs largely yellow, but with the base of the hind coxae infuscated C. oltenicus (BURGHELE)
	Pubescence of mesoscutum more extensive, extending on its dorsal surface at least over the anterior part of the central lobe
4	Petiole bare centrally, with only inconspicuous pubescence near its base, along its sides and at its apical corners. Ovipositor only slightly projecting beyond the apical
	tergite in the retracted position
	towards its apical corners, often forming distinct tufts

- Legs darker, largely yellow-brown or red-brown with the hind coxae usually black.
   Palpi somewhat infuscated. Not more than 31 antennal segments . . . . . . 10
- 8 Petiole somewhat widened towards its apex with conspicuous pale tufts at its apical corners; sides of pronotum above the oblique suture evenly covered with fine pubescence. Hind tarsus slightly shorter than the hind tibia. . . . . C. agraules (NIXON) Host: Cerodontha (Poemyza) muscina MEIGEN
- Petiole almost parallel-sided, with its pubescence becoming denser towards the apical corners, but not forming conspicuous pale tufts: sides of pronotum partly bare. Hind tarsi as long as or longer than the hind tibiae . . . . . . . . . . . . . . . . 9
- 9 Mandible with tooth 2 very long and curved outwards (fig. 136) . . C. fallax (NIXON) Host: Phytomyza cardui HEBING

### Keys to the Dacnusini Parasites of particular Host-Groups

Three keys are given below; to the parasites of the subgenera Butomomyza and Dizygomyza, to the parasites of subgenus Poemyza, and to the parasites of subgenus Icteromyza. I have not treated subgenera Butomomyza and Dizygomyza separately, because two of the commonest parasite species occur on hosts of both subgenera. No keys are offered for subgenera Crastemyza and Cerodontha s.s., since only one and two species of parasites are yet known from these hostgroups (as shown in Table 26).

# 1. Cerodontha subgenera Butomomyza and Dizygomyza (on Cyperaceae, Juncaceae, Iris and Gramineae)

1	Metapleural pubescence directed mainly downwards towards the hind coxa, not forming a rosette
*****	Metapleural pubescence forming a rosette of radiating hairs (compare figs. 22 and 23)
2	Pubescence of metapleuron, propodeum and petiole very long and dense, concealing the surface beneath. Tergite 3 smooth. Wing as fig. 123: pterostigma elongate, widened towards its apex, with vein $2r$ branching from its extreme base. Mandible 3-toothed, not unusually long and pointed Dacnusa maculipes THOMSON Oligophagous species
	Metapleuron, propodeum and petiole only sparsely pubescent. Tergite 3 (i.e. the tergite following the petiole) with conspicuous longitudinal striation. Wing as fig. 186 (vein $2r$ arising further from the base of the pterostigma). Mandible (fig. 37) with its central tooth long and pointed, and usually a small additional tooth before this (so that the central tooth becomes tooth 3) $\ldots \ldots \ldots 3$
3	Metapleuron strongly sculptured Laotris striatula (HALIDAY) Host: C. (Dizygomyza) luctuosa MEIGEN
_	Metapleuron with most of its surfaceal most smooth Laotris rupestris sp. nov. Host: C. (Dizygomyza) sp. on Carez sempervirens
4	Precoxal suture represented by a smooth linear groove, which extends virtually for the entire length of the mesepisternum. Hind coxa with a conspicuous crest or basal tuft of matted pubescence on its posterior margin
	Precoxal suture rugose-costate, usually rather broad. Pubescence at the base of the hind coxa not forming a distinct matted tuft
5	Petiole extraordinarily long and narrow, over three times as long as wide, virtually bare. Gaster beyond petiole bright yellow or orange. Sides of pronotum entirely punctate and covered with dense pubescence. Mesoscutum entirely pubescent 
	Not as above. Petiole not so extremely narrow
	Petiole as fig. 208, entirely covered with extremely dense pubescence which radiates outwards around its apical half. Sides of pronotum largely bare, in strong contrast with the densely pubescent mesoscutum Chorebus cytherea (NIXON) Hosts: C. (Dizygomyza) spinata GROSCHKE, C. (D.) luctuosa MEIGEN, C. (D.) hirtae NOWAKOWSKI, C. (D.) chaixiana GROSCHKE and C. (Poemyza) pygmaea MEIGEN (once)
****	Pubescence of petiole less extensive, not radiating outwards in this way 7
	Sides of pronotum largely bare, with only some fine inconspicuous pubescence along and below the oblique suture. Mesoscutum with its dorsal surface completely bare except for a few hairs along the course of the notaulices. Petiole virtually bare. 
	Host: C. (Dizygomyza) morosa MEIGEN
	Sides of pronotum densely pubescent at least on their lower half. Mesoscutum with         more extensive pubescence, extending at least onto the anterior half of its central         lobe       8

- 8 Petiole with dense, rather long pubescence covering much of its surface, bare only along its centre-line. Pubescence on sides of pronotum dense, but somewhat upstanding, lacking the opaque whitish appearance of *iridis*. Wing (fig. 196) with vein R<sub>s</sub> evenly curved.
   Kontent C. (Dizygomyza) caricicola HERING
- Petiole more sparsely publication. Sides of pronotum with densely matted, opaque whitish publication (similar to that of the metapleuron) covering much of their surface. Wing with vein R<sub>s</sub> slightly sinuate (compare fig. 193). Chorebus iridis sp.nov. Host: C. (Dizygomyza) iridis HENDEL

- 10 Mesoscutum with its dorsal surface bare apart from a few hairs along the course of the notaulices. Petiole with only fine inconspicuous pubescence, which does not form distinct apical tufts. Legs yellow. . . . . . . . . . . . Chorebus flavipes (GOUREAU) Host: C. (Dizygomyza) iraeos ROBINEAU-DESVOIDY
- Mesoscutum with dense pubescence covering almost its entire surface. Petiole covered with dense pubescence which forms distinct apical tufts. Legs ochreous yellow or yellow-brown with the hind coxae infuscated . . Chorebus difficilis sp. nov. Hosts: C. (Dizygomyza) chaixiana GROSCHKE and several C. (Poemyza) spp.
- 11 Mandibles (fig. 203) 3-toothed (similar to those of most species of *Exotela* and *Dacnusa*). Wing (fig. 190) with vein R<sub>s</sub> strongly sinuate. Mesoscutum extensively public public public structure is the lateral lobes bare. . . . Chorebus transversus (NIXON) Host: C. (Butomomyza) angulata LOEW

- Species of more elongate form (thorax 1.3-1.5 times as long as high), with longer hind tarsi (0.9-1.0 times as long as the hind tibiae)

- 14 Central lobe of mesoscutum densely pubescent, roughened anteriorly. All flagellar segments covered with dense pubescence in both sexes . . . Chorebus merella (NIXON) Hosts: C. (Butomomyza) caricivora GROSCHKE, C. (B.) scirpi KARL, C. (B.) slaryi HERING, C. (Dizygomyza) caricicola HERING, C. (D.) luzulae GROSCHKE and C. (D.) chaixiana GROSCHKE

### 2. Cerodontha subgenus Poemyza (on Gramineae)

- 1 Mandibles 3-toothed (compare fig. 61), not unusually long and pointed. Metapleuron without any well-defined raised swelling on its lower half, its pubescence directed mainly downwards towards the hind coxa  $\mathbf{2}$ - Mandibles clearly 4-toothed, or, if virtually 3-toothed, the central tooth is very long and pointed. Metapleuron with a raised swelling on its lower half, which in many species is surrounded by a rosette of radiating pubescence . . . . . . . . . . . . 4 2 Wing (fig. 174) with vein 2r eliminated; the distal section of  $R_s$  is bent at its base and fused with  $(\mathcal{J})$  or contiguous with  $(\mathcal{Q})$  the pterostigma . . Dacnusa adducta (HALIDAY) Hosts: C. (P.) pygmaea MEIGEN and Liriomyza flaveola FALLÉN - Wing with vein 2r well developed  $\ldots$   $\ldots$   $\ldots$   $\ldots$   $\ldots$   $\ldots$ 3 3 Wing as fig. 123: pterostigma conspicuously widened towards its apex, with vein 2rbranching from its extreme base. Pubescence of metapleuron, propodeum and petiole very long and dense, concealing the surface beneath. Legs largely brown. 19-25 antennal segments . . . . . . . . . . . . Dacnusa maculipes Thomson Oligophagous species - Wing as fig. 42 (pterostigma not widened towards its apex, with vein 2r arising further from its base). Metapleuron, propodeum and petiole only sparsely pubescent (fig. 16). Legs largely yellow. 25-34 antennal segments . . . . . . . . Hosts: C. (P.) incisa MEIGEN, C. (P.) pygmaea MEIGEN, C. (P.) phragmitidis NOWAKOWSKI, C. (P.) phalaridis NOWAKOWSKI, C. (Crastemyza) flavocingulata STROBL and Agromyza spp. (nigripes group) 4 Precoxal suture represented by a smooth linear groove, which extends virtually for the entire length of the mesepisternum. Hind coxa with a conspicuous crest or basal tuft of matted pubescence on its posterior margin. Pubescence of metapleuron forming a rosette around the swelling on its lower half (see fig. 23) . . . . . 5- Precoxal suture distinctly rugose-costate (except occasionally in Chorebus talaris (HALIDAY)). Pubescence at the base of the hind coxa not forming a distinct matted  $\mathbf{6}$ 5 Petiole as fig. 208, entirely covered with extremely dense pubescence which radiates outwards around its apical half. Sides of pronotum largely bare, in strong contrast Once bred from C. (P.) pygmaza MEIGEN (usually associated with C. (Dizygomyza) spp.) - Petiole partly bare, with only small patches of pubescence near its base, a few fine hairs along its sides and weak accumulations of fine pubescence at the apical corners. Sides of pronotum with densely matted, opaque whitish pubescence covering most of their surface. Mesoscutum with its dorsal surface largely bare, with pubescence only extending onto the anterior part of the central lobe . . . Chorebus fordi (NIXON) Host: C. (P.) lateralis MACQUART 6 Metapleural swelling evenly covered with long, fairly dense pubescence (fig. 20),

- Metapleuron with dense pubescence forming a distinct rosette around the swelling on its lower half (see fig. 22): the centre of the swelling is almost bare or with only a few outstanding hairs
   9

- 9 Petiole almost bare except for a few fine hairs at its apical corners (not forming distinct tufts). Gaster beyond petiole deep orange-yellow. Mesoscutum almost entirely covered with dense pubescence . . . . . . . . . Chorebus rubicundus sp. nov. Host: C. (P.) pygmaea MEIGEN
- Petiole wholly or partly covered with dense pubescence which becomes denser towards its apical corners, often forming conspicuous whitish tufts: if the central part of the petiole is bare (as in *Chorebus ganesa* (NIXON) and *C. enephes* (NIXON)), these apical tufts are well-developed and conspicuous . . . . . . . . . . . . . . . 10
  10 Mesoscutum with its dorsal surface virtually bare apart from a few hairs along the course of the notaulices. Petiole bare centrally, with conspicuously contrasting white apical tufts. Legs largely yellow: basal antennal segments contrastingly

- Gaster uniformly dark, or at most with tergites 3 and 4 reddish brown. Mesoscutum entirely covered with fine scaly-reticulate ground sculpture. *Chorebus ganesa* (NIXON) Host: C. (P.) deschampsiae SPENCER

- Mesoscutum more extensively roughened, with pubescence covering most of its surface (at most absent from the posterior half of the lateral lobes)

- 15 Thorax about 1.3 times as long as high. Publication of petiole becoming denser towards the apical corners, usually forming distinct apical tufts, but these are not so well developed as in *Chorebus asramenes* (NIXON) and *C. agraules* (NIXON). Mandibles with tooth 2 usually not so long and pointed as in *asramenes* and *agraules*. 27-32 antennal segments. Legs largely ochreous yellow or yellow-brown, with the hind coxae infuscated (more or less dark brown): antennae entirely dark.

- Ovipositor (\$\overline\$) strongly projecting beyond the apical tergite in the retracted position (by over half the length of the petiole) . . . . . . . . . . Chorebus agraules (NIXON) Host: C. (P.) muscina MEIGEN

### 3. Cerodontha subgenus Icteromyza

- 1 Mesoscutum with its dorsal surface completely bare except for a few hairs along the course of the notaulices. Metapleuron with a bare shining appearance, the rosette of pubescence around the swelling on its lower half being extremely fine and inconspicuous. Petiole bare. 21-24 antennal segments . Chorebus metallicus sp. nov. Host: C. (I.) geniculata FALLÉN
- Mesoscutum with at least its central lobe densely pubescent. Metapleural pubescence extremely dense, forming a conspicuous rosette around the raised swelling
- 2 Petiole entirely covered with dense pubescence. Vertex, temples and back of head largely covered with short, rather dense pubescence. Sides of pronotum with dense whitish pubescence covering much of their surface. Antennal segments: 3, 28-30;
  \$\overline\$, 25, ..., Chorebus lanigerus (STELFOX)
  Host: C. (I.) calosoma HENDEL

 $\mathbf{2}$ 

### **Host Association**

The known host ranges of the species of Alysiinae treated in this part are given in Table 25. The definitions of the terms used in the classification were explained in Part I (GRIFFITHS, 1964, page 874): the classification relates to normal host selection and is not affected by cases of xenophagy.

The predominance of very restricted host ranges (monophagy of the first or second degree) in the *Chorebus* species associated with *Cerodontha* hosts is in accordance with the conclusions reached in previous parts of this paper. The two species of *Laotris* are also as far as known host specific. Only the *Exotela*, *Dacnusa* and two of the *Dapsilarthra* species are known to attack other genera of Agromyzidae.

Table 26 opposite lists the known host associations of Dacnusini with *Cerodontha* hosts in Europe.

Tribe Dacnusini			
Laotris striatula (HALIDAY)	Monophagy, 1st degree	agraules (NIXON)	Monophagy, 1st degree
rupestris sp. nov.	Monophagy, 1st degree	difficilis sp. nov.	Disjunctive Mono-
Exotela flavicoxa (Thomson)	Disjunctive Mono- phagy (revised from Table 4)	transversus (NIXON) humeralis sp. nov. merella (NIXON)	phagy Monophagy, 1st degree Monophagy, 1st degree Monophagy, 2nd degree
Daenusa adducta (HALIDAY)	Disjunctive Mono-	nobilis sp. nov.	Monophagy, 1st degree
	phagy or Oligophagy,	pelion (NIXON)	Monophagy, 2nd degree
	1st degree	cytherea (NIXON)	Monophagy, 2nd degree
maculipes THOMSON	Oligophagy, 1st degree	stilifer sp. nov.	Monophagy, 1st degree
Chorebus talaris (HALIDAY)	Monophagy, 2nd degree	iridis sp. nov.	Monophagy, 1st degree
subasper sp. nov.	Monophagy, 1st degree	fordi (NIXON)	Monophagy, 1st degree
asperrimus sp. nov.	Monophagy, 1st degree	siniffa (NIXON)	Monophagy, 1st degree
flavipes (GOUREAU)	Monophagy, 1st degree	vernalis sp. nov.	Monophagy, 1st degree
crenulatus (THOMSON)	Monophagy, 1st degree	metallicus sp. nov.	Monophagy, 1st degree
diremtus (NEES)	Monophagy, 1st degree	nigriscaposus (NIXON)	Monophagy, 2nd degree
rubicundus sp. nov.	Monophagy, 1st degree	lanigerus (STELFOX)	Monophagy, 1st degree
enephes (NIXON) ganesa (NIXON) ninella (NIXON)	Monophagy, 1st degree Monophagy, 1st degree Monophagy, 1st degree	gracilipes (THOMSON) Other Alysiinae	Monophagy, 1st degree
poemyzae sp. nov.	Monophagy, 2nd degree	Dapsilarthra sylvia (HALIDAY)	Oligophagy, 1st degree
vitripennis sp. nov.	Monophagy, 1st degree	balteata (THOMSON)	Oligophagy, 1st degree
asramenes (NIXON)	Monophagy, 1st degree	fuscula sp. nov.	? Monophagy

### Table 25

Classification of host ranges of Alysiinae parasites of Cerodontha s.l.

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Hosts	Laotris	Exotela	Dacnusa	Chorebus talaris group and C. ovalis/lateralis complex	Chorebus affinis group and related species
I. Subgenus Butom	omyza				
angulata					humeralis, transver
scirpi					sus merella
staryi	and the second			3 2 5	merella, stilifer
caricivora					merella
eucaricis					nobilis, pelion
vigneae	1	i			pelion
II. Subgenus Dizyge	omyza				
luctuosa	striatula				cytherea
hirtae					cytherea
spin <b>a</b> ta norosa	-	WYO'L' MAR			cytherea
norosa luzulae					pelion, siniffa merella
caricicola <sup>14</sup>					merella, pelion,
					vernalis
sp.	rupestris				
chaixiana				difficilis	merella, cytherea
raeos ridis			maculipes	flavipes	iridis
	1	I	maaar	ŀ	11 60 60
III. Subgenus Cras	temyza	1	1	4	1
flavocingulata		flavicoxa			*******
V. Subgenus Poen	ryza				
deschampsiae				enephes, ganesa,	
			manlines	poemyzae, difficilis	
calamagrostidis muscina			maculipes	ninella, difficilis agraules	
incisa		flavicoxa		talaris, poemyzae,	
				difficilis	
pygmaea		flavicoxa	adducta,	talaris, rubicundus,	cytherea (once)
			maculipes	poemyzae, asramenes,	
				difficilis	
atrica	· ·			talaris, asperrimus	
ulpina				subasper	
uperciliosa				vitrípennis	
ohragmitidis		flavicoxa			
phalaridis		flavicoxa			
lateralis		1			fordi
V. Subgenus Ictero	myza				
enicu <b>la</b> ta			Land Land		metallicus, nigrisc
calosoma					posus, gracilipes
.a10807114					nigriscaposus, lanigerus
VI. Subgenus Ceroo	lontha	t	ł	1	1
enticornis		}		crenulatus	1

# Table 26 List of Records of Dacnusini Parasites of Cerodontha s.1

<sup>14</sup> Host identification requiring confirmation in the case of the records referring to *pelion*.

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### Summary

1. This paper, the sixth of a series, deals with the Alysiinae parasites in Europe of Cerodontha RONDANI in the wide sense first proposed by NOWAKOWSKI (1962). The parasites belong to four genera of Dacnusini (Laotris, Exotela, Dacnusa and Chorebus) and the non-Dacnusine genus Dapsilarthra.

2. Revised keys are given to the European representatives of several groups of *Chorebus* species, including the *affinis* group (a concept here proposed for the first time) and the very difficult *ovalis/lateralis* complex. Keys are also given to the parasites of certain host-groups, to facilitate the identification of bred material.

3. Of the Dacnusini parasites treated in this paper, all the *Laotris* and *Chorebus* species exhibit a high degree of host specificity, but the single *Exotela* and two *Dacnusa* species are also known to attack other genera of Agromyzidae. A complete host/parasite list has again been prepared, including revision of previous records for Europe.

4. Fourteen new species are described, one in *Dapsilarthra*, one in *Laotris* and twelve in *Chorebus*.

### Zusammenfassung

1. Der sechste Teil der vorliegenden Serie beschäftigt sich mit den europäischen Alysiinae-Parasiten der Gattung Cerodontha RONDANI in ihrer von NOWAKOWSKI (1962) vorgeschlagenen Umgrenzung. Die Parasiten gehören zu vier Dacnusini-Gattungen (Laotris, Exotela, Dacnusa und Chorebus) und zu der außerhalb der Dacnusini stehenden Gattung Dapsilarthra.

2. Für verschiedene Gruppen europäischer Chorebus-Arten einschließlich der affinis-Gruppe (die in vorliegender Arbeit erstmalig in dieser Form umgrenzt wird) und für den sehr schwierigen ovalis/lateralis-Komplex werden revidierte Bestimmungstabellen gegeben. Auch für die Parasiten bestimmter Wirtsgruppen werden Tabellen zur Unterscheidung angeführt, um die Determination gezüchteten Materials zu erleichtern.

3. Von den Dacnusini-Parasiten dieses Publikationsteiles zeigen alle *Laotris*- und *Chorebus*-Arten einen hohen Grad von Wirtsspezifität; nur die eine *Exotela* und die zwei *Dacnusa*-Arten greifen auch andere Agromyziden-Gattungen an. Wiederum wurde eine komplette Wirt-Parasiten-Liste zusammengestellt, die gleichzeitig eine Revision früherer Berichte für Europa berücksichtigt.

4. Vierzehn neue Arten werden beschrieben, eine Dapsilarthra-, eine Laotris- und zwölf Chorebus-Spezies.

### Резюме

1. Шестая часть настоящей серии занимается с европейскими паразитами Alysiini рода Cerodontha Rondani в границах, которых предлагал Nowakowski (1962). Паразиты принадлежат четырмя родам Daenusini (Laotris, Exotela, Daenusa, Daenusa и Chorebus) и одному роду вне Daenusini Dapsilarthra.

2. Для различных групп европейских видов *Chorebus*, включая группу *affinis*, (которая в этой статье впервые обграничивается) и для очень трудного комплекса *ovalis/lateralis* даются ревидированные определительные таблицы. Для паразитов определённых групп даются тоже таблицы для различения, чтобы дать возможность для детерминации выращенного материала.

3. От паразитов Dacnusini этой статьи показывают все виды Laotris и Chorebus высокий уровень специфики к хозяевам; только один вид Exotela и два вида Dacnusa нападают и на другие рода Agromyzidae. Снова составляется полный список хозяин-паразит который включает ревизии раньших данных для Европы.

4. Описывается четыренадцать видов, один у Dapsilarthra, один у Laotris и двенадцать у Chorebus.

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# Table 27

Biometric Data

		1								Abso	lute	Meas	suren	aents	3 (1	= 0.	01 m	m.)								
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25
			Head	l	th (lateral)	Eyes		es Width		ntenr gmei		Max	tillar Segi			r	hora	x	Hi	nd I	eg			d Ta gmer		
		Width	Length	Height	Eye-width	Distance between	Clypeus	Mandibles	3	4	5	3	4	5	6	Length	Width	Height	Femur	Tibia -	Tarsus	1	2	3	4	5
1 2	် ှ	60 62	35 34	48 46	17 17	28 28	22 21	10 9	14 13	12 12	11 11	9	11 11	8	10 9	91 89	45 45	59 58	50 53	78 75	71 69	26 26	14 14	10 10	7	11 10
3	ç	61	34	50	17	27	21	10	13	12	11	9	12	9	10	89	46	61	58	78	71	30	13	9	7	10

Nos. 1-2. Laotris striatula (HALIDAY) ex Cerodontha (Dizygomyza) luctuosa MEIGEN, Stuttgart, Germany.

# Table 28 Biometric Data

									A	bsol	ute 1	Measu	ırem	ents	(1 =	= 0.0	l mn	n.)								
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25
			Head	1	th (lateral)	e between	Width	les Width		nteni egme		Maz	tillar Segn	yPa hents	lpus s	т	hora	x	Hi	nd L	eg			nd Ta gmer		
		Width	Length	Height	Eye-width	Distance Eyes	Clypeus	Mandibles	3	4	5	3	4	5	6	Length	Width	Height	Femur	Tibia	Tarsus	1	2	3	4	5
$\frac{1}{2}$	<b>ở</b> ♀	67 69	39 33	51 50	20 18	36 34	22 23	12 10	17 17	15 14	14 13	13 11	17 15	$13 \\ 13$	16 16	111 100		76 74	69 69	95 96	85 87	34 34	17 17	12 14	9 9	$12 \\ 12$
3	Ŷ	57	32	43	15	28	21	11	16	13	12	9	12	10	11	85	41	58	61	83	74	31	15	11	7	10
4 5	<b>ð</b> ♀	69 56	37 31	50 43	$19 \\ 16$	39 30	24 22	11 8	19 16	15 13	$\frac{14}{12}$	12 10	17 14	13 10	$\frac{16}{12}$	104 77	50 41	74 56	74 61	99 78	85 74	34 30	19 14	12 11	9 9	11 10

Nos. 1–2. Chorebus talaris (HALIDAY): 1, ex Cerodontha (Poemyza) incisa MEIGEN on Festuca gigantea, Poland; 2, ex C. (P.) pygmaea MEIGEN on Brachypodium, Ireland.

238 238 273	Wing L	ength	50
109 104 95	Gaster I	Length	51
23 22 21	Width	Pet	70 TO 00
32 36	Length	iole	10
238 224 209	Total B	ody Length	1
238 224 209 1.8	Length/ Width o	f Head	Þ
1.4	Length/ Height o	of Head	Ŀ
2.2:1:0.8 2.2:1:0.7 2.3:1:0.8	Eyes/	f Head/ e between f Clypeus	¢
3.6 3.4	Width ( Mandib	of /Length les/of Head	2
1.1:1:0.9 1.1:1:0.9 1.1:1:0.9	8 4 5	Antennal Segments	12
1.5.5	Height/ Thorax	Length of	÷
1.3 1.4 1.3	Thorax Width /	/Head Width	4
$1.0 \\ 0.9 \\ 0.8$	Wing Length/	Body Length	F
0.9 0.9 0.9	Hind Ti	bia/Tarsus	۲
1.9:1:0.7:0.5:0.8 1.9:1:0.7:0.5:0.8 2.3:1:0.7:0.6:0.8		Hind Tarsal Segments	c
2.5 2.9	Gaster	of Petiole/	Þ
1.6 1.7 1.5	Width/I Petiole	length of	۲

aotris rupestris sp. nov., holotype.

262	310	271	319 295	 Wing L	ength	28	
108	143	104	148 118	 Gaster	Length	29	
19	24	23	27 28	Width	Peti	30	
32	46	36	39 41	Length	ole	31	
219	281	226	291 248		Body Length	32	
1.8	1.9	1.8	$1.7 \\ 2.1$		of Head	A	
1.4	1.4	1.4	1.5	Length Height	of Head	ß	
1,9:1:0.7	1.7:1:0.6	2.0:1:0.7	1.9:1:0.6 2.1:1:0.7	Distanc Eyes/	of Head/ ee between of Clypeus	C	
3.8	3.3	2.8	రు లు లు లు	Width o Mandib		P	
1.3:1:1.0	1.2:1:0.9	1.2:1:0.9	1.1:1:0.9 1.2:1:0.9	20 14 27	Antennal Segments	a	
1.4	1.4	1.5	1.5	Height/ Thorax	Length of	埬	Ra
1.4   1.4   0.8	1.4	1.4	1.2 1.3	Thorax Width		Ģ	Ratios
0.8	0.9	0.8	0.9	Wing Length/	/Body Length	Ħ	
0.9	0.9	0.9	0.9	Hind T	ibia/Tarsus	I	
2.1:1:0.8:0.6:0.7	1.9:1:0.7:0.5:0.6	$2.0\!:\!1\!:\!0.8\!:\!0.5\!:\!0.7$	1.9:1:0.7:0.5:0.7 1.9:1:0.8:0.5:0.7	1 2 3 4	Hind Tarsal Segments	ų	
		2	1 2 2	 on Lengths	of Petiole/		
3.4	3.1	2.9	9.9	 Gaster		X	
1.6	1.9	1.6	1.4 1.5	Width/L Petiole	ength of	Ч	

DOI: 10.21248/contrib.entomol.18.1-2.63-152

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iorebus subasper sp. nov., holotype. 5. Chorebus asperrimus sp. nov., Tatry, Poland (4 the holotype).

										Abs	olute	Mea	sure	ment	s (1	= 0.	01 m	m.)								
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25
		I	Iead		Eye-width (lateral)	ie a Ey	Clypeus Width	les Width		ntenı gmei				ryP: nent	alpus s		hora	ıx	Hi	nd I	eg			ıd Ta gmer		
		Width	Length	Height	Eye-wi	Distance between I	Clypeus	Mandibles	3	4	5	3	4	5	6	Length	Width	Height	Femur	Tibia	Tarsus	1	2	3	4	5
$\frac{1}{2}$	් ද	55 56	28 34	43 46	16 17	28 26	19 19	12 11	16 17	14 16	12 14	9 10	12 12	7 8	9 10	76 78	39 39	54 56	54 56	74 81	68 72	25 29	13 14	9 10	7 7	11 11
3	3	44	28	39	12	22	17	8	15	11	10	7	10	7	8	65	30	42	43	62	63	24	13	9	7	10
4	ę	40	30	40	15	16	15	7	13	11	10	6	9	7	8	63	28	41	41	63	65	26	14	9	7	9
5	ę	59	32	46	17	28	19	12	16	14	12	9	14	10	13	83	41	58	58	80	72	29	15	11	8	10
6 7	<b>*</b> o o ₽	54 58	32 32	41 43	20 17	26 27	19 19	11 14	14 15	12 14	11 13	- 7	12 12	9 9	10 11	68 72	38 41	56 54	50 52	64 72	59 61	22 23	12 12	9 8	77	9 10
8	5	52	28	39	16	27	19	11	14	12	11	7	11	8	10	67	35	52	49	65	61	22	13	9	7	10
9 10	<b>*o</b> o+	50 54	28 30	40 39	16 15	24 25	17 17	9 11	14 14	$13 \\ 13$	12 12	7 7	12 11	7 8	10 10	64 64	34 34	50 50	49 53	68 70	59 59	22 22	13 12	9 9	7 7	9 9
$\frac{11}{12}$	<b>*o</b> ♀	63 59	35 32	49 46	20 18	30 28	22 21	12 9	17 16	15 15	14 13	9 8	16 14	11 11	13 13	93 80	44 40	65 59	59 56	89 78	81 73	32 28	16 16	11 10	9 9	11 10
$\frac{13}{14}$	<b>*</b> o ♀	69 67	35 35	54 50	15 15	35 34	26 23	11 14	16 15	13 12	11 11	9 9	13 12	9 8	11 11	106 98	49 47	71 74	59 65	93 91	85 87	32 31	19 19	13 13	9 10	13 13
$\frac{15}{16}$	<b>*o</b> ♀	62 53	32 32	47 45	17 16	31 25	21 17	11 7	16 15	13 13	12 12	9 8	14 12	9 8	12 13	83 74	42 34	60 52	61 49	83 73	72 67	28 26	15 13	11 11	8 7	10 9
17	ę	55	34	43	18	25	21	10	16	12	12	9	11	8	11	81	37	58	53	77	69	27	15	10	7	9
18	ð ♀	56	30	41	14	29	19	11 12	15	13 13	12	10 9	13 12	9	11	78 72	41	59	55	75	66	26	13	10	7	9
19 2r	¥	56 58	31 32	43 44	16 17	26 26	$\begin{vmatrix} 21 \\ 19 \end{vmatrix}$	12 9	14   15	13	$\begin{vmatrix} 12 \\ 11 \end{vmatrix}$	9	12	9 8	11 15	72	39 40	58 59	$52 \\ 58$	74 81	68 71	25 26	15 16	10 11	7	10 11

Table 29 Biometric Data

Nos. 1-2. Chorebus flavipes (GOUREAU) ex Cerodontha (Dizygomyza) iraeos ROBINEAU-DESVOIDY; 1, Stuttgart, Germany; 2 Woodwalton, England.

No. 3. Chorebus crenulatus (THOMSON) ex Cerodontha (Cerodontha) denticornis PANZER, Poland.

No. 4. Chorebus diremtus (NRES) ex Cerodontha (Cerodontha) fulvipes MEIGEN, Poland.

No. 5. Chorebus rubicundus sp. nov., holotype.

Nos. 6-7. Chorebus enephes (NIXON) ex Cerodontha (Poemyza) deschampsiae SPENCER: 6, Mühlhausen, Germany; 7, Bieszczady, Poland.

No. 8. Chorebus ganesa (NIXON) ex Cerodontha (Poemyza) deschampsiae SPENCER, Germany.

Nos. 9-10. Chorebus ninella (NIXON) ex Cerodontha (Poemyza) calamagrostidis NOWAKOWSKI: 9, Röderbrunn, Germany; 10, Woodwalton, England.

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														Rat	ios									
26	27	28	29	30	31	32	A	в	C	D		Е		F	G	H	I			J			K	L
Hi Co		ength	length	Pe	tiole	Body Length	/ of Head	/ of Head	Width of Head/ Distance between Eyes/ Width of Clypeus	of /Length les/of Head		nten gme		Length of	/Head Width	/Body Length	Tibia/Tarsus			nd T egme		L	of Petiole/	ength of
Width	Length	Wing Length	Gaster Length	Width	Length	Total B	Length/ Width c	Length/ Height	Width o Distance Eyes/ Width o	Width of Mandibles/	3	4	5	Height/Length Thorax	Thorax Width /	Wing Length/	Hind Ti	1	2	3	4	5	Lengths Gaster	Ho
$16 \\ 14$	21 22	228 246	91 93	20 24	33 36	205 209		1.6 1.4	2.0:1:0.7 2.1:1:0.7	$\begin{array}{c} 2.4 \\ 3.0 \end{array}$		l:1: l:1:		1.4 1.4	1.4 1.4	0.9 0.9	0.9 0.9			.7:0 .7:0			$2.8 \\ 2.6$	1.7 1.5
10	19	190	111	17	28	200	1.6	1.5	2.1:1:0.8	3.3	1.8	3:1:	0.9	1.5	1.5	1.0	1.0	1.9	:1:0	.7:0	.5:0	.8	4.0	1.6
13	18	198	91	14	27	190	1.4	1.3	2.5:1:0.9	4.0	1.2	2:1:	0.9	1.5	1.4	1.0	1.0	1.9	:1:0	.7:0	.5:0	.6	3.3	2.0
15	24	238	109	20	35	214	1.9	1.5	2.1:1:0.7	2.6	1.1	l:1:	0.9	1.4	1.5	0.9	0.9	1.9	:1:0	.8:0	.6:0	7	3.1	1.8
$14 \\ 16$	21 23	205 228	$102 \\ 95$	$17 \\ 19$	32 34	$205 \\ 205$	1	$1.3 \\ 1.3$	2.1:1:0.7 2.1:1:0.7	$2.8 \\ 2.4$		2:1:		$1.2 \\ 1.3$	$1.4 \\ 1.4$	1.0 0.9	0.9 0.8	1		.8:0 .7:0			$3.2 \\ 2.8$	1.9 1.8
14	21	200	87	17	28	184	1.9	1.4	1.9:1:0.7	2.5		3:1:		1.3	1.5	0.9	0.9			.7:0			3.1	1.7
13 13	21 22	$205 \\ 214$	102 80	17 18	28 30	195 172		$1.4 \\ 1.3$	2.1:1:0.7 2.1:1:0.7	3.0 2.7		1:1: 2:1:		$1.3 \\ 1.3$	$1.5 \\ 1.6$	0.9 0.8	0.9 0.8	1.7	:1:0	.7:0 .7:0	.6:0	.7	3.7	1.7
17	26	281	148	28	41	276	1.8	1.4	2.1:1:0.7	3.0	1.2	8:1:	0.9	1.4	1.4	1.0	0.9	1.8	:1:0	.6:0	.5:0.	6	3.6	1.7
16	23	252	139	24	37	243	1.9	1.5	2.1:1:0.7	3.4	1.1	:1:	0.9	1.3	1.5	1.0	0.9	1.8	:1:0	.7:0	.6:0.	7	3.7	1.5
$16 \\ 16$	$\frac{28}{26}$	291 271	$\frac{115}{128}$	$\frac{28}{30}$	$\frac{37}{40}$	267 257	$\begin{array}{c} 1.9\\ 1.9\end{array}$	$1.5 \\ 1.4$	2.0:1:0.7 2.0:1:0.7	$3.2 \\ 2.6$		2:1:		$1.5 \\ 1.3$	1.4 1.4	0.9 0.9	0.9 1.0			.7:0			$3.1 \\ 3.2$	$1.3 \\ 1.3$
15	28	252	104	19	35	214	2.0	1.5	2.0:1:0.7	2.8	1.2	2:1:	0.9	1.4	1.5	0.8	0.9	1.9	:1:0	.8:0	.5:0.	7	2.9	1.9
13	22	214	92	18	30	195	1.7	1.4	2.1:1:0.7	4.2	1.1	:1:	0.9	1.4	1.6	0.9	0.9	2.0	:1:0	.8:0	.6:0.	7	3.1	1.7
17	26	238	87	17	32	200	1.6	1.3	2.2:1:0.8	3.4		3:1:		1.4	1.5	0.8	0.9	1.8	:1:0	.7:0	.5:0.	7	2.8	1.9
14 13	24	$\frac{252}{233}$	96 91	17	$\frac{32}{32}$	205	$1.9 \\ 1.8$	1.4	2.0:1:0.7 2.1:1:0.8	$2.7 \\ 2.5$		:1:		$1.3 \\ 1.8$	$1.4 \\ 1.4$	0.8	0.9 0.9			.8:0			3.0	1.9
13 14	24 24	$\frac{233}{248}$	91 87	19 19	32 32	$\frac{190}{209}$		1.4 1.4	2.1:1:0.8 2.2:1:0.7	$\frac{2.5}{3.5}$		2:1:		$1.3 \\ 1.3$		0.8 0.8				.7:0 .7:0			$\begin{array}{c} 2.9 \\ 2.8 \end{array}$	$1.7 \\ 1.6$

Nos. 11–12. Chorebus poemyzae sp. nov. ex Cerodontha (Poemyza) incisa MEIGEN, Poland : 11, on Agropyron repens, Piska Forest; 12, on Calamagrostis epigejos, Sieraków.

Nos. 13-14. Chorebus vitripennis sp. nov.: 13, Co. Down, Ireland; 14, holotype, Wales.

Nos. 15–16. Chorebus asramenes (NIXON) ex Cerodontha (Poemyza) pygmaea MEIGEN: 15, Oxshott, England; 16, Röderbrunn, Germany.

No. 17. Chorebus agraules (NIXON) ex Cerodontha (Poemyza) muscina MEIGEN, Germany.

Nos. 18-20. Chorebus difficilis sp. nov.: 18, ex Cerodontha (Poemyza) pygmaea MEIGEN, Stuttgart, Germany; 19, holotype ex C. (P.) pygmaea MEIGEN, England; 20, ex C. (Dizygomyza) chaixiana GROSCHKE, Germany.

Table 30 see p. 152.

Table 31 Biometric Data

										Abso	olute	Mea	sure	ment	s (1	= 0.0	)1 m	m.)								
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25
	:		Head	1	th (lateral)	Eyes		es Width		nteni egme		Ma	xilla Seg	ry Pa meni		r	'hora	x	н	ind I	eg			d Ta gme		
		idth	Length	Height	Eye-width	Distance between	Clypeus	Mandibles	3	4	5	3	4	5	6	Length	Width	Height	Femur	Tibia	Tarsus	1	2	3	4	5
1 2 3	<b>*o</b> o≠ o≠	61 59 56	34 35 34	50 46 46	17 17 17	30 24 22	21 20 20	10 9 9	21 16 17	14 13 14	13 12 13	12 9 9	16 10 11	9 9 9	11 10 12	98 87 87	45 39 40	66 56 58	58 51 54	93 79 78	92 78 78	37 34 32	19 15 16	15 10 11	10 7 8	13 10 11
4 5	ð ♀	52 53	31 32	41 43	17 19	24 19	18 17	8 7	18 19	13 15	13 14	8 10	14 15	9 9	11 13	80 80	36 36	54 54	$52 \\ 52$	78 76	78 72	34 32	15 14	11 10	8 7	11 10
6 7	<b>*o</b> ≎	63 68	37 43	54 61	15 21	28 28	$\frac{22}{24}$	11 13	18 21	14 14	13 13	9 9	$\frac{12}{13}$	9 10	9 10	93 104	44 48	67 72	59 63	87 88	83 84	34 34	16 17	$\frac{12}{13}$	9 9	$13 \\ 12$
8	ę	54	34	44	18	26	21	7	16	13	12	10	12	7	9	78	40	59	50	76	72	28	16	10	8	10
9 10	<b>°</b> ♀	46 46	26 28	37 39	14 14	24 22	17 19	7 7	13 14	11 13	11 11	6 7	8 9	6 7	8 9	72 73	$\frac{34}{36}$	44 46	41 46	67 69	69 71	28 29	13 14	9 9	7 7	10 12
11	ð	61	36	49	17	30	19	9	18	15	13	11	16	10	12	97	42	64	65	95	93	37	19	13	10	13
$12 \\ 13$	<b>°</b> ₽	48 51	28 28	39 40	13 14	24 22	21 21	9 10	15 14	14 13	12 11	7 7	9 9	7 6	8 8	71 71	37 39	46 49	46 49	71 74	76 71	30 29	14 13	11 9	8 7	$\frac{13}{12}$
$\begin{array}{c} 14 \\ 15 \end{array}$	ð ₽	50 44	32 30	41 39	13 15	24 21	21 	9 7	15 12	14 11	13 10	6 6	8 7	6 6	8 8	$\frac{79}{72}$	37 34	56 50	54 48	80 67	80 63	13 26	$\frac{16}{12}$	11 8	8 6	13 9
16	ð	60	35	50	16	28		10	21	15	13	9	13	9	11	106	46	74	74	106	95	41	18	31	10	14
17 18	<b>°</b> ♀	44 44	29 28	39 37	12 11	23 22	$\frac{20}{19}$	8 8	15 14	$13 \\ 12$	13 10	7 6	9 8	6 6	7 7	72 71	32 32	50 49	48 48	76 69	77 69	30 25	$\frac{16}{12}$	10 8	8 7	$\frac{12}{13}$

Nos. 1-3. Chorebus cytherea (NIXON), Germany: 1, ex Cerodontha (Dizygomyza) spinata GROSCHKE, Oberbayern; 2, ex C. (D.) luctuosa MEIGEN, Stuttgart; 3, ex C. (D.) chaixiana GROSCHKE, Stuttgart.

Nos. 4-5. Chorebus stilifer sp. nov., Germany.

Nos. 6-7. Chorebus iridis sp. nov., Italy (7 the holotype).

No. 8. Chorebus fordi (NIXON) ex Cerodontha (Poemyza) lateralis MACQUART, Poland.

Nos. 9-10. Chorebus siniffa (NIXON) ex Cerodontha (Dizygomyza) morosa MEIGEN, Poland.

'n

26	27	28	29	30	31	32	A	B	C	D		Е		F	G	H	I			J			ĸ	L
Hi Co		Length	Length	Pe	tiole	Total Body Length	1/ of Head	of Head	Width of Head/ Distance between Eyes/ Width of Clypeus	of /Length les/of Head		nten gme		Height/Length of Thorax	/Head Width	-	<u> </u>			id Ta gmei			of Petiole/	Width/Length of Petiole
Width	Length	Wing L	Gaster Length	Width	Length	Total B	Length/ Width e	Length/ Height of I	Width of Distanc Eyes/ Width of	Width of Mandibles/	3	4	5	Height/ Thorax	Thorax Width	Wing Length/	Hind Ti	1	2	3	4	5	Lengths Gaster	Width/J Petiole
18	29	291	135	19	42	286	1.8	1.5	2.1:1:0.7	3.4	1.4	:1:0	0.9	1.5	1.4	1.0	1.0	1.9	):1;	0.8:0	).5:0	.7	3.2	2.2
15	24	243	98	17	37	228	1.7	1.3	2.4:1:0.8	3.8	1.2	:1:6	).9	1.6	1.5	0.9	1.0	2.5	2:1:	0.7:0	0.5:0	.7	2.7	2.2
17	24	240	106	20	38	226	1.7	1.4	2.5:1:0.9	3.9	1.2	:1:	9.0	1.5	1.4	0.9	1.0	2.	0:1:	0.7:0	).5:0	).7	2.8	1.9
13	23	243	118	13	41	238	1.7	1.3	2.2:1:0.7	3.6	1.4	:1:	1.0	1.5	1.4	1.0	1.0	2.	2:1:	0.7:0	).6:0	).7	2.9	3.1
15	23	238	108	10	39	228	1.7	1.3	2.9:1:0.9	4.3	1.3	:1:	0.9	1.5	1.5	1.0	1.0	2.5	3:1:0	0.7:0	0.5:0	.7	2.8	3.7
17	24	252	122	22	43	262	1.7	1.5	2.3:1:0.8	3.3	1.3	:1:	0.9	1.4	1.4	1.0	1.0	2.	1:1:	0.8:0	0.6:0	.8	2.8	2.0
17	27	276	130	26	46	286	1.6	1.4	2.4:1:0.9	3.3	1.5	:1:	0.9	1.4	1.4	1.0	1.0	2.0	0:1:	0.8:0	).6:0	.7	2.8	1.8
15	24	243	96	17	35	216	1.6	1.3	$2.1\!:\!1\!:\!0.8$	4.9	1.2	:1:	0.9	1.3	1.3	0.9	1.0	1.	8:1:	0.7:0	).5:0	).7	2.7	2.0
12	21	222	106	13	31	209	11	1.4	1.9:1:0.7	3.5		:1:		1.6	1.4	0.9	1.0	2.	1:1:	0.7:0	0:6:0	.8	3.4	2.4
14	21	233	111	17	34	224	1.7	1.4	2.2:1:0.9	3.7	1.1	:1:	0.9	1.6	1.3	1.0	1.0	2.	1:1:	0.7:0	).5:0	.8	3.3	2.0
17	28	310	139	21	44	286	1.7	1.3	2.1:1:0.7	4.2	1.2	:1:	0.9	1.5	1.5	0.9	1.0	2.	0:1:	0.7:	0.5:0	0.7	3.2	2.1
11	21	248	99	19	32	200	1.7	1.4	2.0:1:0.8	3.0	1.1	:1:	0.9	1.5	1.3	0.8	1.1	2.1	:1:(	0.8:0	.6:0	.9	3.2	1.7
13	22	257	108	21	35	226	1.8	1.4	2.4:1:0.9	2.8	1.1	:1:	0.9	1.4	1.3	0.9	1.0	2.3	2:1:	0.7:0	0.6:0	.9	3.1	1.7
15	23	262	102	24	34	219	1.6	1.3	2.1:1:0.8	3.4	1.1	:1:	0.9	1.4	1.3	0.8	1.0	2.0	0:1:	0.7:0	).5:0	.8	3.1	1.4
13	21	228	83	21	26	186	1.5	1.3	2.2:1: -	4.0	1.1	:1:	0.9	1.4	1.3	0.8	0.9	2.5	2:1:	0.7:0	).5:(	).8	3.2	1.2
17	26	327	139	30	36	281	1.7	1.4	2.2:1: -	3.4	1.4	:1:	0.9	1.4	1.3	0.8	0.9	2,	3:1:	0.7:0	).6:0	.8	3.8	1.2
12	23	219	108	21	30	214	1.6	1.4	1.9:1:0.8	3.5	1.1	:1:	1.0	1.4	1.4	1.0	1.0	1.	9:1:	0.6:0	).5:0	.8	3.3	1.4
13	21	228	113	25	30	219	1.6	1.3	2.1:1:0.9	3.3	1.2	:1:	0.9	1.4	1.4	1.0	1.0	2.	1:1:	0.7:0	).6:1	1	3.8	1.2

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No. 11. Chorebus vernalis sp. nov., holotype.

Nos. 12-13. Chorebus metallicus sp. nov., Poland.

Nos. 14–15. Chorebus nigriscaposus (NIXON): 14, ex Cerodontha (Icteromyza) geniculata FALLÉN, Poland; 15, ex C. (I.) calosoma HENDEL, Denmark.

No. 16. Chorebus lanigerus (STELFOX) ex Cerodontha (Icteromyza) calosoma HENDEL, Denmark.

Nos. 17-18. Chorebus gracilipes (THOMSON): 17, lectotype, Sweden; 18, ex Cerodontha (Icteromyza) geniculata FALLÉN, Poland.

			Absolute Measurements $(1 = 0.01 \text{ mm.})$																							
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25
		Head			th (lateral)	Eyes	Width	es Width		aten gme		Maxillary Palpus Segments				Thorax			Hind Leg			Hind Tarsal Segments				
		Width		Height	Eye-width	Distance between	Clypeus	Mandibles	3	4	5	3	4	5	6	Length	Width	Height	Femur	Tibia	Tarsus	1	2	3	4	5
1 2	ð ç	51 65	28 33	39 48	13 18	25 30	19 22	9 11	14 17	10 14	10 13	9	11 15	6 9	9 10	65 76		50 67	43 59	61 75	51 63	21 25	9 11	8 9	6 7	9 11
3 4	đ ç	66 61	36 35	51 46	16 19	32 29	24	13 -	18 16	$13 \\ 13$	12 12	8	12	9 	10 —	86 83		69 62	64 59	86 80	71 64	28 24	13 12	10 9	7 7	12 11
5 6	<b>ð</b> ₽	69 56	39 31	54 44	21 16	32 24	23 21	10 9	21 18	15 15	13 14	11 11	15 14	11 9	13 12	96 85		74 59	63 54	95 78	93 77	37 31	19 15	14 11	10 8	13 11
7 8	<b>ð</b> ♀	78 73	41 39	60 59	22 21	40 37	24 24	12 11	24 22	16 17	14 15	14 14	19 19	13 13	15 15	117 117		81 78		109 109	1	44 44	21 21	14 15	10 10	16 15
9 10	°o ₽	65 64	36 35	51 50	17 19	31 28	$\begin{array}{c} 23\\ 21 \end{array}$	11 10	20 19	$\begin{array}{c} 16\\ 15\end{array}$	15 14	13 11	16 15	11 11	$\begin{array}{c} 13\\14 \end{array}$	96 95		67 65	65 66	95 96	90 88	38 36	17 17	$\begin{array}{c} 13\\12\end{array}$	9 9	12 14

Table 30 Biometric Data

Right part of table 30

							Ratios																	
26	27	28	29	30	31	32	A	в	C	D	Е			F	G	H	I		J					L
Hi: Co		Length	Gaster Length	Petiole		Body Length	/ of Head	/ of Head	of Head/ ce between of Clypeus	of /Length les/of Head	Segments			Length of	/Head Width	Width (Body Length	Tibia/Tarsus				d Tarsal gments		of Petiole/	Length of
Width	Length	Wing Le		Width	Width Length	Total B(	Length/ Width o	Length/ Hei <sub>s</sub> ht o	Width of Distance I Eyes/ Width of	Width of Mandibles	3	4	5	Height/ Thorax	Thorax Width	Wing Length/	Hind Ti	1	2	3	4	5	Lengths Gaster	Width/I Petiole
12	19	190	76	14	27	178	1.8	1.4	2.0:1:0.7	3.0	1.8	3:1:	1.0	1.3	1.5	0.9	0.8	2.5	2:1:	0.8:0	).6:0	.9	2.8	1.9
14	23	238	87	17	40	190	2.0	1.5	2.2:1:0.7	2.9	1.2	2:1:	0.9	1.1	1.4	0.8	0.8	2.2	2:1:0	0.8:0	0.6:0	.8	2.2	2.4
15	24	262	115	17	43	243	1.8	1.4	2.0:1:0.7	2.8	1.4	l:1:	0.9	1.2	1.4	0.9	0.8	2.1	:1:	0.8:0	0.6:0	.9	2.7	2.5
13	21	255	96	20	43	217	1.7	1.3	2.1:1: -	-	1.5	2:1:	0.9	1.3	1.3	0.8	0.8	2.1	:1:	0.8:0	0.6:0	.9	2.2	2.2
19	28	281	148	19	50	286	1.8	1.4	2.1:1:0.7	3.9	1.4	L:1:	0.9	1.3	1.5	1.0	1.0	2.0	):1:	0.7:0	).5:0	.7	3.0	2.6
16	22	257	102	15	39	238	1.8	1.5	2.3:1:0.8	3.5	1.2	2:1:	0.9	1.4	1.4	0.9	1.0	2.1	:1:	0.7:0	0.5:0	.7	2.6	2.6
21	35	300	143	25	59	316	1.9	1.5	2.0:1:0.6	3.4	1.5	5:1:	0.6	1.4	1.3	1.1	1.0	2.2	:1:0	0.7:0	).5:0	.8	2.4	2.4
19	32	300	130	26	54	295	1.9	1.5	2.0:1:0.6	3.5	1.8	3:1:	0.9	1.5	1.3	1.0	0.9	2.1	:1:	0.7:0	).5:0	.7	2.4	2.1
18	28	281	130	20	43	267	1.8	1.4	2.1:1:0.7	3.3	1.2	2:1:	0.9	1.4	1.4	1.0	0.9	2.2	:1:0	0.7:0	0.5:0	.7	3.0	2.2
18	31	276	126	21	43	271	1.8	1.4	2.3:1:0.8	3.4	1.5	3:1:	0.9	1.5	1.5	1.0	0.9	2.2	:1:0	0.7:0	0.6:0	.8	3.0	2.1

Nos. 1-2. Chorebus transversus (NIXON) ex Cerodontha (Butomomyza) angulata LOEW: 1, Stuttgart, Germany; 2, Bookham, England.

Nos. 3-4. Chorebus humeralis sp. nov.: 3, holotype, Ireland; 4, ex Cerodontha (Butomomyza) angulata LOEW, Denmark.

Nos. 5-6. Chorebus merella (NIXON), Germany: 5, ex Cerodontha (Butomomyza) scirpi KARL, Stuttgart; 6, ex C. (Dizygomyza) caricicola HERING, Oberbayern.

Nos. 7-8. Chorebus nobilis sp. nov., Hessen, Germany.

Nos. 9-10. Chorebus pelion (NIXON), Hessen, Germany.